

**NPIP 44th NPIP Biennial Conference
Franklin, TN -- June 28, 2018 Voting Results**

Report of Voting Results on 9-CFR Proposed Changes

9 CFR Proposal Number	Committee Referred To	Committee Recommendations: Passed, Passed as Amended by Technical Committee, Passed as Amended by Break-Out Group, Passed as Amended by Coordinating Committee, Passed as Amended on Floor, Died, Died for Lack of Motion, Died for Lack of Second
1.	Combined	Passed as Amended by Coordinating Committee
2.	Combined	Passed as Amended by Coordinating Committee
3.	145 Combined	Passed
4.	145 Combined	Died for Lack of Motion
5.	145 Combined	Died for Lack of Motion
6.	145 Combined	Passed as Amended by Coordinating Committee
7.	Combined	Passed as Amended by Coordinating Committee
8.	Combined	Passed as Amended by Coordinating Committee
9.	145 B	Passed as Amended by Breakout-Group
10.	145 C	Died for Lack of Motion
11.	145 C	Passed
12.	145 C	Passed
13.	145 C	Died for Lack of Motion
14.	145 D	Passed as Amended by Technical Committee
15.	145 E and 146 E	Passed as Amended by Break-Out Group
16.	145 E	Died for Lack of Motion
17.	145 E	Passed as Amended by Break-Out Group
18.	145 G	Passed
19.	145 G	Passed as Amended by Technical Committee
20.	145 H	Passed
21.	145 H	Passed as Amended by Technical Committee
22.	146 B	Died for Lack of Motion
23.	Combined	Passed as Amended by Coordinating Committee
24.	Combined	Passed
LPC 1.	Combined	Passed as Amended by Coordinating Committee
LPC 2.	Combined	Passed

NPIP 44th NPIP Biennial Conference
Franklin, TN -- June 28, 2018 Voting Results

Report of Voting Results on Program Standards

Program Standard Number	Committee Referred To	Committee Recommendations: Passed, Passed as Amended by Technical Committee, Passed as Recommended by Technical Committee, Passed as Amended by Break-Out Group, Passed as Amended by Coordinating Committee, Passed as Amended on Floor, Died, Died for Lack of Motion, Died for Lack of Second
1.	Combined	Passed as Amended by Technical Committee
2.	Combined	Passed as Amended by Coordinating Committee
3.	Combined	Passed as Amended by Coordinating Committee
4.	Combined	Passed as Amended by Coordinating Committee
5.	Combined	Passed
6.	Combined	Passed
7.	Combined	Passed as Amended by Coordinating Committee
8.	Combined	Passed as Amended by Coordinating Committee
9.	Combined	Passed as Amended by Coordinating Committee
10.	Combined	Passed as Amended on Floor
11.	Combined	Passed as Recommended by Technical Committee
12.	Combined	Passed as Amended on Floor
13.	145 D, G, H	Passed as Amended by Technical Committee
14.	145 D, G, H	Passed as Amended by Technical Committee
15.	145 D, G, H	Passed as Amended by Technical Committee
16.	145 D, G, H	Passed as Amended by Technical Committee

Proposal No. 1 (Amended by TC)

Delegates: Combined

§56.1 Definitions.

~~H5/H7 LPAI exposed.~~ At risk of developing H5/H7 LPAI because of association with birds or poultry infected with H5/H7 LPAI, excrement from birds or poultry infected with H5/H7 LPAI, or other material touched by birds or poultry infected with H5/H7 LPAI, or because there is reason to believe that association has occurred with H5/H7 LPAI or vectors of H5/H7 LPAI, as determined by the Cooperating State Agency and confirmed by APHIS.

H5/H7 LPAI virus exposed (non-infectious).

- (1) Poultry will be considered to be exposed (not infectious) to H5/H7 LPAI ~~V~~ for the purposes of this part if:
 - (i) ~~Antibodies to the H5 or H7 subtype of the AI virus that are not a consequence of vaccination have been detected in poultry; if vaccine is used, methods should be used to distinguish vaccinated birds from birds that are both vaccinated and infected; and~~
 - (ii) ~~Samples collected from the flock using real-time RT-PCR or virus isolation an agent detection test approved by the Department and the Official State Agency are determined to be not infectious for H5/H7 LPAI.~~
- (2) ~~The official determination that H5/H7 LPAI virus exposure has occurred is by the identification of antibodies to the H5 or H7 subtype of AI virus detected and may only be made by the National Veterinary Services Laboratories.~~

H5/H7 LPAI virus, actively infected (infectious).

- (1) Poultry will be considered to be actively infected with H5/H7 LPAI ~~V~~ for the purposes of this part if:
 - (i) H5/H7 LPAI virus has been isolated and identified as such from poultry,; or
 - (ii) Viral antigen or viral RNA specific to the H5 or H7 subtype of AI virus has been detected in poultry. ; or
 - (iii) ~~Antibodies to the H5 or H7 subtype of the AI virus that are not a consequence of vaccination have been detected in poultry. If vaccine is used, methods should be used to distinguish vaccinated birds from birds that are both vaccinated and infected. In the case of isolated serological positive results, H5/H7 LPAI infection may be ruled out on the basis of a thorough epidemiological investigation that does not demonstrate further evidence of H5/H7 LPAI infection, as determined by the Cooperating State Agency, the Official State Agency, and APHIS.~~
- (2) ~~The official determination that H5/H7 LPAI virus has been isolated and identified, or viral antigen or viral RNA specific to the H5 or H7 subtype of AI virus has been detected, or antibodies to the H5 or H7 subtype of the AI virus have been detected may only be made by the National Veterinary Services Laboratories.~~

§145.1 Definitions.

H5/H7 LPAI virus exposed (non-infectious)

- (1) Poultry will be considered to be exposed (not infectious) to H5/H7 LPAI ~~V~~ for the purposes of this part if:

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- (i) Antibodies to the H5 or H7 subtype of the AI virus that are not a consequence of vaccination have been detected in poultry; ~~if vaccine is used, methods should be used to distinguish vaccinated birds from birds that are both vaccinated and infected;~~ and
 - (ii) Samples collected from the flock using ~~real-time RT-PCR or virus isolation an agent detection test approved by the Department and the Official State Agency~~ are determined to be not infectious for H5/H7 LPAI.
- (2) The official determination that H5/H7 LPAI virus exposure has occurred is by the identification of antibodies to the H5 or H7 subtype of AI virus detected and may only be made by the National Veterinary Services Laboratories.

H5/H7 LPAI virus, actively infected (infectious)

- (1) Poultry will be considered to be infected with H5/H7 LPAI for the purposes of this part if:
- (i) H5/H7 LPAI virus has been isolated and identified as such from poultry, or
 - (ii) Viral antigen or viral RNA specific to the H5 or H7 subtype of AI virus has been detected in poultry.
- (2) The official determination that H5/H7 LPAI virus has been isolated and identified, or viral antigen or viral RNA specific to the H5 or H7 subtype of AI virus has been detected, may only be made by the National Veterinary Services Laboratories.

§146.1 Definitions.

H5/H7 LPAI virus exposed (non-infectious).

- (1) Poultry will be considered to be exposed (not infectious) to H5/H7 LPAIV for the purposes of this part if:
- (i) Antibodies to the H5 or H7 subtype of the AI virus that are not a consequence of vaccination have been detected in poultry; ~~if vaccine is used, methods should be used to distinguish vaccinated birds from birds that are both vaccinated and infected;~~ and
 - (ii) Samples collected from the flock using ~~real-time RT-PCR or virus isolation an agent detection test approved by the Department and the Official State Agency~~ are determined to be not infectious for H5/H7 LPAI.
- (2) The official determination that H5/H7 LPAI virus exposure has occurred is by the identification of antibodies to the H5 or H7 subtype of AI virus detected and may only be made by the National Veterinary Services Laboratories.

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H5/H7 LPAI virus, actively infected (infectious).

- (1) Poultry will be considered to be actively infected with H5/H7 LPAIV for the purposes of this part if:
- (i) H5/H7 LPAI virus has been isolated and identified as such from poultry,; or
 - (ii) Viral antigen or viral RNA specific to the H5 or H7 subtype of AI virus has been detected in poultry. ;or
 - (iii) ~~Antibodies to the H5 or H7 subtype of the AI virus that are not a consequence of vaccination have been detected in poultry. If vaccine is used, methods should be used to distinguish vaccinated birds from birds that are both vaccinated and infected. In the case of isolated serological positive results, H5/H7 LPAI infection may be ruled out on the basis of a thorough epidemiological investigation that does not demonstrate further~~

evidence of H5/H7 LPAI infection, as determined by the Cooperating State Agency, the Official State Agency, and APHIS.

- (2) The official determination that H5/H7 LPAI virus has been isolated and identified, or viral antigen or viral RNA specific to the H5 or H7 subtype of AI virus has been detected, or antibodies to the H5 or H7 subtype of the AI virus have been detected may only be made by the National Veterinary Services Laboratories.

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Definitions

H5/H7 LPAI virus exposed (non-infectious).

- (1) Poultry will be considered to be exposed (not infectious) to H5/H7 LPAI V for the purposes of this part if:
- (i) Antibodies to the H5 or H7 subtype of the AI virus that are not a consequence of vaccination have been detected in poultry; if vaccine is used, methods should be used to distinguish vaccinated birds from birds that are both vaccinated and infected; and
 - (ii) Samples collected from the flock using real-time RT-PCR or virus isolation ~~an agent detection test approved by the Department and the Official State Agency~~ are determined to be not infectious for H5/H7 LPAI.
- (2) The official determination that H5/H7 LPAI virus exposure has occurred is by the identification of antibodies to the H5 or H7 subtype of AI virus detected and may only be made by the National Veterinary Services Laboratories.

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H5/H7 LPAI virus, actively infected (infected).

- (1) Poultry will be considered to be actively infected with H5/H7 LPAI V for the purposes of this part if:
- (i) H5/H7 LPAI virus has been isolated and identified as such from poultry,; or
 - (ii) Viral antigen or viral RNA specific to the H5 or H7 subtype of AI virus has been detected in poultry. ~~;~~
 - (iii) Antibodies to the H5 or H7 subtype of the AI virus that are not a consequence of vaccination have been detected in poultry. If vaccine is used, methods should be used to distinguish vaccinated birds from birds that are both vaccinated and infected. In the case of isolated serological positive results, H5/H7 LPAI infection may be ruled out on the basis of a thorough epidemiological investigation that does not demonstrate further evidence of H5/H7 LPAI infection, as determined by the Cooperating State Agency, the Official State Agency, and APHIS.
- (2) The official determination that H5/H7 LPAI virus has been isolated and identified, or viral antigen or viral RNA specific to the H5 or H7 subtype of AI virus has been detected, or antibodies to the H5 or H7 subtype of the AI virus have been detected may only be made by the National Veterinary Services Laboratories.

Reason:

This proposed change adds the definition of H5/H7 LPAI virus infection (infected) and H5/H7 LPAI virus (exposed) to Part 145, Subpart A definitions. The addition of these definitions provides consistency to Parts 145, 146, 56 and Program Standards.

This proposed change amends the definition of H5/H7 LPAI virus infection (infected) and adds the definition of H5/H7 LPAI virus (exposed) to Part 146, Subpart A and Program Standards definitions. The amendment and addition of these definitions provides consistency to Parts 145, 146, 56 and Program Standards.

This proposed change amends the definition of H5/H7 LPAI virus infection (infected) and H5/H7 LPAI virus (exposed) to Part 56.1 definitions. Amending these definitions provides consistency with Parts 145, 146, 56 and Program Standards.

The consistency of H5/H7 LPAI virus (exposed) and H5/H7 LPAI virus infection (infected) definitions define the H5/H7 LPAI transmission risks when test results are reported from an NPIP Authorized Laboratory, NAHLN Laboratory or NVSL. Interpretation of these results by the Department, Official State Agency and Cooperating State Agency provides timely information for the H5/H7 LPAI response as outlined the State's Initial State Response and Containment Plan (ISRCP).

Sponsors: Minnesota H5/H7 LPAI Emergency Disease Management Committee (EDMC)

Proposal No. 2

Delegates: Combined

§56.3 Payment of indemnity.

(b) Percentage of costs eligible for indemnity.

Except for poultry that are described by the categories in paragraphs (b)(1) through (b)(3) of this section, the Administrator is authorized to pay 100 percent of the costs, as determined in accordance with §56.4, of the activities described in paragraphs (a)(1) through (a)(3) of this section, regardless of whether the infected or exposed poultry participate in the Plan. For infected or exposed poultry that are described by the categories in paragraphs (b)(1) through (b)(3) of this section, the Administrator is authorized to pay 25 percent of the costs of the activities described in paragraphs (a)(1) through (a)(3) of this section:

(1) The poultry are from a breeding flock, ~~commercial flock or slaughter plant~~ that participates in any Plan program in part 145 ~~or part 146~~ of this chapter but that does not participate in the U.S. Avian Influenza Clean, ~~or the U.S. H5/H7 Avian Influenza Clean,~~ or U.S. H5/H7 Avian Influenza Monitored program of the Plan available to the flock in part 145 ~~or part 146~~ of this chapter; ~~or and~~

~~(2) The poultry are from a commercial flock or slaughter plant, but the flock or slaughter plant does not participate in the U.S. Avian Influenza Monitored program available to the commercial flock or slaughter plant in part 146 of this chapter; or~~

~~(2) The poultry are from:~~

~~i. A commercial table-egg laying premises with at least 75,000 birds;~~

~~ii. A meat-type chicken slaughter plant that slaughters at least 200,000 meat-type chickens in an operating week;~~

~~iii. A meat-type turkey slaughter plant that slaughters at least 2 million meat-type turkeys in a 12 month period;~~

~~iv. A commercial waterfowl and commercial upland game bird slaughter plant that slaughters at least 50,000 birds annually;~~

~~v. A raised-for-release upland game bird premises, raised-for-release waterfowl premises, and commercial upland game bird or commercial waterfowl producing eggs for human consumption premises that raise at least 25,000 birds annually;~~

~~vi. A breeder flock premises with at least 5,000 birds.~~

(3) The poultry are located in a State that does not participate in the diagnostic surveillance program for H5/H7 LPAI, as described in §146.14 of this chapter, or that does not have an initial State response and containment plan for H5/H7 LPAI that is approved by APHIS under §56.10, unless such poultry participate in the Plan with another State that does participate in the diagnostic surveillance program for H5/H7 LPAI, as described in §146.14 of this chapter, and has an initial State response and containment plan for H5/H7 LPAI that is approved by APHIS under §56.10.

Deleted: or

Reason: Adding the size requirements in this section provides clarity and additional reference for indemnity eligibility. The exemption numbers are already listed in Part 146 Subparts B-E, this proposed change provides an exemption number for breeders participating in Part 145.

Sponsor: Dr. Shauna Voss, Minnesota Board of Animal Health
Dr. Dale Lauer, Minnesota Board of Animal Health
Dr. Michael Kopp, Indiana Board of Animal Health
Mr. Paul Wm. Brennan, Indiana State Poultry Association

Proposal No. 3

Delegates: 145 Combined

§145.7 Specific provisions for participating dealers.

Dealers in ~~poultry breeding stock~~, hatching eggs, ~~or baby~~ newly-hatched poultry or started poultry shall comply with the all provisions in this part Subpart A and Program Standards that which apply to their operations.

Reason: This proposal closely aligns and defines the types of poultry that dealers are associated with as defined in 145.1. It will also require dealers to act in accordance with plan provisions and applicable sanitation details listed in Program Standards.

Sponsor: Dr. Dale Lauer, Minnesota Board of Animal Health, Willmar, MN

Proposal No. 4

Delegates: 145 Combined

§145.7 Specific provisions for participating dealers.

(a) Dealers in poultry breeding stock, hatching eggs, or baby or started poultry shall comply with all provisions in ~~this part~~ Subpart A and the Program Standards that which apply to their operations.

(b) Dealers shall obtain, maintain and comply with licensure and importation requirements for all states where sales are conducted and where products are delivered.

(c) Dealers shall provide to each purchaser a VS Form 9-3 that correctly describes the number and the type of breeding stock, hatching eggs, or baby or started poultry at the time of shipment; the name, physical address and phone number of the purchaser, and the name, physical address and phone number of the dealer. Each VS Form 9-3 shall contain the Report Number of the original hatchery issued VS Form 9-3 listed in "Section 10. Remarks" and also be entered into the dealer's shipping and inventory records. All completed NPIP forms must be returned to the Official State Agency (OSA) within 7 days. The OSAs of the states where business is conducted may also require a weekly sales report submitted by email or fax.

(d) Dealers shall have a biosecurity plan that addresses all aspects of the business including but not limited to the poultry, housing, feed, water, equipment, vehicles and personnel.

Reason: The current language in §145.7 is very generalized and needs more details to address current issues seen with compliance matters with dealers moving poultry and eggs across multiple state lines i.e. incomplete or inaccurate records, non-compliance with state importation statutes, etc.

Sponsor: Dr. Mary Jane Lis
Connecticut Department of Agriculture

Proposal No. 5

Delegates: 145 Combined

§145.12 Inspections.

(a) Each ~~participating~~ Plan participant hatchery shall be audited at least one time annually or a sufficient number of times each year to satisfy the Official State Agency that the operations ~~of the hatchery~~ are in compliance with the provisions of the Plan and Program Standards.

(b) ~~The Records of all flocks maintained primarily~~ for production and distribution of hatching eggs and other products shall be made available to and examined annually by a State Inspector. Records shall include but are not limited to VS Form 9-2, "Flock Selecting and Testing Report"; VS Form 9-3, "Report of Sales of Hatching Eggs, Chicks, and Poults"; set and hatch records; egg receipts; and egg/chick orders or invoices. Records shall be maintained for 3 years. On-site inspections of flocks and premises will be conducted if the State Inspector determines that a breach of sanitation, blood testing, or other provisions has occurred for Plan programs for which the flocks have or are being qualified.

Reason: The current plan provisions only require hatcheries be audited annually, there are no auditing requirements for breeding flocks or dealers. To satisfy Official State Agencies that participants comply with plan provisions and Program Standards, this proposal will make certain adequate oversight of all plan participants is conducted at a minimum annually.

Sponsor: Dr. Shauna Voss, Minnesota Board of Animal Health, Willmar, MN
Dr. Dale Lauer, Minnesota Board of Animal Health, Willmar, MN

Proposal No. 6 (Amended by TC)

Delegates: 145 Combined

§145.14 Testing.

(e) *For Newcastle Disease, as defined by OIE-Virus*. The official tests for ND~~V~~ are ~~the hemagglutination inhibition (HI) test, the enzyme-linked immunosorbent assay (ELISA) test, a serological test for antibody detection~~ or a molecular-based test ~~for antigen detection~~.

Reason: The Primary Breeders propose the addition of an ND Clean program. See corresponding proposal in Program Standards Subpart F.

Sponsor: Primary Breeder Association
Dr. Elena Behnke, Aviagen
Dr. Alberto Torres, Cobb-Vantress
Dr. Travis Schaal, Hy-Line
Dr. Dustin Burch, Aviagen Turkeys

Proposal No. 7

Delegates: Combined

§145.14 Testing.

(d) For Avian Influenza.

The official tests for avian influenza are described in paragraphs (d)(1) and (d)(2) of this section:

(1) Antibody detection tests-

(i) Enzyme-linked immunosorbent assay (ELISA) test.

(A) The ELISA test must be conducted using test kits approved by the Department and the Official State Agency and must be conducted in accordance with the recommendations of the producer or manufacturer.

(B) When positive ELISA samples are identified, an AGID test must be conducted within 48 hours.

(ii) ~~The a~~Agar gel immunodiffusion (AGID) test.

~~(A) The AGID test must be conducted on all ELISA positive samples.~~

(B) The AGID test must be conducted using reagents approved by the Department and the Official State Agency.

(C) The AGID test for avian influenza must be conducted in accordance with part 147 of this subchapter Program Standard A – Blood Testing Procedures (8) Standard test procedures for avian influenza (a) agar gel immunodiffusion (AGID) test. The test can be conducted on egg yolk or blood samples. The AGID test is not recommended for use in waterfowl.

(D) Positive tests for the AGID must be further tested by Federal Reference Laboratories using appropriate tests for confirmation. Final judgment may be based upon further sampling and appropriate tests for confirmation.

§146.13 Testing.

(b) Avian Influenza.

The official tests for avian influenza are described in paragraphs (b)(1) and (b)(2) of this section:

(1) Antibody detection tests-

(i) Enzyme-linked immunosorbent assay (ELISA) test.

(A) The ELISA test must be conducted using test kits approved by the Department and the Official State Agency and must be conducted in accordance with the recommendations of the producer or manufacturer.

(B) When positive ELISA samples are identified, an AGID test must be conducted within 48 hours.

(ii) ~~The a~~Agar gel immunodiffusion (AGID) test.

~~(A) The AGID test must be conducted on all ELISA positive samples.~~

(B) The AGID test must be conducted using reagents approved by the Department and the Official State Agency.

(C) The AGID test for avian influenza must be conducted in accordance with part 147 of this subchapter Program Standard A – Blood Testing Procedures (8) Standard test procedures for avian influenza (a) agar gel immunodiffusion (AGID) test. The test can be conducted on egg yolk or blood samples. The AGID test is not recommended for use in waterfowl.

(D) Positive tests for the AGID must be further tested by Federal Reference Laboratories using appropriate tests for confirmation. Final judgment may be based upon further sampling and appropriate tests for confirmation.

Reason: When samples are submitted from flocks as part of a pre-movement testing program, positive ELISA samples may not be tested via the AGID test in a timely manner to allow the flock to move. This proposal will require timely testing and reporting of AGID test results when ELISA-positive samples are identified at an NPIP Authorized Laboratory.

Sponsor: Minnesota H5/H7 LPAI Emergency Disease Management Committee (EDMC)

Proposal No. 8

Delegates: Combined

§145.14 Testing.

(d) For avian influenza.

The official tests for avian influenza are described in paragraphs (d)(1) and (d)(2) of this section.

- (1) Antibody detection tests
 - (i) Enzyme-linked immunosorbent assay (ELISA). ELISA must be conducted using test kits approved by the Department and the Official State Agency and must be conducted in accordance with the recommendations of the producer or manufacturer.
 - (ii) The agar gel immunodiffusion (AGID) test.
 - (A) The AGID test must be conducted on all ELISA-positive samples.
 - (B) The AGID test must be conducted using reagents approved by the Department and the Official State Agency.
 - (C) The AGID test for avian influenza must be conducted in accordance with part 147 of this subchapter. The test can be conducted on egg yolk or blood samples. The AGID test is not recommended for use in waterfowl.
 - (D) Positive tests for the AGID must be further tested by Federal Reference Laboratories using appropriate tests for confirmation. Final judgment may be based upon further sampling and appropriate tests for confirmation.
- (2) Agent detection tests. Agent detection tests may be used to detect influenza A ~~virus matrix gene or protein~~ virus but not to determine hemagglutinin or neuraminidase subtypes. Samples for agent detection testing should be collected from naturally occurring flock mortality or clinically ill birds.

§146.13 Testing.

(b) Avian influenza.

The official tests for avian influenza are described in paragraphs (b)(1) and (b)(2) of this section.

- (1) Antibody detection tests-
 - (i) Enzyme-linked immunosorbent assay (ELISA). ELISA must be conducted using test kits approved by the Department and the Official State Agency and must be conducted in accordance with the recommendations of the producer or manufacturer.
 - (ii) The agar gel immunodiffusion (AGID) test.
 - (A) The AGID test must be conducted on all ELISA-positive samples.
 - (B) The AGID test must be conducted using reagents approved by the Department and the Official State Agency.
 - (C) The AGID test for avian influenza must be conducted in accordance with part 147 of this subchapter. The test can be conducted on egg yolk or blood samples. The AGID test is not recommended for use in waterfowl.
 - (D) Positive tests for the AGID must be further tested by Federal Reference Laboratories using appropriate tests for confirmation. Final judgment may be based upon further sampling and appropriate tests for confirmation.

- (2) Agent detection tests. Agent detection tests may be used to detect influenza A ~~virus matrix gene or protein~~ but not to determine hemagglutinin or neuraminidase subtypes. Samples for agent detection testing should be collected from naturally occurring flock mortality or clinically ill birds.

Reason: This imposes an unnecessary technical restriction on test design and precludes the use of lateral flow antigen immunoassays which target the NP protein.

Sponsor: Dr. Erica Spackman
USDA-Agricultural Research Service

Proposal No. 9

Delegates: 145 B

§145.23 Terminology and classification; flocks and products.

(d) U.S. S. Enteritidis Clean.

This classification is intended for egg-type breeders wishing to assure their customers that the hatching eggs and chicks produced are certified free of Salmonella enteritidis.

- (1) A flock and the hatching eggs and chicks produced from it which have met the following requirements as determined by the Official State Agency:
 - (i) The flock originated from a U.S. S. enteritidis Clean flock, or meconium from the chick boxes and a sample of chicks that died within 7 days after hatching are examined bacteriologically for salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped.
 - (ii) All feed fed to the flock shall meet the following requirements:
 - (A) Pelletized feed shall contain either no animal protein or only animal protein products produced under the Animal Protein Products Industry (APPI) Salmonella Education/Reduction Program. The protein products must have a minimum moisture content of 14.5 percent and must have been heated throughout to a minimum temperature of 190 °F., or above, or to a minimum temperature of 165 °F. for at least 20 minutes, or to a minimum temperature of 184 °F. under 70 lbs. pressure during the manufacturing process.
 - (B) Mash feed may contain no animal protein other than an APPI animal protein product supplement manufactured in pellet form and crumbled: Provided, that mash feed may contain nonpelleted APPI animal protein product supplements if the finished feed is treated with a salmonella control product approved by the Food and Drug Administration.
 - (iii) Feed shall be stored and transported in such a manner as to prevent possible contamination;
 - (iv) The flock is maintained in accordance with part 147 of this subchapter with respect to flock sanitation, cleaning and disinfection, and Salmonella isolation, sanitation, and management. Rodents and other pests should be effectively controlled;
 - (v) Environmental samples shall be collected from the flock by an Authorized Agent, in accordance with part 147 of this subchapter, when the flock is 2 to 4 weeks of age. The samples shall be examined bacteriologically for group D salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped. The authorized agent shall also collect samples every 30 days after the first sample has been collected.
 - ~~(vi) If a Salmonella vaccine is used that causes positive reactions with pullorum typhoid antigen, one of the following options must be utilized:
 - (A) Administer the vaccine after the pullorum typhoid testing is done as described in paragraph (d)(1)(vii) of this section.
 - (B) If an injectable bacterin or live vaccine that does not spread is used, keep a sample of 350 birds unvaccinated and banded for identification until the flock reaches at least 4 months of age. Following negative serological and bacteriological examinations as described in paragraph (d)(1)(vii) of this section, vaccinate the banded, non-vaccinated birds.~~
 - ~~(vii) Blood samples from 300 non-vaccinated birds as described in paragraph (d)(1)(vi) of this section shall be tested with either pullorum antigen or by a federally licensed Salmonella enteritidis enzyme-linked immunosorbent assay (ELISA) test when the flock is more than 4 months of age. All birds with positive or inconclusive reactions, up to a maximum of 25 birds, shall be submitted to an authorized laboratory and~~

~~examined for the presence of group D salmonella, in accordance with part 147 of this subchapter. Cultures from positive samples shall be serotyped.~~

- ~~(viii)vi~~ Hatching eggs are collected as quickly as possible, and their sanitation is maintained in accordance with part 147 of this subchapter.
 - ~~(ix)vii~~ Hatching eggs produced by the flock are incubated in a hatchery whose sanitation is maintained in accordance with part 147 of this subchapter and sanitized either by a procedure approved by the Official State Agency or in accordance with part 147 of this subchapter.
- (2) A flock shall not be eligible for this classification if Salmonella enteritidis ser enteritidis (SE) is isolated from a specimen taken from a bird in the flock. Isolation of SE from an environmental or other specimen, as described in paragraph (d)(1)(v) of this section, will require bacteriological examination for SE in an authorized laboratory, in accordance with part 147 of this subchapter, of a random sample of 60 live birds from a flock of 5,000 birds or more, or 30 live birds from a flock with fewer than 5,000 birds. If only one specimen is found positive for SE, the participant may request bacteriological examination of a second sample, equal in size to the first sample, from the flock. If no SE is recovered from any of the specimens in the second sample, the flock will be eligible for the classification.
- ~~(3) A non-vaccinated flock shall be eligible for this classification if Salmonella enteritidis (S. enteritidis ser Enteritidis) is isolated from an environmental sample collected from the flock in accordance with paragraph (d)(1)(v) of this section. Provided, That testing is conducted in accordance with paragraph (d)(1)(vii) of this section each 30 days and no positive samples are found.~~
- (43) In order for a hatchery to sell products of this classification, all products handled shall meet the requirements of the classification.
- (54) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures.

Reason:

The pullorum-typhoid (PT) agglutination test was added to the NPIP for testing egg-type breeder flocks in the late 1980's. It first shows up in the "white book" dated August 1989 under U.S. Sanitation Monitored. When the Salmonella Enteritidis (SE) outbreak in humans associated with eggs was identified in the late 1980's there was a need to identify infected flocks that may produce contaminated eggs. Since SE is a serogroup D1 Salmonella as is S. Pullorum and S. Gallinarum, it was assumed (hoped) that the PT agglutination test would also detect SE infected flocks. Over time, results have shown that the PT agglutination test is not an effective method for the detection of SE infected flocks.

This proposal only removes the PT agglutination test from the U.S. S. Enteritidis Clean classification. It DOES NOT remove it from the U.S. Pullorum-Typhoid Clean classification. In addition to the fact that the PT agglutination test is not an effective method for detecting SE infected flocks, it causes problems for companies that are vaccinating for *Salmonella*. Also, this change resolves the confusion in Standard B(2)(a) over how many reactors to submit for culture.

Sponsor:

Dr. Doug Waltman
Georgia Poultry Laboratory Network

Deleted: Standard B—Bacteriological Examination Procedure¶

(1) Reserved¶

~~(1) Laboratory procedure recommended for the bacteriological examination of egg-type and meat-type breeding flocks with salmonella enteritidis positive environments.¶~~

~~Birds selected for bacteriological examination from egg-type and meat-type breeding flocks positive for Salmonella enteritidis after environmental monitoring should be examined as described in Section B(2)(a) of these Program Standards, with the following exceptions and modifications allowed due to the high number of birds required for examination: ¶~~

~~(a) Except when visibly pathological tissues are present, direct culture, Section B(2)(a)(1) of these standards, may be omitted.¶~~

~~¶~~

(2) Laboratory procedure recommended for the bacteriological examination of Salmonella from birds ¶

~~For egg- and meat-type chickens, turkeys, waterfowl, exhibition poultry, and game birds¶~~

~~All reactors to the pullorum-typhoid tests, up to 25 birds, and birds from Salmonella enteritidis (SE) positive environments should be cultured in accordance with both the direct enrichment (paragraph (a)(1)) and selective enrichment (paragraph (a)(2)) procedures described in this section. Provided, if there are more than four reactors to the pullorum-typhoid tests in the flock, a minimum of four reactors as provided for in 9 CFR~~

~~145.14(a)(6)(ii) shall be submitted to the authorized laboratory for bacteriological examination. Careful aseptic technique should be used when collecting all tissue samples.¶~~

~~For reactors to the pullorum-typhoid tests, if there are more than four reactors in a flock, a minimum of four reactors shall be submitted to the authorized laboratory; if the flock has four or fewer reactors all the reactors must be submitted [145.14(a)(6)(ii)]. The isolation of S. Enteritidis from U.S. S. Enteritidis Clean flocks will result in the submission of 60 live birds from a flock with fewer than 5,000 birds from multiplier egg-type chicken breeding flocks [145.23(d)(2)] or primary egg-type chicken breeding flocks [145.73(d)(2)] and 25 birds from primary meat-type chicken breeding flocks [145.83(e)(3)]. These birds should be cultured in accordance with both direct culture (paragraph (a)(1)) and selective enrichment (paragraph (a)(2)) procedures described in this section. Provided, if there are no grossly abnormal or diseased tissues present, direct culture may be omitted. Careful aseptic technique should be used when collecting all tissue samples.¶~~

Proposal No. 10

Delegates: 145 C

§145.33 Terminology and classification; flocks and products.

(1) U.S. Avian Influenza Clean

This program is intended to be the basis from which the breeding-hatchery industry may conduct a program for the prevention and control of avian influenza. It is intended to determine the presence of avian influenza in multiplier breeding chickens through routine surveillance of each participating breeding flock. A flock and the hatching eggs and chicks produced from it will qualify for this classification when the Official State Agency determines that they have met the following requirements:

- (1) It is a multiplier breeding flock in which a minimum of 30 birds have been tested negative for antibodies to avian influenza using an approved test as described in §145.14 when more than 4 months of age. To retain this classification:
 - (i) A sample of at least 15 birds must be tested negative at intervals of 90 days;
or
 - (ii) A sample of fewer than 15 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of 30 birds is tested within each 90-day period;
or
 - (iii) The flock is tested as provided in §145.14(d) at intervals of 30 days or less and found to be negative, and a total of 15 samples are collected and tested within each 90-day period;
and
- (2) ~~During each 90-day period, all multiplier spent fowl, up to a maximum of 30, A~~ sample of 11 birds must be tested serologically and found negative for antibodies for avian influenza within 21 days prior to movement to slaughter.

Reason: Approved antigen and antibody tests should be allowed to determine Avian Influenza Clean status. Approved tests, including PCR, are referenced in CFR 145.14(d). Additionally, striking the “during each 90 day period, all multiplier spent fowl, up to a maximum of 30” will make this program consistent with the language in the AI programs in 145.23 (h) and 145.73 (f).

Sponsor: Dr. Ken Opengart
Keystone Foods

Proposal No. 11

Delegates: 145 C

§145.33 Terminology and classification; flocks and products.

(1) U.S. Avian Influenza Clean.

This program is intended to be the basis from which the breeding-hatchery industry may conduct a program for the prevention and control of avian influenza. It is intended to determine the presence of avian influenza in multiplier breeding chickens through routine surveillance of each participating breeding flock. A flock and the hatching eggs and chicks produced from it will qualify for this classification when the Official State Agency determines that they have met the following requirements:

- (1) It is a multiplier breeding flock in which a minimum of 30 birds have been tested negative for antibodies to avian influenza using an approved test as described in §145.14 when more than 4 months of age. To retain this classification:
 - (i) A sample of at least 15 birds must be tested negative at intervals of 90 days; **or**
 - (ii) A sample of fewer than 15 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of ~~30~~ 15 birds is tested within each 90-day period;
- or**
- (iii) The flock is tested as provided in §145.14(d) at intervals of 30 days or less and found to be negative, and a total of 15 samples are collected and tested within each 90-day period;

Reason: To correct a previous omission or mistake. A total of 15 birds must be tested in each of the three options to retain the AI Clean classification. (ii) requires 30 birds to be tested, which is inconsistent with the other two options.

Sponsor: Dr. Denise Heard
USDA NPIP Senior Coordinator

Proposal No. 12

Delegates: 145 C

§145.33 Terminology and classification; flocks and products.

(1) U.S. Avian Influenza Clean.

This program is intended to be the basis from which the breeding-hatchery industry may conduct a program for the prevention and control of avian influenza. It is intended to determine the presence of avian influenza in multiplier breeding chickens through routine surveillance of each participating breeding flock. A flock and the hatching eggs and chicks produced from it will qualify for this classification when the Official State Agency determines that they have met the following requirements:

- (1) It is a multiplier breeding flock in which a minimum of 30 birds have been tested negative for antibodies to avian influenza using an approved test as described in §145.14 when more than 4 months of age. To retain this classification:
 - (i) A sample of at least 15 birds must be tested negative at intervals of 90 days; **or**
 - (ii) A sample of fewer than 15 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of 30 birds is tested within each 90-day period;

or

 - (iii) The flock is tested as provided in §145.14(d) at intervals of 30 days or less and found to be negative, and a total of 15 samples are collected and tested within each 90-day period;

and
- (2) During each 90-day period, all multiplier spent fowl, up to a maximum of 30, must be tested ~~serologically~~ and found negative ~~for antibodies~~ for avian influenza within 21 days prior to movement to slaughter.

Reason: Approved antigen and antibody tests should be allowed to determine Avian Influenza Clean status. Approved tests, including PCR, are referenced in CFR 145.14(d). **THIS PROPOSAL RECEIVED INTERIM APPROVAL DURING THE 2017 GENERAL CONFERENCE COMMITTEE MEETING.**

Sponsor: Proposal was amended by the GCC during the 2017 GCC meeting and received Interim approval until the 44th NPIP Biennial Conference.

Proposal No. 13

Delegates: 145 C

§145.33 Terminology and classification; flocks and products.

(n) U.S. H5/H7 Avian Influenza Clean.

This program is intended to be the basis from which the breeding-hatchery industry may conduct a program for the prevention and control of the H5/H7 subtypes of avian influenza. It is intended to determine the presence of the H5/H7 subtypes of avian influenza in multiplier breeding chickens through routine surveillance of each participating breeding flock. A flock and the hatching eggs and chicks produced from it will qualify for this classification when the Official State Agency determines that they have met the following requirements:

- (1) It is a multiplier breeding flock in which a minimum of 30 birds have been tested negative to the H5/H7 subtypes of avian influenza as provided in §145.14(d) when more than 4 months of age. To retain this classification:
 - (i) A sample of at least 15 birds must be tested negative at intervals of 90 days; **or**
 - (ii) A sample of fewer than 15 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of 15 birds is tested within each 90-day period; **or**
 - (iii) The flock is tested as provided in §145.14(d) at intervals of 30 days or less and found to be negative, and a total of 15 samples are collected and tested within each 90-day period; **and**
- (2) During each 90-day period, all multiplier spent fowl, up to a maximum of 30, must be tested negative for H5/H7 subtypes of avian influenza within 21 days prior to movement to slaughter.

Reason: Creating a Subpart C H5/H7 specific AI Clean program similar to H5/H7 AI programs in Subpart D and Subpart E and allowing approved antibody and/or antigen testing.

Sponsor: Dr. Julie Helm
Clemson Livestock Poultry Health

Proposal No. 14 (Amended by TC)

Delegates: 145 D

§145.43 Terminology and classification; flocks and products.

(h) U.S. Newcastle Disease ~~Virus~~ Clean. This program is intended to be the basis from which the breeding-hatchery industry may conduct a program for the prevention and control of Newcastle Disease. It is intended to determine the presence of Newcastle Disease ~~Virus~~ in primary breeding turkeys through vaccination and monitoring of each participating breeding flock. A flock and the hatching eggs and poult produced from it will qualify for this classification when the Official State Agency determines that they have met the following requirements:

(1) It is a primary breeding flock that is either:

(i) Vaccinated for Newcastle Disease ~~Virus~~ using USDA-licensed ~~approved~~ vaccines and response to vaccination is serologically monitored using an approved test as described in §145.14 when more than 4 months of age and meets the criteria in §145.43(h)(2) to retain classification.

OR

(ii) Unvaccinated for Newcastle Disease ~~Virus~~ in which a minimum of 30 birds have tested negative to ND using an approved test as described in §145.14 when more than 4 months of age and meets criteria in §145.43(h)(3) to retain classification.

(2) To retain this classification, **for vaccinated flocks,**

(i) Vaccines for ND~~V~~ must be USDA-licensed ~~approved~~ vaccines ~~manufactured with low virulence live strains-administered~~ during early stages of development ~~up to grow-out-through rearing,~~ and ~~killed-inactivated~~ vaccines as final vaccination ~~no later than 6 weeks~~ prior to the onset of egg production

AND

(ii) The flock has been monitored for antibody response using approved serological tests as listed in §145.14 and the results are compatible with immunological response against ND vaccination

AND

(iii) Testing must include a minimum of 30 birds with a serologic monitoring program ~~beginning at approximately 10 weeks when more than 4 months~~ of age and prior to the onset of production, and not longer than every 90 days thereafter.

(3) To retain this classification **for unvaccinated flocks,**

(i) A minimum of 30 birds per flock must ~~be~~ test negative using an approved test in §145.14 at intervals of 90 days OR

(ii) A sample of fewer than 30 birds may be tested, and found negative, at any one time if all pens are equally represented and a total of 30 birds is tested within each 90-day period; AND

(iii) During each 90-day period, all primary spent fowl, up to a maximum of 30, must test negative to ND within 21 days prior to movement to slaughter.

(4) Newcastle Disease ~~Virus~~ must be a disease reportable to the responsible State authority (State veterinarian, etc.) by all licensed veterinarians. To accomplish this, all laboratories (private, State, and university laboratories) that perform diagnostic procedures on poultry must examine all submitted cases of unexplained respiratory disease, egg production drops, and mortality for ND~~V~~.

§145.45 Terminology and classification; compartments.

(a) U.S. H5/H7 Avian Influenza and ND~~V~~ Clean Compartment.

This program is intended to be the basis from which the primary turkey breeding-hatchery industry may demonstrate the existence and implementation of a program that has been approved by the Official State Agency and the Service to establish a compartment consisting of a primary breeding-hatchery company that is free of H5/H7 avian influenza (AI) and ND~~V~~. For the purpose of the compartment, avian influenza is defined according to the OIE Terrestrial Animal Health Code Chapter 10.4 and Newcastle Disease ~~Virus~~ is defined according to the OIE Terrestrial Animal Health Code Chapter 10.9. This compartment has the purpose of protecting the defined subpopulation and avoiding the introduction and spread of H5/H7 AI and ND~~V~~ within that subpopulation by prohibiting contact with other commercial poultry operations, other domestic and wild birds, and other intensive animal operations. The program shall consist of the following:

(1) Definition of the compartment. Based on the guidelines established by the World Organization for Animal Health (OIE) in the Terrestrial Animal Health Code and the guidelines in this paragraph (a), the primary breeder company will define the compartment with respect to H5/H7 AI and ND~~V~~. Specifically, the company will use a comprehensive biosecurity program to define the compartment as a subpopulation of poultry with a health status for H5/H7 AI and ND~~V~~ that is separate from birds and poultry outside the compartment. The Official State Agency and the Service must approve all documentation submitted to substantiate the defined compartment as adequate to qualify for epidemiological separation from other potential sources of infection of H5/H7 AI and ND~~V~~. Guidelines for the definition of the compartment include:

(i) Definition and description of the subpopulation of birds and their health status. All birds included in the compartment must be U.S. H5/H7 Avian Influenza Clean in accordance with §145.43(g) and ND~~V~~ Clean in accordance with §145.43(h). The poultry must also be located in a State that has an initial State response and containment plan approved by APHIS under §56.10 of this chapter and that participates in the diagnostic surveillance program for H5/H7 low pathogenicity AI as described in §145.15. Within the compartment, all official tests for AI and ND~~V~~, as described in §145.14(d) and §145.14(e), must be conducted in

State or Federal laboratories or in NPIP authorized laboratories that meet the minimum standards described in §147.52 of this subchapter. In addition, the company must provide to the Service upon request any relevant historical and current H5/H7 AI and ~~NDV~~-related data for reference regarding surveillance for the disease within the compartment. Upon request, the Official State Agency may provide such data for other commercial poultry populations located in the State.

(ii) Description of animal identification and traceability processes.

The primary breeder company must also include a description of its animal identification and traceability records, including examples of Veterinary Services (VS) Form 9-5, "Report of Hatcheries, Dealers and Independent Flocks"; VS Form 9-2, "Flock Selection and Testing Report"; VS Form 9-3, "Report of Sales of Hatching Eggs, Chicks and Poults"; VS Form 9-9, "Hatchery Inspection Report"; set and hatch records; egg receipts; and egg/chick invoices for the subpopulation. Documentation must also include breed identification (NPIP stock code). The Service should ensure that an effective flock identification system and traceability system are in place.

(iii) Definition and description of the physical components or establishments of the defined compartment. The primary breeder company must provide documentation establishing that the defined compartment is epidemiologically separated from other poultry and bird populations. The documentation must be approved by the Official State Agency and the Service as indicating adequate epidemiological separation to maintain the compartment's separate health status with respect to H5/H7 AI and ~~NDV~~. The documentation should include descriptions of:

- (A) The physical and spatial factors that separate the compartment from surrounding bird populations and affect the biosecurity status of the compartment.
- (B) Relevant environmental factors that may affect exposure of the birds to AI and ~~NDV~~.
- (C) The functional boundary and fencing that are used to control access to the compartment.
- (D) Facilities and procedures to prevent access by wild birds and to provide separation from other relevant hosts.
- (E) The relevant infrastructural factors that may affect exposure to AI and ~~NDV~~, including the construction and design of buildings or physical components, cleaning and disinfection of buildings and physical components between production groups with quality assurance verification, cleaning and disinfection of equipment, and introduction of equipment or material into the compartment.

(iv) Definition and description of the functional relationships between components of the defined compartment. Functional relationships between components of the compartment include traffic movement and flow at and among premises, personnel movement at and among premises, exposure to live bird populations, and any other factors that could affect biosecurity of the compartment. All physical components of the compartment must be maintained in compliance with hygiene and biosecurity procedures for poultry primary breeding flocks and hatcheries in accordance with part 147 of this subchapter. In addition, the company must provide a biosecurity plan for the compartment and all included components. The biosecurity plan should include:

- (A) Requirements that company employees and contract growers limit their contact with live birds outside the compartment.
- (B) An education and training program for company employees and contractors.
- (C) Standard operating procedures for company employees, contractors, and outside maintenance personnel.
- (D) Requirements for company employees and non-company personnel who visit any premises within the compartment.
- (E) Company veterinary infrastructure to ensure flock monitoring and disease diagnosis and control measures.
- (F) Policies for management of vehicles and equipment used within the compartment to connect the various premises.
- (G) Farm site requirements (location, layout, and construction).
- (H) Pest management program.
- (I) Cleaning and disinfection process.
- (J) Requirements for litter and dead bird removal and/or disposal.

(v) Description of other factors important for maintaining the compartment. The company veterinary infrastructure will assess sanitary measures, environmental risk factors, and management and husbandry practices that relate to the separation of the compartment and the health status of the birds contained within the compartment that may affect risk of exposure to H5/H7 AI and NDV. This assessment must include a description of internal monitoring and auditing systems (e.g., quality assurance and quality control programs) to demonstrate the effectiveness of the compartment. Upon request, the Service will provide the company with information on the epidemiology of H5/H7 AI and NDV and the associated risk pathways in which the components of the compartment are located.

(vi) *Approval or denial.* Based on this documentation provided under this paragraph (a)(1), as well as any other information the Service and the Official State Agency determine to be necessary, the Service and the Official State Agency will approve or deny the classification of the compartment as U.S. H5/H7 Avian Influenza and ND~~V~~ Clean.

(2) *Company activities for maintenance of the compartment.*

(i) The primary breeder company's management of biosecurity, surveillance, and disease control efforts must be uniform and equivalent among all components that are a part of the compartment. Oversight and inspection of these management practices must be conducted by the company's licensed, accredited veterinarians.

(ii) Veterinary staff from the Official State Agency and NPIP staff will work in partnership with licensed, accredited veterinarians to train and certify auditors through Service-approved workshops. The trained auditors will conduct biosecurity and operational audits at least once every 2 years to ensure the integrity of the compartment. These audits will include evaluation of the critical control points and standard operating practices within the compartment, verification of the health status of the flock(s) contained within the compartment, and examination of the biosecurity and management system of the integrated components of the compartment.

(iii) In addition, the company must demonstrate compliance with paragraph (a)(1) of this section for remaining in the U.S. H5/H7 Avian Influenza and ND~~V~~ Clean classifications, surveillance for H5/H7 AI and ND~~V~~ within the compartment, and conducting tests in State or Federal laboratories or in NPIP authorized laboratories. Accredited veterinarians are responsible for the enforcement of active and passive surveillance of H5/H7 AI and ND~~V~~ in primary breeder flocks. Baseline health status must be maintained for all flocks or subpopulations within the compartment, indicating the dates and negative results of all avian influenza and ND~~V~~ surveillance and monitoring testing, the dates and history of last disease occurrence (if any), the number of outbreaks, and the methods of disease control that were applied.

(iv) Documentation will be maintained in the company's database and will be verified as required by the Service and/or the Official State Agency.

(3) *Service and Official State Agency activities for maintenance of the compartment.* The Service will work in cooperation with the Official State Agencies to ensure the continued integrity of any recognized compartments. Activities will include:

(i) Oversight of the establishment and management of compartments;

- (ii) Establishment of effective partnerships between the Service, the Plan, and the primary breeder industry;
- (iii) Approval or denial of classification of compartments as U.S. H5/H7 Avian Influenza and ND~~V~~ Clean Compartments under paragraph (a)(1) of this section;
- (iv) Official certification of the health status of the compartment, and commodities that may be traded from it through participation in the Plan for avian diseases, including the U.S. H5/H7 Avian Influenza Clean program as described in §145.43(g) and ND~~V~~ Clean program as described in §145.43(h) and diagnostic surveillance for H5/H7 low pathogenicity AI as described in §145.15;
- (v) Conducting audits of compartments at least once every 2 years to:

- (A) Confirm that the primary breeding company's establishments are epidemiologically distinct and pathways for the introduction of disease into the compartment are closed through routine operational procedures;

and

- (B) Evaluate and assess the management and husbandry practices relating to biosecurity to determine whether they are in compliance with hygiene and biosecurity procedures for poultry primary breeding flocks and hatcheries in accordance with part 147 of this subchapter;

- (vi) Providing, upon request, model plans for management and husbandry practices relating to biosecurity in accordance with part 147 of this subchapter, risk evaluations in conjunction with the primary breeder industry (including disease surveillance such as VS Form 9-4, "Summary of Breeding Flock Participation"), and diagnostic capability summaries and systems for initial State response and containment plans in accordance with §56.10 of this chapter; and

- (vii) Publicizing and sharing compartment information with international trading partners, upon request, to establish approval and recognition of the compartment, including timeliness and accuracy of disease reporting and surveillance measures as described in §§145.15, ~~and~~ 145.43(g), and 145.43(h).

(4) *Emergency response and notification.* In the case of a confirmed positive of H5/H7 AI and/or ND~~V~~ in the subpopulation of the compartment, the management of the compartment must notify the Service. The Service will immediately suspend the status of the compartment. A compartment will be eligible to resume trade with importing countries only after the compartment has adopted the necessary measures to reestablish the biosecurity level and confirm that H5/H7 AI and/or ND~~V~~ is not present in the compartment and the Service has

reevaluated the management and biosecurity measures of the compartment and approved said compartment for trade.

(b) [Reserved]

Reason: The Primary Breeders propose the addition of an NDV Clean program. See corresponding proposal in Program Standards Subpart F.

Sponsor: Primary Breeder Association
Dr. Dustin Burch, Aviagen Turkeys
Dr. Elena Behnke, Aviagen
Dr. Alberto Torres, Cobb-Vantress
Dr. Travis Schaal, Hy-Line

Proposal No. 15

Delegates: 145 E, 6E

Subpart E—Special Provisions for Hobbyist and Exhibition ~~Waterfowl, Exhibition Poultry, and Game Bird~~ Raised-for-Release Waterfowl Breeding Flocks and Products

§145.51 Definitions.

Except where the context otherwise requires, for the purposes of this subpart the following terms shall be construed, respectively, to mean:

Exhibition Poultry. Domesticated fowl which are bred for the combined purposes of meat or egg production and competitive showing.

Hobbyist Poultry. Domesticated fowl which are bred for the purpose of meat and/or egg production on a small scale as determined by the Official State Agency.

Raised-for-Release Waterfowl. Domesticated fowl that normally swim, such as ducks and geese, grown under confinement for the primary purpose of producing eggs, chicks, started, or mature birds for release on game preserves or in the wild.

Game birds. Domesticated fowl such as pheasants, partridge, quail, grouse, and guineas, but not doves and pigeons.

Waterfowl. Domesticated fowl that normally swim, such as ducks and geese.

§145.52 Participation.

Participating flocks of hobbyist and exhibition ~~waterfowl, exhibition poultry, and game birds,~~ raised-for-release waterfowl, and the eggs, chicks, started, and mature ~~and baby~~ poultry produced from them shall comply with the applicable general provisions of subpart A of this part and the special provisions of this subpart E. The special provisions that apply to meat-type waterfowl flocks are found in subpart I of this part. The special provisions that apply to game bird flocks are found in subpart J of this part.

- (c) It is recommended that waterfowl flocks and gallinaceous flocks ~~in open air facilities~~ be kept separate.
- (f) All participating raised-for-release waterfowl flocks, regardless of whether they are breeders or non-breeders, shall be enrolled under this part 145 subpart E.

§145.53 Terminology and classification; flocks and products.

Participating flocks, and the eggs, chicks, started, and mature ~~and baby~~ poultry produced from them, which have met the respective requirements specified in this section may be designated by the following terms and the corresponding designs illustrated in §145.10.

(a) [Reserved]

(b) ***U.S. Pullorum-Typhoid Clean.***

A flock in which freedom from pullorum and typhoid has been demonstrated to the Official State Agency under the criteria in one of the following paragraphs (b)(1) through (5) of this section (See §145.14 relating to the official blood test where applicable.):

- (5) It is a primary breeding flock located in a State determined to be in compliance with the provisions of paragraph (b)(4) of this section, and in which a sample of 300 birds from flocks of more than 300, and each bird in flocks of 300 or less, has been officially tested for pullorum-typhoid within the past 12 months with no reactors: *Provided*, That a bacteriological examination monitoring program or serological examination monitoring program for game birds acceptable to the Official State Agency and approved by the Service may be used in lieu of annual blood testing: *And Provided further*, That when a flock is a hobbyist or ~~exhibition waterfowl~~ or exhibition poultry primary breeding flock located in a State which has

been deemed to be a U.S. Pullorum-Typhoid Clean State for the past three years, and during which time no isolation of pullorum or typhoid has been made that can be traced to a source in that State, a bacteriological examination monitoring program or a serological examination monitoring program acceptable to the Official State Agency and approved by the Service may be used in lieu of annual blood testing.

(e) ***U.S. H5/H7 Avian Influenza Clean.***

This program is intended to be the basis from which the breeding-hatchery and raised-for-release waterfowl industry may conduct a program for the prevention and control of the H5 and H7 subtypes of avian influenza. It is intended to determine the presence of the H5 and H7 subtypes of avian influenza in hobbyist or exhibition ~~waterfowl, exhibition poultry, and game bird~~ breeding flocks and raised-for-release waterfowl through routine surveillance of each participating ~~breeding~~ flock. A flock or premise, and the hatching eggs, ~~and chicks, started, and mature poultry~~ produced from it, will qualify for this classification when the Official State Agency determines that it has met one of the following requirements:

- (1) It is a primary or multiplier breeding flock in which a minimum of 30 birds has been tested negative to the H5 and H7 subtypes of avian influenza as provided in §145.14(d) when more than 4 months of age; *Provided*, that waterfowl flocks may test a minimum of 30 cloacal swabs for virus isolation. To retain this classification:
 - (i) A sample of at least 30 birds must be tested negative at intervals of ~~180~~ 180 days; or
 - (ii) A sample of fewer than 30 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of 30 birds are tested within each ~~180~~ 180-day period.
- (2) ~~It is a multiplier breeding flock in which a minimum of 30 birds has been tested negative to the H5 and H7 subtypes of avian influenza as provided in §145.14(d) when more than 4 months of age; *Provided*, that waterfowl flocks may test a minimum of 30 cloacal swabs for virus isolation. To retain this classification:~~
 - (i) ~~A sample of at least 30 birds must be tested negative at intervals of 180 days; or~~
 - (ii) ~~A sample of fewer than 30 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of 30 birds are tested within each 180-day period.~~
- (3) ~~A sample of at least 30 birds must be tested and found negative to H5/H7 avian influenza within 21 days prior to movement to slaughter.~~
- (2) For participants with non-breeding flocks retained for raised-for-release or other purposes on the same premises as a breeding flock, a representative sample of at least 30 birds from the participating premise must be tested negative to the H5 and H7 subtypes of avian influenza as provided in §145.14(d) when more than 4 months of age, every 180 days.

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Subpart J—Special Provisions for Egg/Meat-Type Game Bird and Raised-for-Release Game Bird Breeding Flocks and Products

§145.101 Definitions.

Except where the context otherwise requires, for the purposes of this subpart the following terms shall be construed, respectively, to mean:

Egg/Meat-Type Bird. Birds grown under confinement for the primary purpose of producing eggs and/or meat for human consumption.

Raised-for-Release Bird. Birds grown under confinement for the primary purpose of producing eggs, chicks, started, or mature birds for release on game preserves or in the wild.

Game birds. Domesticated fowl such as pheasants, partridge, quail, grouse, and guineas, but not doves and pigeons.

Categories of Operations:

- (a) **Breeder.** An individual or business that maintains a breeding flock for the purpose of producing eggs, chicks, started, or mature birds. A breeder that is also a hatchery and/or grower shall be categorized as a breeder.
- (b) **Hatchery.** An individual or business that does not have a breeding flock, but hatches eggs for the purpose of producing chicks, started, or mature birds. A hatchery that is also a grower shall be categorized as a hatchery.
- (c) **Grower.** An individual or business that does not have a breeding flock or hatchery, but raises birds for the purpose of selling started or mature birds.
- (d) **Dealer.** An individual or business that resells eggs, chicks, started, or mature birds. Products a dealer handles are typically resold within 30 days or less.

Categories of Products:

- (a) **Egg.** Laid by a female bird for the purpose of hatching a chick.
- (b) **Chick.** A bird that is newly hatched from an egg.
- (c) **Started Bird.** A bird that is between the age of a newly, hatched chick and a mature bird.
- (d) **Mature Bird.** A bird that is fully colored and has reached the average maximum size specific to each species.

§145.102 Participation.

Participating flocks of egg/meat-type game birds, raised-for-release game birds, and the products produced from them shall comply with the applicable general provisions of subpart A of this part and the special provisions of this subpart J.

- (a) Products shall lose their identity under Plan terminology when not maintained by Plan participants under the conditions prescribed in §145.5(a).
- (b) Hatching eggs produced by breeding flocks shall be nest clean, fumigated, or otherwise sanitized in accordance with part 147 of this subchapter.
- (c) It is recommended that gallinaceous flocks and waterfowl flocks be kept separate.
- (d) Any nutritive material provided to baby poultry must be free of the avian pathogens that are officially represented in the Plan disease classifications listed in §145.10.
- (e) A flock of game birds that are not breeders, but are located on the same premise as game bird breeders, shall be covered under the same NPIP hatchery approval number as long as the appropriate testing requirements have been met.
- (f) All participating raised-for-release game bird flocks, regardless of whether they are breeders or non-breeders, shall be enrolled under this part 145 subpart J.
- (g) A breeder, hatchery, or grower may also be a dealer without being categorized as a dealer. To resell products under the assigned NPIP number and avoid losing NPIP flock classifications, products must be purchased from an NPIP participant with equal or

greater classifications or from a flock with equivalent or greater testing requirements under official supervision.

- (h) Subject to the approval of the Service and the Official State Agencies in the importing and exporting States, participating flocks may report poultry sales to importing States by using either VS Form 9-3, "Report of Sales of Hatching Eggs, Chicks, and Poults," or by using an invoice form (9-3I) approved by the Official State Agency and the Service to identify poultry sales to clients. If the 9-3I form is used, the following information must be included on the form:
- (1) The form number "9-3I", printed or stamped on the invoice;
 - (2) The seller name and address;
 - (3) The date of shipment;
 - (4) The invoice number;
 - (5) The purchaser name and address;
 - (6) The quantity of products sold;
 - (7) Identification of the products by bird variety or by NPIP stock code as listed in the NPIP APHIS 91-55-078 appendix; and
 - (8) The appropriate NPIP illustrative design in §145.10. One of the designs in §145.10(b) or (g) must be used. The following information must be provided in or near the NPIP design:
 - (i) The NPIP State number and NPIP approval number; and
 - (ii) The NPIP classification for which product is qualified (e.g., U.S. Pullorum-Typhoid Clean).

§145.103 Terminology and classification; flocks and products.

Participating flocks, and the eggs, chicks, started, and mature birds produced from them, which have met the respective requirements specified in this section may be designated by the following terms and the corresponding designs illustrated in §145.10.

(a) **[Reserved]**

(b) **U.S. Pullorum-Typhoid Clean.**

A flock in which freedom from pullorum and typhoid has been demonstrated to the Official State Agency under the criteria in one of the following paragraphs (b)(1) through (3) of this section (See §145.14 relating to the official blood test where applicable.):

- (1) It has been officially blood tested within the past 12 months with no reactors.
- (2) It is a started or mature bird flock that meets the following specifications as determined by the Official State Agency and the Service:
 - (i) The flock is located in a State where all persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which *S. pullorum* or *S. gallinarum* is isolated;
 - (ii) The flock is composed entirely of birds that originated from U.S. Pullorum-Typhoid Clean breeding flocks or from flocks that met equivalent requirements under official supervision; and
 - (iii) The flock is located on a premises where a flock not classified as U.S. Pullorum-Typhoid Clean was located the previous year; *Provided*, That an Authorized Testing Agent must blood test up to 300 birds per flock, as described in §145.14, if the Official State Agency determines that the flock has been exposed to pullorum-typhoid. In making determinations of exposure and setting the number of birds to be blood tested, the Official State Agency shall evaluate the results of any blood tests, described in §145.14(a)(1), that were performed on an unclassified flock located on the

premises during the previous year; the origins of the unclassified flock; and the probability of contacts between the flock for which qualification is being sought and (a) infected wild birds, (b) contaminated feed or waste, or (c) birds, equipment, supplies, or personnel from flocks infected with pullorum-typhoid.

- (3) It is a breeding flock that originated from U.S. Pullorum-Typhoid Clean breeding flocks or from flocks that met equivalent requirements under official supervision, and in which a sample of 300 birds from flocks of more than 300, and each bird in flocks of 300 or less, has been officially tested for pullorum-typhoid within the past 12 months with no reactors: *Provided*, That a bacteriological examination monitoring program or serological examination monitoring program for game birds acceptable to the Official State Agency and approved by the Service may be used in lieu of annual blood testing: And provided Further, That it is located in a State in which it has been determined by the Service that:
- (i) All hatcheries within the State are qualified as “National Plan Hatcheries” or have met equivalent requirements for pullorum-typhoid control under official supervision;
 - (ii) All hatchery supply flocks within the State, are qualified as U.S. Pullorum-Typhoid Clean or have met equivalent requirements for pullorum-typhoid control under official supervision: *Provided*, That if other domesticated fowl, except waterfowl, are maintained on the same premises as the participating flock, freedom from pullorum-typhoid infection shall be demonstrated by an official blood test of each of these fowl;
 - (iii) All shipments of products other than U.S. Pullorum-Typhoid Clean, or equivalent, into the State are prohibited;
 - (iv) All persons performing poultry disease diagnostic services within the State are required to report to the Official State Agency within 48 hours the source of all poultry specimens from which *S. pullorum* or *S. gallinarum* is isolated;
 - (v) All reports of any disease outbreak involving a disease covered under the Plan are promptly followed by an investigation by the Official State Agency to determine the origin of the infection; *Provided*, That if the origin of the infection involves another State, or if there is exposure to poultry in another State from the infected flock, then the National Poultry Improvement Plan will conduct an investigation;
 - (vi) All flocks found to be infected with pullorum or typhoid are quarantined until marketed or destroyed under the supervision of the Official State Agency, or until subsequently blood tested, following the procedure for reacting flocks as contained in §145.14(a)(5), and all birds fail to demonstrate pullorum or typhoid infection;
 - (vii) All poultry, including exhibition, exotic, and game birds, but excluding waterfowl, going to public exhibition shall come from U.S. Pullorum-Typhoid Clean or equivalent flocks, or have had a negative pullorum-typhoid test within 90 days of going to public exhibition;
 - (viii) The flock is located in a State in which pullorum disease or fowl typhoid is not known to exist nor to have existed in hatchery supply flocks within the State during the preceding 24 months.
 - (ix) Discontinuation of any of the conditions or procedures described in paragraphs (b)(3)(i), (ii), (iii), (iv), (v), (vi), (vii), and (viii) of this section.

or the occurrence of repeated outbreaks of pullorum or typhoid in poultry breeding flocks within or originating within the State shall be grounds for the Service to revoke its determination that such conditions and procedures have been met or complied with. Such action shall not be taken until a thorough investigation has been made by the Service and the Official State Agency has been given an opportunity to present its views.

(c) U.S. H5/H7 Avian Influenza Clean.

This program is intended to be the basis from which the game bird industry may conduct a program for the prevention and control of the H5 and H7 subtypes of avian influenza. It is intended to determine the presence of the H5 and H7 subtypes of avian influenza in game bird flocks through routine surveillance of each participating flock. A flock or premise, and the hatching eggs, chicks, started, and mature birds produced from it, will qualify for this classification when the Official State Agency determines that it has met the following requirements:

- (1) It is a flock in which a minimum of 30 birds has been tested negative to the H5 and H7 subtypes of avian influenza as provided in §145.14(d) when more than 4 months of age. To retain this classification:
 - (i) A sample of at least 30 birds must be tested negative at intervals of 90 days; or
 - (ii) A sample of fewer than 30 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of 30 birds are tested within each 90-day period.
- (2) For participants with non-breeding flocks retained for raised-for-release or other purposes on the same premises as a breeding flock, a representative sample of at least 30 birds from the participating premise must be tested negative to the H5 and H7 subtypes of avian influenza as provided in §145.14(d) when more than 4 months of age, every 90 days.

(d) U.S. Salmonella Monitored.

This program is intended to be the basis from which the game bird industry may conduct a program for the prevention and control of salmonellosis. It is intended to reduce the incidence of *Salmonella* organisms in day-old poultry through an effective and practical sanitation program in the hatchery. This will afford other segments of the poultry industry an opportunity to reduce the incidence of *Salmonella* in their products. The following requirements must be met for a flock to be of this classification:

- (1) An Authorized Agent shall collect a minimum of five environmental samples, e.g., chick papers, hatching trays, and chick transfer devices, from the hatchery at least every 30 days. Testing must be performed at an authorized laboratory.
- (2) To claim products are of this classification, all products shall be derived from a hatchery that meets the requirements of the classification.
- (3) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures.

§145.104 Terminology and classification; States.

(a) U.S. Pullorum-Typhoid Clean State.

- (1) A State will be declared a U.S. Pullorum-Typhoid Clean State when it has been determined by the Service that:
 - (i) The State is in compliance with the provisions contained in §145.23(b)(3)(i) through (vii), §145.33(b)(3)(i) through (vii), §145.43(b)(3)(i) through (vi),

§145.53(b)(3)(i) through (vii), §145.73(b)(2)(i), §145.83(b)(2)(i), §145.93(b)(3)(i) through (vii), and 145.103(b)(3)(i) through (ix).
(ii) No pullorum disease or fowl typhoid is known to exist nor to have existed in hatchery supply flocks within the State during the preceding 12 months;
Provided, That pullorum disease or fowl typhoid found within the preceding 24 months in waterfowl, exhibition poultry, and game bird breeding flocks will not prevent a State, which is otherwise eligible, from qualifying.

- (2) Discontinuation of any of the conditions described in paragraph (a)(1)(i) of this section, or repeated outbreaks of pullorum or typhoid occur in hatchery supply flocks described in paragraph (a)(1)(ii) of this section, or if an infection spreads from the originating premises, the Service shall have grounds to revoke its determination that the State is entitled to this classification. Such action shall not be taken until a thorough investigation has been made by the Service and the Official State Agency has been given an opportunity for a hearing in accordance with rules of practice adopted by the Administrator.

Subpart E—Special Provisions for Commercial Upland Egg/Meat-Type Game Birds, Commercial Egg/Meat-Type Waterfowl, Meat-Type Game Bird Slaughter Plants, and Meat-Type Waterfowl Slaughter Plants, Raised for Release Upland Game Birds, and Raised for Release Waterfowl

§146.51 Definitions.

Commercial upland Egg/Meat-Type game birds. ~~Upland game bird pheasants, quail, or partridges~~ Domesticated fowl such as pheasants, partridge, quail, grouse, and guineas, but not doves and pigeons grown under confinement for the primary purposes of producing eggs and/or meat for human consumption.

Commercial Egg/Meat-Type waterfowl. Domesticated ducks or geese grown under confinement for the primary purposes of producing eggs and/or meat for human consumption.

Commercial upland Meat-Type game bird slaughter plant. A ~~commercial upland~~ meat-type game bird slaughter plant that is federally inspected or under State inspection that the U.S. Department of Agriculture's Food Safety and Inspection Service has recognized as equivalent to Federal inspection.

Commercial Meat-Type waterfowl slaughter plant. A ~~commercial~~ meat-type waterfowl slaughter plant that is federally inspected or under State inspection that the U.S. Department of Agriculture's Food Safety and Inspection Service has recognized as equivalent to Federal inspection.

Raised for release upland game birds. ~~Pheasants, quail, and partridge that are raised under confinement for release in game preserves and are not breeding stock.~~

Raised for release waterfowl. ~~Waterfowl that are raised under confinement for release in game preserves and are not breeding stock.~~

Shift. The working period of a group of employees who are on duty at the same time.

§146.52 Participation.

- (a) Participating ~~commercial upland~~ meat-type game bird slaughter plants, ~~commercial~~ meat-type waterfowl slaughter plants, ~~raised for release upland game bird~~ premises, and ~~raised for release waterfowl~~ premises, and ~~commercial upland~~ egg-type game bird and ~~commercial~~ egg-type waterfowl premises producing eggs for human consumption ~~premises~~ shall comply with the applicable general provisions of subpart A of this part and the special provisions of this subpart E.

- (b) ~~Commercial waterfowl~~ Meat-type game bird slaughter plants and ~~commercial upland game bird~~ Meat-type waterfowl slaughter plants that slaughter fewer than 50,000 birds annually are exempt from the special provisions of this subpart E.
- (c) ~~Raised for release upland game bird premises, raised for release waterfowl premises, and commercial upland~~ Egg-type game bird and commercial Egg-type waterfowl producing eggs for human consumption premises that raise with fewer than 25,000 birds annually are exempt from the special provisions of this subpart E.

§146.53 Terminology and classification; slaughter plants and premises.

Participating slaughter plants and flocks which have met the respective requirements specified in this section may be designated by the following terms and the corresponding designs illustrated in §146.9 of this part:

(a) ***U.S. H5/H7 Avian Influenza ~~onitored.~~***

This program is intended to be the basis from which the ~~commercial waterfowl egg/meat-type game bird~~ and ~~commercial upland game bird egg/meat-type waterfowl~~ industry may conduct a program to monitor for the H5/H7 subtypes of avian influenza. It is intended to determine the presence of the H5/H7 subtypes of avian influenza in ~~commercial waterfowl egg/meat-type game birds and commercial upland game birds egg/meat-type waterfowl~~ through routine surveillance of each participating slaughter plant or, in the case of egg-producing flocks, the regular surveillance of these flocks. A slaughter plant or flock will qualify for this classification when the Official State Agency determines that it has met one of the following requirements:

- (1) It is a ~~commercial upland meat-type~~ game bird slaughter plant or ~~commercial meat-type waterfowl~~ slaughter plant where a minimum of 11 birds per shift are tested negative for the H5/H7 subtypes of avian influenza, as provided in §146.13(b), at slaughter;
- (2) It is a ~~commercial upland meat-type~~ game bird slaughter plant or ~~commercial meat-type waterfowl~~ slaughter plant that only accepts ~~commercial upland egg/meat-type~~ game birds or ~~commercial egg/meat-type~~ waterfowl from flocks where a minimum of 11 birds per flock have been tested negative for the H5/H7 subtypes of avian influenza, as provided in §146.13(b), no more than 21 days prior to slaughter; ~~or~~
- (3) It is a ~~commercial upland meat-type~~ game bird slaughter plant or ~~commercial meat-type waterfowl~~ slaughter plant that has an ongoing active and passive surveillance program for H5/H7 subtypes of avian influenza that is approved by the Official State Agency and the Service.
- (4) It is an ~~commercial upland egg-type~~ game bird or ~~egg-type~~ waterfowl flock that produces eggs for human consumption where a minimum of 11 birds per flock have been tested negative to the H5/H7 subtypes of avian influenza as provided in §146.13(b) within 30 days of disposal or within a 12 month period.
- (5) It is an ~~commercial upland egg-type~~ game bird or ~~egg-type~~ waterfowl flock that has an on-going active and passive surveillance program for H5/H7 subtypes of avian influenza that is approved by the Official State Agency and the Service.

(b) ***U.S. H5/H7 Avian Influenza ~~Monitored.~~***

This program is intended to be the basis from which the ~~raised for release upland game bird and raised for release waterfowl~~ industries may conduct a program to monitor for the H5/H7 subtypes of avian influenza. It is intended to determine the presence of the H5/H7 subtypes of avian influenza through routine surveillance of each participating premises. A premises will qualify for the classification when the Official State Agency determines that a representative sample of 30 birds from the participating premises has

~~been tested with negative results for the H5/H7 subtypes of avian influenza, as provided in §146.13(b), every 90 days.~~

Reason: The size, complexity, and uniqueness of the game bird industry has made the facilitation of NPIP provisions extremely difficult and confusing for everyone involved. The current definitions and provisions do not match the production methods and end uses for the game bird industry. As a result, there is a sufficient need to separate the game bird industry from the rest of the hobby and exhibition poultry industry.

We recommend the following changes to simplify the NPIP program and better protect each industry by clarifying where farms should be enrolled, how they should be testing, and promote the uniform application of NPIP provisions throughout the country:

(145E) Remove all references to game birds, because of the creation of subpart 145J. Use more general definitions and cleanup the language to ensure all poultry operations can be captured in 145 E. Create a definition for a hobbyist operation, since it is not currently defined.

(145J) Add 145 J to create a subpart for the game bird industry for the reasons stated above. Combine the breeding and grow out of game birds, rather than having them separate in 145E & 146E. The change will allow all game bird operations to be listed together on the NPIP website. In addition, grow out birds can become breeders when they are released or sold. Therefore, the same testing and classifications as breeder flocks should be available to grow out flocks.

Allow premises with breeders and grow out birds to be covered under one NPIP number, because breeders typically also grow out birds on the same premises. In addition, replacement breeders are usually raised with grow out birds, and spent breeders are typically reconditioned and sold with grow out birds.

Define egg/meat-type vs raised-for-release gamebirds, since it isn't done in part 145. Categorize the types of operations and products that exist in the game bird industry to provide information that allows NPIP officials to accurately register participants and enforce the proper provisions necessary to keep the industry safe.

Add "nest clean", in addition to fumigating and disinfecting, to accurately reflect egg production methods of birds that lay eggs on wire or away from litter.

Allow a breeder, hatchery, or grower, to also be a dealer without the dealer categorization to simplify the registration and record keeping process. A clear statement is provided that mandates products must be purchased from NPIP participants with equal or greater classifications to prevent a loss of classifications. It encourages everyone in the industry to belong to NPIP and test accordingly to ensure products are safer to move and more desirable.

Simplify the Pullorum-Typhoid Clean section to reflect the terminology and production methods in the game bird industry. Include started and mature birds, and define how they can be classified as PT Clean.

Increase the H5/H7 AI Clean testing requirements from 180 to 90 days to match the H5/H7 AI Monitored requirements that were in part 146 subpart E for game birds.

(146E) Update and standardize the layout, terminology, and definitions to match the other subparts within 146. Replace the term "commercial" with "egg/meat-type" to match the other subparts and eliminate confusion with the true definition of the word "commercial". Eliminate all references to grow out and raised-for-release production, since it is included in part 145.

Allow grow out birds to be included with part 145 to increase the testing and classifications available to the flocks making the industry safer. It also enrolls all game bird operations together to make it much easier for NPIP officials to facilitate the program.

Sponsor: Troy L. Laudenslager, Mahantongo Game Farms, Dalmatia, PA
North American Gamebird Association
Dr. Nan Hanshaw, Chief Animal Health Division, PA Department of Agriculture
Dr. Eva Wallner-Pendleton, Penn State Animal Diagnostic Laboratory
Dr. Doug Anderson, GA Poultry Lab Network

Proposal No. 16

Delegates: 145E

§145.53 Terminology and classification; flocks and products.

(e) U.S. H5/H7 Avian Influenza Clean.

This program is intended to be the basis from which the breeding-hatchery industry may conduct a program for the prevention and control of the H5 and H7 subtypes of avian influenza. It is intended to determine the presence of the H5 and H7 subtypes of avian influenza in hobbyist or exhibition waterfowl, exhibition poultry, and game bird breeding flocks through routine surveillance of each participating breeding flock. A flock, and the hatching eggs, ~~and chicks, and mature poultry~~ produced from it, will qualify for this classification when the Official State Agency determines that it has met ~~one of~~ the following requirements:

- (1) It is a primary or multiplier breeding flock in which a minimum of 30 birds has been tested negative to the H5 and H7 subtypes of avian influenza as provided in §145.14(d) when more than 4 months of age; Provided, that waterfowl flocks may test a minimum of 30 cloacal swabs for virus isolation. To retain this classification:

(i) A sample of at least 30 birds must be tested negative at intervals of ~~180~~ 90 days; **or**

(ii) A sample of fewer than 30 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of 30 birds are tested within each 180-day period.

- (2) For participants with non-breeding poultry flocks on the same breeding flock premises that are retained for raise-for-release or other purposes, a minimum of 30 birds must be tested and negative to the H5 and H7 subtypes of avian influenza as provided in §145.14(d) when more than 4 months of age; Provided, that waterfowl flocks may test a minimum of 30 cloacal swabs for virus isolation. To retain this classification:

(i) A sample of at least 30 birds must be tested negative at intervals of 180 days;

or

(ii) A sample of fewer than 30 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of 30 birds are tested within each 180-day period.

- ~~(2) It is a multiplier breeding flock in which a minimum of 30 birds has been tested negative to the H5 and H7 subtypes of avian influenza as provided in §145.14(d) when more than 4 months of age; Provided, that waterfowl flocks may test a minimum of 30 cloacal swabs for virus isolation. To retain this classification:~~

~~(i) A sample of at least 30 birds must be tested negative at intervals of 180 days;~~

~~**or**~~

~~(ii) A sample of fewer than 30 birds may be tested, and found to be negative, at any one time if all pens are equally represented and a total of 30 birds are tested within each 180-day period.~~

- ~~(3) A sample of at least 30 birds must be tested and found negative to H5/H7 avian influenza within 21 days prior to movement to slaughter.~~

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Reason: To provide better credibility to these classifications, requirements for Subpart 145 E, U.S. H5/H7 Avian Influenza Clean participation (145.53) should be equivalent to or greater than the Subpart 146 E, H5/H7 Avian Influenza Monitored participation requirements.

Sponsor: Dr. Shauna Voss, Minnesota Board of Animal Health, Willmar, MN
Dr. Dale Lauer, Minnesota Board of Animal Health, Willmar, MN
Scott Meyer, Oakwood Game Farm, Princeton, MN
Michael Forsgren, Forsgren Pheasant Farm, Pelican Rapids, MN

Proposal No. 17 (AMENDMENT TO ORIGINAL PROPOSAL)

Delegates: 145E

§145.53 Terminology and classification; flocks and products
(f) U.S. Salmonella Monitored

This program is intended to be the basis from which the breeding-hatching industry may conduct a program for the prevention and control of salmonellosis. It is intended to reduce the incidence of Salmonella organisms in hatching eggs and day-old poultry through an effective and practical sanitation and testing program at the breeder farm and in the hatchery. This will afford other segments of the poultry industry an opportunity to reduce the incidence of Salmonella in their products. The following requirements must be met for a flock or hatchery to be eligible for ~~of~~ this classification as determined by the Official State Agency:

- ~~(1) An Authorized Agent shall collect a minimum of five environmental samples, e.g., chick papers, hatching trays, and chick transfer devices, from the hatchery at least every 30 days. Testing must be performed at an authorized laboratory.~~
- ~~(2) To claim products are of this classification, all products shall be derived from a hatchery that meets the requirements of the classification.~~
- ~~(3) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures.~~
- (1) Hatcheries must be kept in a sanitary condition as applicable and as outlined in plan provisions §145.6 Specific provisions for participating hatcheries and NPIP Program Standards C.
- (2) An Authorized Agent shall collect and submit to an authorized laboratory:
 - (i) A minimum of five samples from the hatchery at least every 30 days while in operation. These samples may include: hatchery debris, swabs from hatcher, setters, hatchery environment, hatchery equipment, sexing tables and belts, meconium, chick box papers, hatching trays, or chick transfer devices. Samples will be examined bacteriologically at an authorized laboratory for *Salmonella*; and
 - (ii) Annual environmental samples from each pullet and breeder farm flock in accordance with Program Standards Standard B (3)(a)(1). Samples will be examined bacteriologically at an authorized laboratory for *Salmonella*.
- (3) If *Salmonella* is identified through this testing:
 - (i) A qualified poultry health professional knowledgeable with the operation will be consulted and will:
 - (A) Review test results to evaluate the *Salmonella* monitoring program.
 - (B) Use the *Salmonella* monitoring program test results to develop appropriate and practical *Salmonella* intervention measures.
 - ~~(ii) conduct periodic on-site visits until samples at the hatchery are *Salmonella* negative;~~
 - ~~(iii) As recommended by the poultry health professional, additional testing will be conducted to determine the source of *Salmonella*.~~
 - ~~(iii) The participant shall provide a corrective action plan to the OSA within ten business days of the final test report. The corrective action plan shall include:~~

Deleted: E

Deleted: pre-lay (before egg production begins)

Deleted: and mid-lay (mid-point of scheduled egg production) when the flock is in production.

~~(A) Notification of the customer and chick distributor(s) of the hatchery status;~~

~~(B) Plans for cleaning and disinfection of the hatchery and/or poultry house per NPIP Program Standards C;~~

~~(C) Development of appropriate and practical *Salmonella* control measures at the breeder flock and hatchery acceptable to the OSA;~~

~~(D) A review of the breeder flock biosecurity plan to evaluate *Salmonella* control measures;~~

~~(iv) Allow OSA officials to inspect the hatchery and breeder flocks upon request to monitor compliance.~~

(2) (4) To claim products are of this classification, all products shall be derived from a farm or hatchery that meets the requirements of the classification.

~~(3) (5) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures.~~

Deleted: flock

Reason:

Backyard poultry flocks have increased in popularity in recent years. Poultry are well recognized as possible carriers of *Salmonella* which result in thousands of illnesses each year, with some of these illnesses resulting in human death. Between 1996 and 2017, 65 outbreaks of human *Salmonella* infections linked to live poultry from mail-order hatcheries were documented, however, these outbreaks likely underestimate the true burden of illness resulting from contact with backyard poultry. Live poultry such as chickens and ducks can be carrying *Salmonella* bacteria but appear healthy and clean, with no sign of illness, and as raising backyard flocks becomes more popular, more people are having contact with chickens and ducks – and may not understand the risk of *Salmonella* infection.

The sponsors of this proposal recognize that additional poultry dealer and retail outlet sanitation measures, public education and public awareness efforts are needed to further reduce the number of human *Salmonella* cases. This proposal is a first step in the *Salmonella* surveillance and control process. Currently the Subpart E U.S. *Salmonella* Monitored classification only requires hatchery testing which is inadequate to assess the *Salmonella* burden from hatching eggs feeding into the hatchery. This proposal will outline a process that Subpart E participants who wish to obtain this classification can follow to reduce *Salmonella* in both breeder flocks and hatcheries.

Sponsors:

Dr. Shauna Voss, Minnesota Board of Animal Health, Willmar, MN

Dr. Dale Lauer, Minnesota Board of Animal Health, Willmar, MN

Dr. Nanette Hanshaw, Pennsylvania Department of Agriculture

Dr. Doug Waltman, Georgia Poultry Laboratory Network, Gainesville, GA

RED HIGHLIGHTED TEXT INDICATE DELETIONS AND/OR ADDITIONS TO ORIGINAL PROPOSAL #17

Proposal No. 18

Delegates: 145 G

§145.73 Terminology and classification; flocks and products.

(d) U.S. S. Enteritidis Clean.

This classification is intended for primary egg-type breeders wishing to assure their customers that the hatching eggs and multiplier chicks produced are certified free of Salmonella enteritidis.

- (1) A flock and the hatching eggs and chicks produced from it which have met the following requirements as determined by the Official State Agency:
 - (i) The flock originated from a U.S. S. Enteritidis Clean flock, or meconium from the chick boxes and a sample of chicks that died within 7 days after hatching are examined bacteriologically for salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped.
 - (ii) All feed fed to the flock shall meet the following requirements:
 - (A) Pelletized feed shall contain either no animal protein or only animal protein products produced under the Animal Protein Products Industry (APPI) Salmonella Education/Reduction Program. The protein products must have a minimum moisture content of 14.5 percent and must have been heated throughout to a minimum temperature of 190 °F, or above, or to a minimum temperature of 165 °F for at least 20 minutes, or to a minimum temperature of 184 °F under 70 lbs. pressure during the manufacturing process.
 - (B) Mash feed may contain no animal protein other than an APPI animal protein product supplement manufactured in pellet form and crumbled: Provided, That mash feed may contain nonpelleted APPI animal protein product supplements if the finished feed is treated with a salmonella control product approved by the U.S. Food and Drug Administration.
 - (iii) Feed shall be stored and transported in such a manner as to prevent possible contamination;
 - (iv) The flock is maintained in accordance with part 147 of this subchapter with respect to flock sanitation, cleaning and disinfection, and Salmonella isolation, sanitation, and management. Rodents and other pests should be effectively controlled;
 - (v) Environmental samples shall be collected from the flock by an Authorized Agent, in accordance with part 147 of this subchapter, when the flock is 2 to 4 weeks of age. The samples shall be examined bacteriologically for group D salmonella at an authorized laboratory. Cultures from positive samples shall be serotyped. The Authorized Agent shall also collect samples every 30 days after the first sample has been collected.
 - ~~(vi) If a Salmonella vaccine is used that causes positive reactions with pullorum typhoid antigen, one of the following options must be utilized:
 - ~~(A) Administer the vaccine after the pullorum typhoid testing is done as described in paragraph (d)(1)(vii) of this section.~~
 - ~~(B) If an injectable bacterin or live vaccine that does not spread is used, keep a sample of 350 birds unvaccinated and banded for identification until the flock reaches at least 4 months of age. Following negative serological and bacteriological examinations as described in paragraph (d)(1)(vii) of this section, vaccinate the banded, non-vaccinated birds.~~~~
 - ~~(vii) Blood samples from 300 non-vaccinated birds as described in paragraph (d)(1)(vi) of this section shall be tested with either pullorum antigen or by a federally licensed Salmonella enteritidis enzyme-linked immunosorbent assay (ELISA) test when the flock is more than 4 months of age. All birds with positive or inconclusive reactions, up to a maximum of 25 birds, shall be submitted to an authorized laboratory and~~

examined for the presence of group D salmonella, in accordance with part 147 of this subchapter. Cultures from positive samples shall be serotyped.

- (viii) Hatching eggs are collected as quickly as possible and are handled as described in §147.22 of this subchapter and are sanitized or fumigated (see §147.25 of this subchapter).
- (ix) Hatching eggs produced by the flock are incubated in a hatchery whose sanitation is maintained in accordance with part 147 of this subchapter and sanitized either by a procedure approved by the Official State Agency or in accordance with part 147 of this subchapter.
- (2) A flock shall not be eligible for this classification if Salmonella enteritidis serotype enteritidis (SE) is isolated from a specimen taken from a bird in the flock. Isolation of SE from an environmental or other specimen, as described in paragraph (d)(1)(v) of this section, will require bacteriological examination for SE in an authorized laboratory, in accordance with part 147 of this subchapter, of a random sample of 60 live birds from a flock of 5,000 birds or more, or 30 live birds from a flock with fewer than 5,000 birds. If only one specimen is found positive for SE, the participant may request bacteriological examination of a second sample, equal in size to the first sample, from the flock. If no SE is recovered from any of the specimens in the second sample, the flock will be eligible for the classification.
- ~~(3) A non-vaccinated flock shall be eligible for this classification if SE is isolated from an environmental sample collected from the flock in accordance with paragraph (d)(1)(v) of this section. Provided, That testing is conducted in accordance with paragraph (d)(1)(vii) of this section each 30 days and no positive samples are found.~~
- (4) In order for a hatchery to sell products of this classification, all products handled shall meet the requirements of the classification.
- (5) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures. The Official State Agency shall not revoke the participant's classification until the participant has been given an opportunity for a hearing in accordance with rules of practice adopted by the Official State Agency.

Reason:

The pullorum-typhoid (PT) agglutination test was added to the NPIP for testing egg-type breeder flocks in the late 1980's. It first shows up in the "white book" dated August 1989 under U.S. Sanitation Monitored. When the Salmonella Enteritidis (SE) outbreak in humans associated with eggs was identified in the late 1980's there was a need to identify infected flocks that may produce contaminated eggs. Since SE is a serogroup D1 Salmonella as is S. Pullorum and S. Gallinarum, it was assumed (hoped) that the PT agglutination test would also detect SE infected flocks. Over time, results have shown that the PT agglutination test is not an effective method for the detection of SE infected flocks.

This proposal only removes the PT agglutination test from the U.S. S. Enteritidis Clean classification. It DOES NOT remove it from the U.S. Pullorum-Typhoid Clean classification. In addition to the fact that the PT agglutination test is not an effective method for detecting SE infected flocks, it causes problems for companies that are vaccinating for *Salmonella*. Also, this change resolves the confusion in Standard B(2)(a) over how many reactors to submit for culture.

Sponsor:

Dr. Doug Waltman
Georgia Poultry Laboratory Network

Deleted: Standard B—Bacteriological Examination Procedure¶

(1) Reserved¶

~~(1) Laboratory procedure recommended for the bacteriological examination of egg-type and meat-type breeding flocks with salmonella enteritidis positive environments.¶~~

~~Birds selected for bacteriological examination from egg-type and meat-type breeding flocks positive for Salmonella enteritidis after environmental monitoring should be examined as described in Section B(2)(a) of these Program Standards, with the following exceptions and modifications allowed due to the high number of birds required for examination: ¶~~

~~(a) Except when visibly pathological tissues are present, direct culture, Section B(2)(a)(1) of these standards, may be omitted.¶~~

~~¶~~

(2) Laboratory procedure recommended for the bacteriological examination of Salmonella from birds ¶

For egg- and meat-type chickens, turkeys, waterfowl, exhibition poultry, and game birds¶

All reactors to the pullorum-typhoid tests, up to 25 birds, and birds from Salmonella enteritidis (SE) positive environments should be cultured in accordance with both the direct enrichment (paragraph (a)(1)) and selective enrichment (paragraph (a)(2)) procedures described in this section: Provided, if there are more than four reactors to the pullorum-typhoid tests in the flock, a minimum of four reactors as provided for in 9 CFR 145.14(a)(6)(ii) shall be submitted to the authorized laboratory for bacteriological examination. Careful aseptic technique should be used when collecting all tissue samples.¶

For reactors to the pullorum-typhoid tests, if there are more than four reactors in a flock, a minimum of four reactors shall be submitted to the authorized laboratory; if the flock has four or fewer reactors all the reactors must be submitted [145.14(a)(6)(ii)]. The isolation of S. Enteritidis from U.S. S. Enteritidis Clean flocks will result in the submission of 60 live birds from a flock of 5,000 birds or more, or 30 live birds from a flock with fewer than 5,000 birds from multiplier egg-type chicken breeding flocks [145.23(d)(2)] or primary egg-type chicken breeding flocks [145.73(d)(2)] and 25 birds from primary meat-type chicken breeding flocks [145.83(e)(3)]. These birds should be cultured in accordance with both direct culture (paragraph (a)(1)) and selective enrichment (paragraph (a)(2)) procedures described in this section. Provided, if there are no grossly abnormal or diseased tissues present, direct culture may be omitted. Careful aseptic technique should be used when collecting all tissue samples.¶

Proposal No. 19 (Amended by TC)

Delegates: 145 G

§145.73 Terminology and classification; flocks and products.

(h) U.S. Newcastle Disease ~~Virus~~ Clean. This program is intended to be the basis from which the breeding-hatchery industry may conduct a program for the prevention and control of Newcastle Disease. It is intended to determine the presence of Newcastle Disease ~~Virus~~ in primary breeding chickens through vaccination and monitoring of each participating breeding flock. A flock and the hatching eggs and chicks produced from it will qualify for this classification when the Official State Agency determines that they have met the following requirements:

(1) It is a primary breeding flock that is either:

(i) Vaccinated for Newcastle Disease ~~Virus~~ using USDA-licensed approved vaccines and response to vaccination is serologically monitored using an approved test as described in §145.14 when more than 4 months of age and meets the criteria in §145.73(h)(2) to retain classification.

OR

(ii) Unvaccinated for Newcastle Disease ~~Virus~~ in which a minimum of 30 birds have tested negative to ND using an approved test as described in §145.14 when more than 4 months of age and meets criteria in §145.73(h)(3) to retain classification.

(2) To retain this classification, for vaccinated flocks,

(i) Vaccines for ND~~V~~ must be USDA-licensed approved vaccines ~~manufactured with low-virulence live strains administered~~ during early stages of development ~~up to grow-out through rearing~~, and ~~killed-inactivated~~ vaccines as final vaccination ~~no later than 6 weeks~~ prior to the onset of egg production

AND

(ii) The flock has been monitored for antibody response using approved serological tests as listed in §145.14 and the results are compatible with immunological response against ND vaccination

AND

(iii) Testing must include a minimum of 30 birds with a serologic monitoring program ~~beginning at approximately 10 weeks when more than 4 months~~ of age ~~and prior to the onset of production~~, and not longer than every 90 days thereafter.

(3) To retain this classification for unvaccinated flocks,

(i) A minimum of 30 birds per flock must ~~be~~ test negative using an approved test in §145.14 at intervals of 90 days OR

(ii) A sample of fewer than 30 birds may be tested, and found negative, at any one time if all pens are equally represented and a total of 30 birds is tested within each 90-day period;

AND

(iii) During each 90-day period, all primary spent fowl, up to a maximum of 30, must test negative to ND within 21 days prior to movement to slaughter.

(4) Newcastle Disease ~~Virus~~ must be a disease reportable to the responsible State authority (State veterinarian, etc.) by all licensed veterinarians. To accomplish this, all laboratories (private, State, and university laboratories) that perform diagnostic procedures on poultry must examine all submitted cases of unexplained respiratory disease, egg production drops, and mortality for ND~~V~~.

§145.74 Terminology and classification; compartments.

(a) U.S. Avian Influenza and NDV Clean Compartment.

This program is intended to be the basis from which the primary egg-type chicken breeding-hatchery industry may demonstrate the existence and implementation of a program that has been approved by the Official State Agency and the Service to establish a compartment consisting of a primary breeding-hatchery company that is free of H5/H7 avian influenza (AI) and NDV. This compartment has the purpose of protecting the defined subpopulation and avoiding the introduction and spread of H5/H7 AI and NDV within that subpopulation by prohibiting contact with other commercial poultry operations, other domestic and wild birds, and other intensive animal operations. The program shall consist of the following:

(1) Definition of the compartment. Based on the guidelines established by the World Organization for Animal Health (OIE) in the Terrestrial Animal Health Code and the guidelines in this paragraph (a), the primary breeder company will define the compartment with respect to H5/H7 AI and NDV. Specifically, the company will use a comprehensive biosecurity program to define the compartment as a subpopulation of poultry with a health status for H5/H7 AI and NDV that is separate from birds and poultry outside the compartment. The Official State Agency and the Service must first approve all documentation submitted by the company to substantiate the defined compartment as adequate to qualify for epidemiological separation from other potential sources of infection of H5/H7 AI and NDV. Guidelines for the definition of the compartment include:

- (i) Definition and description of the subpopulation of birds and their health status. All birds included in the compartment must be U.S. Avian Influenza Clean in accordance with §145.73(f) and NDV Clean in accordance with §145.73(h). The poultry must also be located in a State that has an initial State response and containment plan approved by APHIS under §56.10 of this chapter and that participates in the diagnostic surveillance program for H5/H7 low pathogenicity AI as described in §145.15. Within the compartment, all official tests for AI and NDV, as described in §145.14(d) and §145.14(e), must be conducted in State or Federal laboratories or in NPIP authorized laboratories that meet the minimum standards described in §147.52 of this subchapter. In addition, the company must provide to the Service upon request any relevant historical and current H5/H7 AI and NDV-related data for reference regarding surveillance for the disease within the compartment. Upon request, the Official State Agency may provide such data for other commercial poultry populations located in the State.
- (ii) Description of animal identification and traceability processes. The primary breeder company must also include a description of its animal identification and traceability records, including examples of Veterinary Services (VS) Form 9-5, "Report of Hatcheries, Dealers and Independent Flocks"; VS Form 9-2, "Flock Selection and Testing Report"; VS Form 9-3, "Report of Sales of Hatching Eggs, Chicks and Poults"; VS Form 9-9, "Hatchery Inspection Report"; set and hatch records; egg receipts; and egg/chick invoices for the subpopulation. Documentation must also include breed identification (NPIP stock code). The Service should ensure that an effective flock identification system and traceability system are in place.
- (iii) Definition and description of the physical components or establishments of the defined compartment. The primary breeder company must provide documentation establishing that the defined compartment is epidemiologically separated from other poultry and bird populations. The documentation must be approved by the Official State Agency and the Service as indicating adequate epidemiological separation to maintain the compartment's separate health status with respect to H5/H7 AI and NDV. The documentation should include descriptions of:

- (A) The physical and spatial factors that separate the compartment from surrounding bird populations and affect the biosecurity status of the compartment.
- (B) Relevant environmental factors that may affect exposure of the birds to AI and NDV.
- (C) The functional boundary and fencing that are used to control access to the compartment.
- (D) Facilities and procedures to prevent access by wild birds and to provide separation from other relevant hosts.
- (E) The relevant infrastructural factors that may affect exposure to AI and NDV, including the construction and design of buildings or physical components, cleaning and disinfection of buildings and physical components between production groups with quality assurance verification, cleaning and disinfection of equipment, and introduction of equipment or material into the compartment.

(iv) Definition and description of the functional relationships between components of the defined compartment. Functional relationships between components of the compartment include traffic movement and flow at and among premises, personnel movement at and among premises, exposure to live bird populations, and any other factors that could affect biosecurity of the compartment. All physical components of the compartment must be maintained in compliance with hygiene and biosecurity procedures for poultry primary breeding flocks and hatcheries in accordance with part 147 of this subchapter. In addition, the company must provide a biosecurity plan for the compartment and all included components. The biosecurity plan should include but not be limited to:

- (A) Requirements that company employees and contract growers limit their contact with live birds outside the compartment.
- (B) An education and training program for company employees and contractors.
- (C) Standard operating procedures for company employees, contractors, and outside maintenance personnel.
- (D) Requirements for company employees and non-company personnel who visit any premises within the compartment.
- (E) Company veterinary infrastructure to ensure flock monitoring and disease diagnosis and control measures.
- (F) Policies for management of vehicles and equipment used within the compartment to connect the various premises.
- (G) Farm site requirements (location, layout, and construction).
- (H) Pest management program.
- (I) Cleaning and disinfection process.
- (J) Requirements for litter and dead bird removal and/or disposal.

(v) Description of other factors important for maintaining the compartment. The company veterinary infrastructure will assess sanitary measures, environmental risk factors, and management and husbandry practices that relate to the separation of the compartment and the health status of the birds contained within the compartment that may affect risk of exposure to H5/H7 AI and NDV. This assessment must include a description of internal monitoring and auditing systems (e.g., quality assurance and quality control programs) to demonstrate the effectiveness of the compartment. Upon request, the Service will provide the company with information on the epidemiology of H5/H7 AI and NDV and the associated risk pathways in which the components of the compartment are located.

(vi) Approval or denial. Based on the documentation provided under this paragraph (a)(1), as well as any other information the Service and the Official State Agency determine to be necessary, the Service and the Official State Agency will approve or deny the classification of the compartment as U.S. Avian Influenza and ND~~V~~ Clean.

(2) *Company activities for maintenance of the compartment.*

(i) The primary breeder company's management of biosecurity, surveillance, and disease control efforts must be uniform and equivalent among all components that are a part of the compartment. Oversight and inspection of these management practices must be conducted by the company's licensed, accredited veterinarians.

(ii) Veterinary staff from the Official State Agency and NPIP staff will work in partnership with licensed, accredited veterinarians to train and certify auditors through Service-approved workshops. The trained auditors will conduct biosecurity and operational audits at least once every 2 years to ensure the integrity of the compartment. These audits will include evaluation of the critical control points and standard operating practices within the compartment, verification of the health status of the flock(s) contained within the compartment, and examination of the biosecurity and management system of the integrated components of the compartment.

(iii) In addition, the company must demonstrate compliance with paragraph (a)(1) of this section for remaining in the U.S. Avian Influenza and ND~~V~~ Clean classifications, surveillance for H5/H7 AI and ND~~V~~ within the compartment, and conducting tests in State or Federal laboratories or in NPIP authorized laboratories. Accredited veterinarians are responsible for the enforcement of active and passive surveillance of H5/H7 AI and ND~~V~~ in primary breeder flocks. Baseline health status must be maintained for all flocks or subpopulations within the compartment, indicating the dates and negative results of all avian influenza and ND~~V~~ surveillance and monitoring testing, the dates and history of last disease occurrence (if any), the number of outbreaks, and the methods of disease control that were applied.

(iv) Documentation will be maintained in the company's database and will be verified as required by the Service and/or the Official State Agency.

(3) *Service and Official State Agency activities for maintenance of the compartment.* The Service will work in cooperation with the Official State Agencies to ensure the continued integrity of any recognized compartments. Activities include:

(i) Oversight of the establishment and management of compartments;

(ii) Establishment of effective partnerships between the Service, the Plan, and the primary breeder industry;

(iii) Approval or denial of classification of compartments as U.S. Avian Influenza and ND~~V~~ Clean Compartments under paragraph (a)(1) of this section;

(iv) Official certification of the health status of the compartment, and commodities that may be traded from it through participation in the Plan for avian diseases, including the U.S. Avian Influenza Clean program as described in §145.73(f) and ND~~V~~ Clean program as described in §145.73(h) and diagnostic surveillance for H5/H7 low pathogenicity AI as described in §145.15;

(v) Conducting audits of compartments at least once every 2 years to:

(A) Confirm that the primary breeding company's establishments are epidemiologically distinct and pathways for the introduction of disease into the compartment are closed through routine operational procedures;

and

(B) Evaluate and assess the management and husbandry practices relating to biosecurity to determine whether they are in compliance with hygiene

and biosecurity procedures for poultry primary breeding flocks and hatcheries in accordance with part 147 of this subchapter;

(vi) Providing, upon request, model plans for management and husbandry practices relating to biosecurity in accordance with part 147 of this subchapter, risk evaluations in conjunction with the primary breeder industry (including disease surveillance such as VS Form 9-4, “Summary of Breeding Flock Participation”), and diagnostic capability summaries and systems for initial State response and containment plans in accordance with §56.10 of this chapter; and

(vii) Publicizing and sharing compartment information with international trading partners, upon request, to establish approval and recognition of the compartment, including timeliness and accuracy of disease reporting and surveillance measures as described in §§145.15, ~~and~~ 145.73(f), ~~and~~ 145.73(h).

(4) *Emergency response and notification.* In the case of a confirmed positive of H5/H7 AI ~~and/or NDV~~ in the subpopulation of the compartment, the management of the compartment must notify the Service. The Service will immediately suspend the status of the compartment. A compartment will be eligible to resume trade with importing countries only after the compartment has adopted the necessary measures to reestablish the biosecurity level and confirm that H5/H7 AI ~~and/or NDV~~ is not present in the compartment and the Service has reevaluated the management and biosecurity measures of the compartment and approved said compartment for trade.

Reason: The Primary Breeders propose the addition of an NDV Clean program. See corresponding proposal in Program Standards Subpart F.

Sponsor: Primary Breeder Association
Dr. Travis Schaal, Hy-Line
Dr. Elena Behnke, Aviagen
Dr. Alberto Torres, Cobb-Vantress
Dr. Dustin Burch, Aviagen Turkeys

Proposal No. 20

Delegates: 145 H

§145.83 Terminology and classification; flocks and products.

(e) U.S. S. Enteritidis Clean.

This classification is intended for primary meat-type breeders wishing to assure their customers that the chicks produced are certified free of *Salmonella enteritidis*.

- (1) A flock and the hatching eggs and chicks produced from it shall be eligible for this classification if they meet the following requirements, as determined by the Official State Agency:
 - (i) The flock originated from a U.S. S. Enteritidis Clean flock, or one of the following samples has been examined bacteriologically for *S. enteritidis* at an authorized laboratory in accordance with part 147 of this subchapter and any group D *Salmonella* samples have been serotyped:
 - (A) A sample of chick papers, hatcher tray swabs, or fluff collected and cultured in accordance with part 147 of this subchapter; and
 - (B) Samples of intestinal and liver or spleen tissues from a minimum of 30 chicks that died within 7 days after hatching and have been preserved daily by freezing prior to shipment to an authorized laboratory.
 - (ii) The flock is maintained in compliance with isolation, sanitation, and management procedures for *Salmonella* in accordance with part 147 of this subchapter.
 - (iii) Environmental samples are collected from the flock by or under the supervision of an Authorized Agent, in accordance with part 147 of this subchapter, when the flock reaches 4 months of age and every 30 days thereafter. Once the flock is in egg production and chicks are hatching from it, the samples must include at least 4 individual test assay results every 30 days in flocks of more than 500 birds or 2 individual assays per month in flocks of 500 birds or fewer. One of these results must come from samples collected from hatched chicks at a participating hatchery derived from said flock. These individual test assays may be derived from pooled samples from the farm or hatchery in accordance with part 147 of this subchapter, but must be run as separate test assays in the laboratory. The environmental samples shall be examined bacteriologically for group D *Salmonella* at an authorized laboratory, and cultures from group D positive samples shall be serotyped.
 - ~~(iv) Blood samples from 300 birds from the flock are officially tested with pullorum antigen when the flock is at least 4 months of age. All birds with positive or inconclusive reactions, up to a maximum of 25 birds, shall be submitted to an authorized laboratory and examined for the presence of group D *Salmonella* in accordance with part 147 of this subchapter. Cultures from group D positive samples shall be serotyped.~~
 - ~~(v) Hatching eggs produced by the flock are collected as quickly as possible and their sanitation is maintained in accordance with part 147 of this subchapter.~~
 - ~~(vi) Hatching eggs produced by the flock are incubated in a hatchery whose sanitation is maintained in accordance with part 147 of this subchapter, and the hatchery must have been sanitized either by a procedure approved by the Official State Agency or by fumigation in accordance with part 147 of this subchapter.~~
- (2) If *Salmonella enteritidis* serotype enteritidis (SE) is isolated from a specimen taken from a bird in the flock, except as provided in paragraph (e)(3) of this section, the flock shall not be eligible for this classification.
- (3) If SE is isolated from an environmental sample collected from the flock in accordance with paragraph (e)(1)(iii) of this section, an additional environmental sampling and 25 live cull birds or fresh dead birds (if present), or other randomly selected live birds if fewer than 25

culls can be found in the flock, must be bacteriologically examined for SE in accordance with part 147 of this subchapter. If only 1 bird from the 25-bird sample is found positive for SE., the participant may request bacteriological examination of a second 25-bird sample from the flock. In addition, if the flock with the SE isolation is in egg production and eggs are under incubation, the next four consecutive hatches shall be examined bacteriologically in accordance with part 147 of this subchapter. Samples shall be collected from all of the hatching unit's chick trays and basket trays of hatching eggs, or from all chick box papers from the flock, and tested, pooling the samples into a minimum of 10 separate assays. Any followup hatchery-positive SE isolations shall result in discontinuation of subsequent hatches until the flock status is determined by bird culture. The flock will be disqualified for the U.S. S. Enteritidis Clean classification if a bird or subsequent flock environmental assay results in isolation of SE.

- (4) In order for a hatchery to sell products of this classification, all products handled by the hatchery must meet the requirements of this paragraph.
- (5) This classification may be revoked by the Official State Agency if the participant fails to follow recommended corrective measures. The Official State Agency shall not revoke the participant's classification until the participant has been given an opportunity for a hearing in accordance with rules of practice adopted by the Official State Agency.
- (6) A pedigree, experimental, great-grandparent, or grandparent flock that is removed from the U.S. S. Enteritidis Clean program may be reinstated whenever the following conditions are met:
 - (i) The owner attests that corrective measures have been implemented, which may include one or more of the following:
 - (A) Test and slaughter infected birds based on blood tests of every bird in the flock, with either pullorum antigen or by a federally licensed Salmonella enteritidis enzyme-linked immunosorbent assay (ELISA) test when the flock is more than 4 months of age.
 - (B) Perform other corrective actions including, but not limited to, vaccination, medication, cleaning and disinfection of houses, rodent control, and movement of uninfected birds to premises that have been determined to be environmentally negative for S. enteritidis in accordance with par 147 of this subchapter.
 - (C) One hundred percent of blood samples from the birds moved to the clean premises are tested negative for Salmonella pullorum and group D Salmonella. All birds with positive or inconclusive reactions, up to a maximum of 25 birds, shall be submitted to an authorized laboratory and examined for the presence of group D Salmonella, in accordance with part 147 of this subchapter. Cultures from positive samples shall be serotyped.
 - (D) Two consecutive environmental drag swabs taken at the clean premises collected in accordance with part 147 of this subchapter 4 weeks apart are negative for S. enteritidis.
 - (E) Other corrective measures at the discretion of the Official State Agency.
 - (ii) Following reinstatement, a flock will remain eligible for this classification if the flock is tested in accordance with paragraph (e)(1)(v) of this section every 30 days and no positive samples are found and the flock meets the requirements set forth in §145.83(e).

Standard B—Bacteriological Examination Procedure

(1) Reserved

- ~~(1) Laboratory procedure recommended for the bacteriological examination of egg-type and meat-type breeding flocks with salmonella enteritidis positive environments. Birds selected for bacteriological examination from egg-type and meat-type breeding flocks positive for Salmonella enteritidis after environmental monitoring should be~~

examined as described in Section B(2)(a) of these Program Standards, with the following exceptions and modifications allowed due to the high number of birds required for examination:

(a) Except when visibly pathological tissues are present, direct culture, Section B(2)(a)(1) of these standards, may be omitted.

(2) Laboratory procedure recommended for the bacteriological examination of Salmonella from birds

(a) For egg- and meat-type chickens, turkeys, waterfowl, exhibition poultry, and game birds

~~All reactors to the pullorum-typhoid tests, up to 25 birds, and birds from Salmonella enteritidis (SE) positive environments should be cultured in accordance with both the direct enrichment (paragraph (a)(1)) and selective enrichment (paragraph (a)(2)) procedures described in this section: Provided, if there are more than four reactors to the pullorum-typhoid tests in the flock, a minimum of four reactors as provided for in 9 CFR 145.14(a)(6)(ii) shall be submitted to the authorized laboratory for bacteriological examination. Careful aseptic technique should be used when collecting all tissue samples.~~

For reactors to the pullorum-typhoid tests, if there are more than four reactors in a flock, a minimum of four reactors shall be submitted to the authorized laboratory; if the flock has four or fewer reactors all the reactors must be submitted [145.14(a)(6)(ii)]. The isolation of S. Enteritidis from U.S. S. Enteritidis Clean flocks will result in the submission of 60 birds from multiplier egg-type chicken breeding flocks [145.23(d)(2)] or primary egg-type chicken breeding flocks [145.73(d)(2)] and 25 birds from primary meat-type chicken breeding flocks [145.83(e)(3)]. These birds should be cultured in accordance with both direct culture (paragraph (a)(1)) and selective enrichment (paragraph (a)(2)) procedures described in this section. *Provided*, if there are no grossly abnormal or diseased tissues present, direct culture may be omitted. Careful aseptic technique should be used when collecting all tissue samples.

Reason:

The pullorum-typhoid (PT) agglutination test was added to the NPIP for testing egg-type breeder flocks in the late 1980's. It first shows up in the "white book" dated August 1989 under U.S. Sanitation Monitored. When the Salmonella Enteritidis (SE) outbreak in humans associated with eggs was identified in the late 1980's there was a need to identify infected flocks that may produce contaminated eggs. Since SE is a serogroup D1 Salmonella as is S. Pullorum and S. Gallinarum, it was assumed (hoped) that the PT agglutination test would also detect SE infected flocks. Over time, results have shown that the PT agglutination test is not an effective method for the detection of SE infected flocks.

This proposal only removes the PT agglutination test from the U.S. S. Enteritidis Clean classification. It DOES NOT remove it from the U.S. Pullorum-Typhoid Clean classification. In addition to the fact that the PT agglutination test is not an effective method for detecting SE infected flocks, it causes problems for companies that are vaccinating for *Salmonella*. Also, this change resolves the confusion in Standard B(2)(a) over how many reactors to submit for culture.

Sponsor:

Dr. Doug Waltman
Georgia Poultry Laboratory Network

Proposal No. 21 (Amended by TC)

Delegates: 145 H

§145.83 Terminology and classification; flocks and products.

(h) U.S. Newcastle Disease ~~Virus~~ Clean. This program is intended to be the basis from which the breeding-hatchery industry may conduct a program for the prevention and control of Newcastle Disease. It is intended to determine the presence of Newcastle Disease ~~Virus~~ in primary breeding chickens through vaccination and monitoring of each participating breeding flock. A flock and the hatching eggs and chicks produced from it will qualify for this classification when the Official State Agency determines that they have met the following requirements:

(1) It is a primary breeding flock that is either:

(i) Vaccinated for Newcastle Disease ~~Virus~~ using USDA-licensed approved vaccines and response to vaccination is serologically monitored using an approved test as described in §145.14 when more than 4 months of age and meets the criteria in §145.83(h)(2) to retain classification.

OR

(ii) Unvaccinated for Newcastle Disease ~~Virus~~ in which a minimum of 30 birds have tested negative to ND using an approved test as described in §145.14 when more than 4 months of age and meets criteria in §145.83(h)(3) to retain classification.

(2) To retain this classification, for vaccinated flocks,

(i) Vaccines for ND~~V~~ must be USDA-licensed approved vaccines ~~manufactured with low-virulence live strains administered~~ during early stages of development ~~up to grow-out through rearing~~, and ~~killed-inactivated~~ vaccines as final vaccination ~~no later than 6 weeks~~ prior to the onset of egg production

AND

(ii) The flock has been monitored for antibody response using approved serological tests as listed in §145.14 and the results are compatible with immunological response against ND vaccination

AND

(iii) Testing must include a minimum of 30 birds with a serologic monitoring program ~~beginning at approximately 10 weeks when more than 4 months~~ of age ~~and prior to the onset of production~~, and not longer than every 90 days thereafter.

(3) To retain this classification for unvaccinated flocks,

(i) A minimum of 30 birds per flock must ~~be~~ test negative using an approved test in §145.14 at intervals of 90 days

OR

(ii) A sample of fewer than 30 birds may be tested, and found negative, at any one time if all pens are equally represented and a total of 30 birds is tested within each 90-day period;

AND

(iii) During each 90-day period, all primary spent fowl, up to a maximum of 30, must test negative to ND within 21 days prior to movement to slaughter.

(4) Newcastle Disease ~~Virus~~ must be a disease reportable to the responsible State authority (State veterinarian, etc.) by all licensed veterinarians. To accomplish this, all laboratories (private, State, and university laboratories) that perform diagnostic procedures on poultry must examine all submitted cases of unexplained respiratory disease, egg production drops, and mortality for ND~~V~~.

§145.84 Terminology and classification; compartments

(a) U.S. Avian Influenza and NDV Clean Compartment

This program is intended to be the basis from which the primary meat-type chicken breeding-hatchery industry may demonstrate the existence and implementation of a program that has been approved by the Official State Agency and the Service to establish a compartment consisting of a primary breeding-hatchery company that is free of H5/H7 avian influenza (AI) and NDV. This compartment has the purpose of protecting the defined subpopulation and avoiding the introduction and spread of H5/H7 AI and NDV within that subpopulation by prohibiting contact with other commercial poultry operations, other domestic and wild birds, and other intensive animal operations. The program shall consist of the following:

(1) *Definition of the compartment.* Based on the guidelines established by the World Organization for Animal Health (OIE) in the Terrestrial Animal Health Code and the guidelines in this paragraph (a), the primary breeder company will define the compartment with respect to H5/H7 AI and NDV. Specifically, the company will use a comprehensive biosecurity program to define the compartment as a subpopulation of poultry with a health status for H5/H7 AI and NDV that is separate from birds and poultry outside the compartment. The Official State Agency and the Service must first approve all documentation submitted by the company to substantiate the defined compartment as adequate to qualify for epidemiological separation from other potential sources of infection of H5/H7 AI and NDV. Guidelines for the definition of the compartment include:

(i) Definition and description of the subpopulation of birds and their health status. All birds included in the compartment must be U.S. Avian Influenza Clean in accordance with §145.83(g) and NDV Clean in accordance with §145.83(h). The poultry must also be located in a State that has an initial State response and containment plan approved by APHIS under §56.10 of this chapter and that participates in the diagnostic surveillance program for H5/H7 low pathogenicity AI as described in §145.15. Within the compartment, all official tests for AI and NDV, as described in §145.14(d) and §145.14(e), must be conducted in State or Federal laboratories or in NPIP authorized laboratories that meet the minimum standards described in §147.52 of this subchapter. In addition, the company must provide to the Service upon request any relevant historical and current H5/H7 AI and NDV-related data for reference regarding surveillance for the disease and the health status of the compartment. Upon request, the Official State Agency may provide such data for other commercial poultry populations located in the State.

(ii) Description of animal identification and traceability processes. The primary breeder company must also include a description of its animal identification and traceability records, including examples of Veterinary Services (VS) Form 9-5, "Report of Hatcheries, Dealers and Independent Flocks"; VS Form 9-2, "Flock Selection and Testing Report"; VS Form 9-3, "Report of Sales of Hatching Eggs, Chicks and Poults"; VS Form 9-9, "Hatchery Inspection Report"; set and hatch records; egg receipts; and egg/chick invoices for the subpopulation. Documentation must also include breed identification (NPIP stock code). The Service should ensure that an effective flock identification system and traceability system are in place.

(iii) Definition and description of the physical components or establishments of the defined compartment. The primary breeder company must provide documentation establishing that the defined compartment is epidemiologically separated from other poultry and bird populations. The documentation must be approved by the Official State Agency and the Service as indicating adequate epidemiological separation to maintain the compartment's separate health status

with respect to H5/H7 AI and NDV. The documentation should include descriptions of:

- (A) The physical and spatial factors that separate the compartment from surrounding bird populations and affect the biosecurity status of the compartment.
- (B) Relevant environmental factors that may affect exposure of the birds to AI and NDV.
- (C) The functional boundary and fencing that are used to control access to the compartment.
- (D) Facilities and procedures to prevent access by wild birds and to provide separation from other relevant hosts.
- (E) The relevant infrastructural factors that may affect exposure to AI and NDV, including the construction and design of buildings or physical components, cleaning and disinfection of buildings and physical components between production groups with quality assurance verification, cleaning and disinfection of equipment, and introduction of equipment or material into the compartment.

(iv) Definition and description of the functional relationships between components of the defined compartment. Functional relationships between components of the compartment include traffic movement and flow at and among premises, personnel movement at and among premises, exposure to live bird populations, and any other factors that could affect biosecurity of the compartment. All physical components of the compartment must be maintained in compliance with hygiene and biosecurity procedures for poultry primary breeding flocks and hatcheries in accordance with part 147 of this subchapter. In addition, the company must provide a biosecurity plan for the compartment and all included components. The biosecurity plan should include but not be limited to:

- (A) Requirements that company employees and contract growers limit their contact with live birds outside the compartment.
- (B) An education and training program for company employees and contractors.
- (C) Standard operating procedures for company employees, contractors, and outside maintenance personnel.
- (D) Requirements for company employees and non-company personnel who visit any premises within the compartment.
- (E) Company veterinary infrastructure to ensure flock monitoring and disease diagnosis and control measures.
- (F) Policies for management of vehicles and equipment used within the compartment to connect the various premises.
- (G) Farm site requirements (location, layout, and construction).
- (H) Pest management program.
- (I) Cleaning and disinfection process.
- (J) Requirements for litter and dead bird removal and/or disposal.

(v) Description of other factors important for maintaining the compartment. The company veterinary infrastructure will assess sanitary measures, environmental risk factors, and management and husbandry practices that relate to the separation of the compartment and the health status of the birds contained within the compartment that may affect risk of exposure to H5/H7 AI and NDV. This assessment must include a description of internal monitoring and auditing systems (e.g., quality assurance and quality control programs) to demonstrate the effectiveness of the compartment. Upon request, the Service will provide the company with information on the epidemiology of H5/H7 AI and NDV and the

associated risk pathways in which the components of the compartment are located.

(vi) Approval or denial. Based on the documentation provided under this paragraph (a)(1), as well as any other information the Service and the Official State Agency determine to be necessary, the Service and the Official State Agency will approve or deny the classification of the compartment as U.S. Avian Influenza and NDV Clean.

(2) *Company activities for maintenance of the compartment.*

(i) The primary breeder company's management of biosecurity, surveillance, and disease control efforts must be uniform and equivalent among all components that are a part of the compartment. Oversight and inspection of these management practices must be conducted by the company's licensed, accredited veterinarians.

(ii) Veterinary staff from the Official State Agency and NPIP staff will work in partnership with licensed, accredited veterinarians to train and certify auditors through Service-approved workshops. The trained auditors will conduct biosecurity and operational audits at least once every 2 years to ensure the integrity of the compartment. These audits will include evaluation of the critical control points and standard operating practices within the compartment, verification of the health status of the flock(s) contained within the compartment, and examination of the biosecurity and management system of the integrated components of the compartment.

(iii) In addition, the company must demonstrate compliance with paragraph (a)(1) of this section for remaining in the U.S. Avian Influenza and NDV Clean classifications, surveillance for H5/H7 AI and NDV within the compartment, and conducting tests in State or Federal laboratories or in NPIP authorized laboratories. Accredited veterinarians are responsible for the enforcement of active and passive surveillance of H5/H7 AI and NDV in primary breeder flocks. Baseline health status must be maintained for all flocks or subpopulations within the compartment, indicating the dates and negative results of all avian influenza and NDV surveillance and monitoring testing, the dates and history of last disease occurrence (if any), the number of outbreaks, and the methods of disease control that were applied.

(iv) Documentation will be maintained in the company's database and will be verified as required by the Service and/or the Official State Agency.

(3) *Service and Official State Agency activities for maintenance of the compartment.* The Service will work in cooperation with the Official State Agencies to ensure the continued integrity of any recognized compartments. Activities include:

(i) Oversight of the establishment and management of compartments;

(ii) Establishment of effective partnerships between the Service, the Plan, and the primary breeder industry;

(iii) Approval or denial of classification of compartments as U.S. Avian Influenza and NDV Clean Compartments under paragraph (a)(1) of this section;

(iv) Official certification of the health status of the compartment, and commodities that may be traded from it through participation in the Plan for avian diseases, including the U.S. Avian Influenza Clean program as described in §145.83(g) and NDV Clean Program as described in §145.83(h) and diagnostic surveillance for H5/H7 low pathogenicity AI as described in §145.15;

(v) Conducting audits of compartments at least once every 2 years to:

(A) Confirm that the primary breeding company's establishments are epidemiologically distinct and pathways for the introduction of disease into the compartment are closed through routine operational procedures;

and

(B) Evaluate and assess the management and husbandry practices relating to biosecurity to determine whether they are in compliance with hygiene and biosecurity procedures for poultry primary breeding flocks and hatcheries in accordance with part 147 of this subchapter;

(vi) Providing, upon request, model plans for management and husbandry practices relating to biosecurity in accordance with part 147 of this subchapter, risk evaluations in conjunction with the primary breeder industry (including disease surveillance such as VS Form 9-4, “Summary of Breeding Flock Participation”), and diagnostic capability summaries and systems for initial State response and containment plans in accordance with §56.10 of this chapter; and
(vii) Publicizing and sharing compartment information with international trading partners, upon request, to establish approval and recognition of the compartment, including timeliness and accuracy of disease reporting and surveillance measures as described in §§145.15, ~~and 145.83(g)~~, and 145.83(h).

(4) *Emergency response and notification.* In the case of a confirmed positive of H5/H7 AI and/or NDV in the subpopulation of the compartment, the management of the compartment must notify the Service. The Service will immediately suspend the status of the compartment. A compartment would be eligible to resume trade with importing countries only after the compartment has adopted the necessary measures to reestablish the biosecurity level and confirm that H5/H7 AI and/or NDV is not present in the compartment and the Service has reevaluated the management and biosecurity measures of the compartment and approved said compartment for trade.

Reason: The Primary Breeders propose the addition of an NDV Clean program. See corresponding proposal in Program Standards Subpart F.

Sponsor: Primary Breeder Association
Dr. Elena Behnke, Aviagen
Dr. Alberto Torres, Cobb-Vantress
Dr. Travis Schaal, Hy-Line
Dr. Dustin Burch, Aviagen Turkeys

Proposal No. 22

Delegates: 146 B

§146.23 Terminology and classification; flocks and products.

Participating flocks which have met the respective requirements specified in this section may be designated by the following terms and the corresponding designs illustrated in §146.9 of this part:

(a) U.S. H5/H7 Avian Influenza Monitored-(1) *Table-egg layer pullet flocks*. This program is intended to be the basis from which the table-egg layer industry may conduct a program to monitor for the H5/H7 subtypes of avian influenza. It is intended to determine the presence of the H5/H7 subtypes of avian influenza in table-egg layer pullets through routine surveillance of each participating commercial table-egg layer pullet flock. A flock will qualify for this classification when the Official State Agency determines that it has met one of the following requirements:

(i) It is a commercial table-egg layer pullet flock in which a minimum of 11 birds have been tested negative to the H5/H7 subtypes of avian influenza as provided in §146.13(b) within 21 days prior to movement; or

(ii) It is a commercial table-egg layer pullet flock that has an ongoing active and diagnostic surveillance program for the H5/H7 subtypes of avian influenza in which the number of birds tested is equivalent to the number required in paragraph (a)(1)(i) of this section and that is approved by the Official State Agency and the Service.

(iii) Pullet flocks of 75,000 or more birds located on a participating table egg layer premises may be enrolled under the same NPIP number as the layer flock;

(iv) If a premises owner grows pullets for multiple owners, and has the housing capacity of 75,000 or more pullets, that grower should be enrolled as the NPIP participant and an NPIP number should be assigned to that grower;

Reason: The additional language regarding enrollment of pullet flocks would help reduce confusion for participants.

Sponsor: Ron Ballew, Hillandale-Gettysburg LP
Nan Hanshaw, PA Dept. of Agriculture

Proposal No. 23

Delegates: Combined

§147.45 Official delegates.

Each cooperating State shall be entitled to one official delegate for each of the programs prescribed in parts 145 and 146 of this chapter in which it has one or more participants at the time of the Conference. The official delegates shall be elected by a representative group of participating industry members and be certified by the Official State Agency. It is recommended but not required that the official delegates be Plan participants. Individuals may be allowed to be an official delegate or alternate delegate for up to three States in which that delegate has flocks or is a plan participant with acknowledgement and approval of the Official State Agencies. Each official delegate shall endeavor to obtain, prior to the Conference, the recommendations of industry members of his State with respect to each proposed change.

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Reason: This proposal is to provide a more clear allowance for representation of a single delegate in multiple states in which that delegate has participation. For the 2016 NPIP Biennial meeting several participating companies with operations in several states were contacted by multiple states to represent their subsections. Several industry individuals obtained approval from the respective OSA's in each state to have that one individual vote for the applicable subpart in each of the states thus having multiple votes. However it was disallowed based on unknown reasons not contained in the above CFR section in 147.45 stating that multiple votes for multiple states by a single person was not allowed. This is contradictory to what already occurs routinely within a state with little participation in which an OSA holds multiple votes for subsections with no industry participants in attendance. The concern expressed by some is that they don't want to get in a situation where company X with operations in multiple states "controls votes" with a single person. Although this is understandable, all that company X has to do is send additional people to each individually vote for each separate state. All that this does is increase the costs to both the company and NPIP for dinners and meeting space arrangements associated with getting these additional voting members to the Biennial meeting. Ultimately it will still be up to the OSA's to make sure they get the right people that will accurately represent their entire industry/subpart fairly no matter if multiple votes is allowed or not. The last sentence of section 147.45 states this requirement for the delegate to seek out the recommendations of industry members of that state prior to the meeting. For some subsections, for example turkey breeders, there just aren't a lot of choices for many states for delegate participation from different companies due to consolidation, so multi-state representation by a single individual would be a justifiable allowance and is often requested by the OSA's of that state. The proposed wording above also provides the restriction such that a delegate is only allowed to represent multiple states if they have flocks or participation in that state as to not allow nefarious representation in states with no active stake or participation in by that delegate.

Sponsors: Ben Wileman, Select Genetics, Willmar, MN
Kabel Robbins, Butterball, Ozark, AR
Brian Wooming, Cargill, Springdale, AR
Bill Pittenger, Missouri Department of Agriculture, Jefferson City, MO
Katie Schlist, Forsman Farms, Howard Lake, MN
Rosemary Marusak, Daybreak Foods, Lake Mills, WI
Travis Schaal, Hy-Line International, Dallas Center, IA
Carrie Cremers, Pilgrim's, Cold Spring, MN

Proposal No. 24

Delegates: Combined

§147.52 Authorized laboratories.

These minimum requirements are intended to be the basis on which an authorized laboratory of the Plan can be evaluated to ensure that official Plan assays are performed in accordance with the NPIP Program Standards or other procedures approved by the Administrator in accordance with §147.53(d)(1) and reported as described in paragraph (f) of this section. A satisfactory evaluation will result in the laboratory being recognized by the NPIP office of the Service as an authorized laboratory qualified to perform the assays provided for in this part.

(a) *Check-test proficiency.* The NPIP will serve as the lead agency for the coordination of available check tests from the National Veterinary Services Laboratories. Further, the NPIP may approve and authorize additional laboratories to produce and distribute a check test as needed.

The authorized laboratory must use the next available check test for each assay that it performs.

(b) *Trained technicians.* ~~The~~ Testing procedures at the all authorized laboratory laboratories must be run or overseen by a laboratory technician who every 4 years has attended, and satisfactorily completed, Service-approved laboratory workshops for Plan-specific diseases, ~~within the past 4 years.~~

Reason: By removing the word “within” and substituting “every”, the proposed change to paragraph (b) clears up any confusion that might arise as to the timing of when a laboratory technician is required to attend a Service-approved laboratory workshop. This change makes clear that a workshop for an individual Plan disease must be attended by a laboratory technician at 4-year intervals, rather than any time during a 4-year span. Additionally, including the word “authorized” allows for consistency with other paragraphs in the section.

Sponsor: Monica Della Maggiore
California Poultry Federation

Late Proposal No. 1

Delegates: Combined

PART 56—CONTROL OF H5/H7 LOW PATHOGENIC AVIAN INFLUENZA

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§56.1 Definitions.

Cleaning. The removal of gross contamination, organic material, and debris from the premises or respective structures, via mechanical means like sweeping (dry cleaning) and/or the use of water and soap or detergent (wet cleaning). The goal is to minimize organic material to prepare for effective disinfection.

Compensation. In the case of AI infection, compensation specifically refers to reimbursement for the activities associated with the depopulation of infected or exposed poultry, including the disposal of contaminated carcasses and materials and the cleaning and disinfection of premises, conveyances, and materials that came into contact with infected or exposed poultry. In the case of contaminated materials, if the cost of cleaning and disinfection would exceed the value of the materials, or cleaning and disinfection would be impracticable for any reason, VS will base compensation on the fair market value (depreciated value) of those materials. Compensation does NOT include payment for depopulated birds or eggs destroyed (see definition of Indemnity).

Disinfection. Methods used on surfaces to destroy or eliminate Avian Influenza through physical (e.g., heat) or chemical (e.g., disinfectant) means. A combination of methods may be required.

Indemnity. AI indemnity specifically refers to payments representing the fair market value of destroyed birds and eggs. AI indemnity does not include reimbursements for depopulation, disposal, destroyed materials, or cleaning and disinfection (virus elimination) activities; these are covered under compensation.

Virus Elimination (VE): Cleaning and disinfection measures primarily conducted to destroy or eliminate all AI virus on the premises in a cost-effective manner.

§56.2 Cooperation with States

(a) The Administrator has been delegated the authority to cooperate with Cooperating State Agencies in the eradication of H5/H7 LPAI. This cooperation may include, but is not necessarily limited to, the following activities:

- (1) Payment to Cooperating State Agencies for surveillance and monitoring associated with poultry that have been infected with or exposed to H5/H7 LPAI;
 - (2) Transfer of vaccine for H5/H7 LPAI to Cooperating State Agencies if provided for in the initial State response and containment plan approved by APHIS under §56.10; and
 - (3) Payment for vaccine administration by Cooperating State Agencies, if provided for in the initial State response and containment plan approved by APHIS under §56.10.
- (b) (1) Any payment made to a State or an Official State Agency for the activities listed in paragraphs (a)(1) and (a)(3) of this section must be made through a cooperative agreement between the Cooperating State Agency and APHIS. The payment for which the Cooperating State Agency is eligible will be determined in the cooperative agreement.

(i) For any Cooperating State Agency that participates in the National Poultry Improvement Plan diagnostic surveillance program for H5/H7 LPAI, as described in §146.14 of this chapter, and has an initial State response and containment plan for H5/H7 LPAI that is approved by APHIS, as described in §56.10 of this part, the cooperative agreement will provide that the Cooperating State Agency is eligible for payment of ~~100~~ percent of the costs of surveillance and monitoring and ~~100~~ percent of the costs of vaccine administration, as determined in the cooperative agreement.

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(ii) For any Cooperating State Agency that does not meet the criteria in paragraph (b)(1)(i) of this section, the cooperative agreement will provide that the Cooperating State Agency is eligible for payment of ~~25~~ percent of the costs of surveillance and monitoring and ~~25~~ percent of the costs of vaccine administration, as determined in the cooperative agreement.

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(2) Transfer of vaccine under paragraph (a)(2) of this section must be accomplished through a cooperative agreement between the Cooperating State Agency and APHIS.

(c) Cooperating State Agencies will be responsible for making the determination to request Federal assistance under this part in the event of an outbreak of H5/H7 LPAI.

§56.3 Payment of indemnity and/or compensation

(a) *Activities eligible for indemnity and/or compensation.* The Administrator ~~shall pay~~ indemnity and/or compensation for the activities listed in paragraphs (a)(1) through (a)(3) of this section, as provided in paragraph (b) of this section:

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- (1) Destruction and disposal of poultry that were infected with or exposed to H5/H7 LPAI;
- (2) Destruction of any eggs destroyed during testing of poultry for H5/H7 LPAI during an outbreak of H5/H7 LPAI; and
- (3) Cleaning and disinfection of premises, conveyances, and materials that came into contact with poultry that were infected with or exposed to H5/H7 LPAI; or, in the case of materials, if the cost of cleaning and disinfection would exceed the value of the materials or cleaning and disinfection would be impracticable for any reason, the destruction and disposal of the materials.

(b) *Percentage of costs eligible for indemnity and/or compensation.*

Except for poultry that are described by the categories in paragraphs (b)(1) through (b)(3) of this section, the Administrator is authorized to pay 100 percent of the costs, as determined in accordance with §56.4, of the activities described in paragraphs (a)(1) through (a)(3) of this section, regardless of whether the infected or exposed poultry participate in the Plan. For infected or exposed poultry that are described by the categories in paragraphs (b)(1) through (b)(3) of this section, the Administrator is authorized to pay 25 percent of the costs of the activities described in paragraphs (a)(1) through (a)(3) of this section:

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- (1) The poultry are from a breeding flock that participates in any Plan program in part 145 of this chapter but that does not participate in the U.S. Avian Influenza Clean or the U.S. H5/H7 Avian Influenza Clean program of the Plan available to the flock in part 145 of this chapter; or
- (2) The poultry are from a commercial flock or slaughter plant, but the flock or slaughter plant does not participate in the U.S. Avian Influenza Monitored program available to the commercial flock or slaughter plant in part 146 of this chapter; or
- (3) The poultry are located in a State that does not participate in the diagnostic surveillance program for H5/H7 LPAI, as described in §146.14 of this chapter, or that does not have an initial State response and containment plan for H5/H7 LPAI that is approved by APHIS under §56.10, unless such poultry participate in the Plan with another State that does participate in the diagnostic surveillance program for H5/H7 LPAI, as described in §146.14 of this chapter, and has an initial State response and containment plan for H5/H7 LPAI that is approved by APHIS under §56.10.

(c) *Other sources of payment.* If the recipient of indemnity and/or compensation for any of the activities listed in paragraphs (a)(1) through (a)(3) of this section also receives payment for any of those activities from a State or from other sources, the indemnity and/or compensation provided under this part ~~will~~ may be reduced by the total amount of payment received from the State or other sources to the extent that total payments do not exceed 100% of total reimbursable indemnity and/or compensation amounts.

§56.4 Determination of indemnity and/or compensation amounts

(a) *Destruction and disposal of poultry.*

- (1) Indemnity for the destruction of poultry and/or eggs infected with or exposed to H5/H7 LPAI will be based on the fair market value of the poultry and/or eggs, as determined by an appraisal. ~~Poultry infected with or exposed to H5/H7 LPAI that are removed by APHIS or a Cooperating State Agency from a flock will be appraised by an APHIS official appraiser and a State official appraiser jointly, or, if APHIS and State authorities agree, by either an APHIS official appraiser or a State official appraiser alone. For laying hens, the appraised value should include the hen's projected future egg production. Appraisals of poultry must be reported on forms furnished by APHIS and signed by the appraisers and must be signed by the owners of the poultry to indicate agreement with the appraisal amount.~~
 - i. For commercial poultry and most other flocks, these values will be determined using the current APHIS appraisal calculator values. For laying hens, the appraisal calculator values include the hen's projected future egg production.
 - ii. For specialty birds or other flocks for which a calculator is not available, the Avian Health staff will contact the compensation specialist at the Centers for Epidemiology and Animal Health to establish an appropriate value for the birds and/or eggs.

- iii. An indemnity request form Appraisals of poultry must be signed by the owners and grower (if applicable) of the poultry and received by APHIS prior to the destruction of the poultry and eggs, unless the owners, grower, APHIS, and the Cooperating State Agency agree in writing that the poultry may be destroyed immediately. Reports of appraisals must show the number of birds and the value per head. Complete inventory records of all birds and eggs on the premises must be provided to APHIS prior to the start of depopulation.
- (2) Indemnity Compensation for disposal of poultry and/or eggs infected with or exposed to H5/H7 LPAI will be based on receipts or other documentation maintained by the claimant verifying expenditures for disposal activities authorized by this part. Any disposal of poultry infected with or exposed to H5/H7 LPAI for which indemnity compensation is requested must be performed under a compliance agreement between the claimant, ~~the Cooperating State Agency~~, and APHIS. APHIS will review claims for indemnity compensation for disposal to ensure that all expenditures relate directly to activities described in §56.5 and in the initial State response and containment plan described in §56.10. If disposal is performed by the Cooperating State Agency, APHIS will indemnify compensate the Cooperating State Agency for disposal under a cooperative agreement.
- (3) The destruction and disposal of the indemnified poultry and/or eggs must be conducted in accordance with the initial State response and containment plan for H5/H7 LPAI, as described in §56.10.
- ~~(b) Destruction of eggs. Indemnity for eggs destroyed during an outbreak for testing for H5/H7 LPAI will be based on the fair market value of the eggs, as determined by an appraisal. Eggs destroyed for testing for H5/H7 LPAI will be appraised by an APHIS official appraiser and a State official appraiser jointly, or, if APHIS and State authorities agree, by either an APHIS official appraiser or a State official appraiser alone. Appraisals of eggs must be reported on forms furnished by APHIS and signed by the appraisers and must be signed by the owners of the eggs to indicate agreement with the appraisal amount. Appraisals of eggs must be signed by the owners of the eggs prior to the destruction of the poultry, unless the owners, APHIS, and the Cooperating State Agency agree that the eggs may be destroyed immediately. Reports of appraisals must show the number of eggs and the value per egg.~~
- ~~(e) Cleaning and disinfection (virus elimination).~~
- (1) Indemnity Compensation for cleaning and disinfection (virus elimination) of premises, conveyances, and materials that came into contact with poultry that are infected with or exposed to H5/H7 LPAI will be based on receipts or other documentation maintained by the claimant verifying expenditures for cleaning and disinfection activities authorized by this part determined using the current APHIS flat-rate virus elimination calculator in effect at the time of the infection. Any cleaning and disinfection of premises, conveyances, and materials for which indemnity is requested must be performed under a compliance agreement between the claimant, the Cooperating State Agency, and APHIS. APHIS will review claims for indemnity for cleaning and disinfection to ensure that all expenditures relate directly to activities described in §56.5 and in the initial State response and containment plan described in §56.10.
- (2) For premises types for which a flat-rate VE calculator is not available, reimbursement will be based on receipts or other documentation maintained by the claimant verifying expenditures for cleaning and disinfection (virus elimination) activities authorized by this part. Any cleaning and disinfection (virus elimination) of premises, conveyances, and materials for which compensation is requested must be performed under a compliance agreement between the claimant, the Cooperating State Agency, and APHIS. APHIS will

review claims for compensation for cleaning and disinfection (virus elimination) to ensure that all expenditures relate directly to activities described in §56.5 and in the initial State response and containment plan described in §56.10. In the case of materials, if the cost of cleaning and disinfection would exceed the value of the materials or cleaning and disinfection would be impracticable for any reason, indemnity for the destruction of the materials will be based on the fair market value of those materials, as determined by an appraisal. Materials will be appraised by an APHIS official appraiser and a State official appraiser jointly, or, if APHIS and State authorities agree, by either an APHIS official appraiser or a State official appraiser alone. Indemnity for disposal of the materials will be based on receipts or other documentation maintained by the claimant verifying expenditures for disposal activities authorized by this part. Appraisals of materials must be reported on forms furnished by APHIS and signed by the appraisers and must be signed by the owners of the materials to indicate agreement with the appraisal amount. Appraisals of materials must be signed prior to the destruction of the materials, unless the owners, APHIS, and the Cooperating State Agency agree that the materials may be destroyed immediately. Any disposal of materials for which indemnity is requested must be performed under a compliance agreement between the claimant, the Cooperating State Agency, and APHIS. APHIS will review claims for compensation for disposal to ensure that all expenditures relate directly to activities described in §56.5 and in the initial State response and containment plan described in §56.10.

(i) In the case of materials, if the cost of cleaning and disinfection (virus elimination) would exceed the value of the materials or cleaning and disinfection (virus elimination) would be impracticable for any reason, compensation for the destruction of the materials will be based on the fair market value (depreciated value) of those materials, as determined by an appraisal. Materials will be appraised by an APHIS official appraiser. Compensation for disposal of the materials will be based on receipts or other documentation maintained by the claimant verifying expenditures for disposal activities authorized by this part. Appraisals of materials must be reported on forms furnished by APHIS and must be signed by the appraisers and by the owners of the materials to indicate agreement with the appraisal amount. Appraisals of materials must be signed and received by APHIS prior to the disassembly or destruction of the materials, unless the owners, APHIS, and the Cooperating State Agency agree in writing that the materials may be disassembled and/or destroyed immediately. Any disposal of materials for which compensation is requested must be performed under a compliance agreement between the claimant, the Cooperating State Agency, and APHIS. APHIS will review claims for compensation for disposal to ensure that all expenditures relate directly to activities described in §56.5 and in the initial State response and containment plan described in §56.10.

(d) Requirements for compliance agreements. The compliance agreement is a comprehensive document that describes the depopulation, disposal, and cleaning and disinfection plans for poultry that were infected with or exposed to H5/H7 LPAI, or a premises that contained such poultry. The compliance agreement sets out ~~APHIS responsibilities, owner responsibilities, and Cooperating State Agency responsibilities~~ cost estimates that include labor, materials, supplies, equipment, personal protective equipment, and any additional information deemed necessary by APHIS. A compliance agreement is comparable to a statement of work and indicates what tasks will be completed, who will be responsible for each task, and how much the work is expected to cost. A compliance agreement may also be referred to as a detailed financial plan. The compliance agreement must include the owner's name and the name and address of the

~~affected premises. The compliance agreement must have signatories that include, but are not necessarily limited to, the owner, the grower (if applicable), the Cooperating State Agency representative, the State veterinarian, and the APHIS area supervisor. In addition, the compliance agreement must contain a flock plan with estimated cost breakdowns that include labor, materials, personal protective equipment, travel expenses for personnel involved, and any additional information deemed necessary by the Service. A signed compliance agreement is required before beginning any work for which indemnity funds will be requested. Once work associated with the compliance agreement is completed, receipts and documentation detailing the activities specified in the agreement should be forwarded to APHIS for review, approval, and final payment. This documentation should be submitted to APHIS no later than 30 days after the quarantine release of the affected or exposed premises.~~

§56.5 Destruction and disposal of poultry and cleaning and disinfection (virus elimination) of premises, conveyances, and materials

(a) *Destruction of poultry.* Poultry that are infected with or exposed to H5/H7 LPAI may be required to be destroyed at the discretion of the Cooperating State Agency and APHIS and in accordance with the initial State response and containment plan described in §56.10. The Cooperating State Agency and APHIS will select a method to use for the destruction of such poultry based on the following factors:

- (1) The species, size, and number of the poultry to be destroyed;
- (2) The environment in which the poultry are maintained;
- (3) The risk to human health or safety of the method used;
- (4) Whether the method requires specialized equipment or training;
- (5) The risk that the method poses of spreading the H5/H7 LPAI virus;
- (6) Any hazard the method could pose to the environment;
- (7) The degree of bird control and restraint required to administer the destruction method;
- (8) The speed with which destruction must be conducted; and
- (9) Consistency of the method with humane euthanasia guidelines.

(b) *Disposal of poultry.* Carcasses of poultry that have died from H5/H7 LPAI infection or poultry that have been humanely slaughtered to fulfill depopulation requirements must be disposed of promptly and efficiently in accordance with the initial State response and containment plan described in §56.10 to prevent the spread of H5/H7 LPAI infection. Disposal methods will be selected by the Cooperating State Agency and APHIS and may include one or more of the following: Burial, incineration, composting, or rendering. Regardless of the method used, strict biosecurity procedures must be implemented and enforced for all personnel and vehicular movement into and out of the area in accordance with the initial State response and containment plan to prevent dissemination of the H5/H7 LPAI virus.

(c) *Controlled marketing.*

(1) At the discretion of the Cooperating State Agency and APHIS, poultry that has been infected with or exposed to H5/H7 LPAI may be allowed to move for controlled marketing and maintain their current NPIP certifications in accordance with the initial State response and containment plan described in §56.10 and in accordance with the following requirements:

- (i) Poultry infected with or exposed to H5/H7 LPAI must not be transported to a market for controlled marketing until approved by the Cooperating State Agency in accordance with the initial State response and containment plan described in §56.10.

(ii) Within 7 days prior to slaughter, each flock to be moved for controlled marketing must be tested for H5/H7 LPAI using a test approved by the Cooperating State Agency and found to be free of the virus.

(iii) Routes to slaughter must avoid other commercial poultry operations whenever possible. All load-out equipment, trailers, and trucks used on premises that have housed poultry that were infected with or exposed to H5/H7 LPAI must be cleaned and disinfected and not enter other poultry premises or facilities for 48 hours after removing such poultry from their premises.

(iv) Flocks moved for controlled marketing must be the last poultry marketed during the week they are marketed.

(2) Poultry moved for controlled marketing will not be eligible for indemnity under §56.3. However, any costs related to cleaning and disinfection (virus elimination) of premises, conveyances, and materials that came into contact with poultry that are moved for controlled marketing will be eligible for indemnity compensation under §56.3.

(d) *Cleaning and disinfection (virus elimination) of premises, conveyances, and materials.*

Premises, conveyances, and materials that came into contact with poultry infected with or exposed to H5/H7 LPAI must be cleaned and disinfected; *Provided*, that materials for which the cost of cleaning and disinfection would exceed the value of the materials or for which cleaning and disinfection would be impracticable for any reason may be destroyed and disposed. Cleaning and disinfection must be performed in accordance with the initial State response and containment plan described in §56.10, which must be approved by APHIS. Cleaning and disinfection must also be performed in accordance with any applicable State and local environmental regulations. ~~This paragraph (d) provides guidelines for the development of a cleaning and disinfection plan for a premises and for the materials and conveyances on that premises.~~

~~(1) *Preparation for cleaning and disinfection.* Following the depopulation or controlled marketing of all poultry infected with or exposed to H5/H7 LPAI on a premises, the following procedures should be completed prior to cleaning and disinfection:~~

~~(i) Secure all feathers and debris that might blow around outside the house in which the infected or exposed poultry were held by gathering and pushing the material into the house;~~

~~(ii) Apply insecticides and rodenticides immediately after the removal of the birds, before the house cools;~~

~~(iii) Close the house in which the poultry were held, maintaining just enough ventilation to remove moisture. Leave the house undisturbed for a minimum of 72 hours.~~

~~(2) *Cleaning and disinfection.* All premises, conveyances, and materials that came into contact with poultry that were infected with or exposed to H5/H7 LPAI must be cleaned and disinfected. Cleaning and disinfection must be performed on all buildings that came into contact with poultry that were infected with or exposed to H5/H7 LPAI within a premises, including pumphouses and service areas. To accomplish cleaning and disinfection, the following procedures should be completed:~~

~~(i) *Disposal of manure, debris, and feed.* Clean up all manure, debris, and feed. Compost manure, debris, and feed by windrowing in the house if possible. If this is not possible, set up a system for hauling manure, debris, and feed to an approved site for burial, piling, or composting. Manure, debris, and feed may be removed from the house or premises and disposed of by composting it on site, leaving it in a undisturbed pile on site, or removing it from the site in covered vehicles. Land application of manure, debris, and feed should only be performed in accordance with the initial State response and containment plan described in §56.10. Clean out the house or move or spread litter as determined by the~~

Cooperating State Agency and in accordance with the initial State response and containment plan. If composting is used as a disposal method, manure and litter should be composted in accordance with State and local regulations. If litter is piled, the litter pile must be covered and allowed to sit undisturbed for an amount of time approved by the Cooperating State Agency and APHIS and in accordance with the initial State response and containment plan described in §56.10. Drying and heat *in situ* over time are effective and may be used in place of composting if weather conditions or conditions in the building are favorable. After use, equipment used to clean out manure, debris, and feed must be cleaned, disinfected, and inspected at the site to which the manure and litter was transported. In the case of inclement weather, the equipment may be cleaned, disinfected, and inspected at off-site wash stations at the discretion of the Cooperating State Agency and APHIS.

(ii) *Cleaning of premises and materials.* Cleaning and washing should be thorough to ensure that all materials or substances contaminated with H5/H7 LPAI virus, especially manure, dried blood, and other organic materials, are removed from all surfaces. Spray all contaminated surfaces above the floor with detergent and water to knock dust down to the floor, using no more water than necessary. Wash equipment and houses with detergent and water. Disassemble equipment as required to clean all contaminated surfaces. Special attention should be given to automatic feeders and other closed areas to ensure adequate cleaning. Inspect houses and equipment to ensure that cleaning has removed all contaminated materials or substances. Rinse with fresh water and let houses and equipment dry completely before applying disinfectant.

(iii) *Disinfection of premises and materials.* When cleaning has been completed and all surfaces are dry, all interior surfaces of the structure should be saturated with a disinfectant registered with the U.S. Environmental Protection Agency (EPA) for AI virus per label instructions or a disinfectant approved by the EPA for use under a Federal Insecticide, Rodenticide, and Fungicide Act section 18 exemption. A power spray unit should be used to spray the disinfectant on all surfaces that may be treated with the disinfectant according to its EPA label, making sure that the disinfectant gets into cracks and crevices. Special attention should be given to automatic feeders and other closed areas to ensure adequate disinfection.

(vi) *Cleaning and disinfection of conveyances.* Clean and disinfect all trucks and vehicles used in transporting affected poultry or materials before soil dries in place. Both exterior, including the undercarriage, and interior surfaces, including truck cabs, must be cleaned. The interior of the truck cabs should be washed with clean water and sponged with a disinfectant authorized in §71.10(a) of this chapter. Manure and litter removed from these vehicles should be handled in a manner similar to that described in paragraph (d)(2)(i) of this section.

(3) *Activities after cleaning and disinfection.* Premises should remain empty until testing provides negative virus detection results and checked by the Cooperating State Agency in accordance with the initial State response and containment plan described in §56.10. The premises may not be restocked with poultry until after the date specified in the initial State response and containment plan described in §56.10.

(4) *Destruction and disposal of materials.* In the case of materials for which the cost of cleaning and disinfection would exceed the value of the materials or for which cleaning and disinfection would be impracticable for any reason, the destruction and disposal of

~~the materials must be conducted in accordance with the initial State response and containment plan described in §56.10.~~

§56.6 Presentation of claims for indemnity and/or compensation.

Claims for the following must be documented on a form furnished by APHIS and presented to an APHIS employee or the State representative authorized to accept the claims:

- (a) ~~Compensation Indemnity~~ for the value of poultry to be destroyed due to infection with or exposure to H5/H7 LP AI;
- (b) ~~Compensation Indemnity~~ for the value of eggs to be destroyed ~~during testing for~~ due to infection or exposure to H5/H7 LP AI; and
- (c) Compensation for the cost of cleaning and disinfection (virus elimination) of premises, conveyances, and materials that came into contact with poultry infected with or exposed to H5/H7 LP AI, or, in the case of materials, if the cost of cleaning and disinfection (virus elimination) would exceed the value of the materials or cleaning and disinfection (virus elimination) would be impracticable for any reason, the cost of destruction and disposal for the materials.

§56.7 Mortgage against poultry or eggs.

When poultry or eggs have been destroyed under this part, any claim for indemnity must be presented on forms furnished by APHIS. The owner of the poultry or eggs must certify on the forms that the poultry or eggs covered are, or are not, subject to any mortgage as defined in this part. If the owner states there is a mortgage, the owner and each person holding a mortgage on the poultry or eggs must sign the APHIS-furnished form, consenting to the payment of indemnity to the person specified on the form.

§56.8 Conditions for payment

(a) When poultry or eggs have been destroyed pursuant to this part, the Administrator shall pay claims to any party with which the owner of the poultry or eggs has entered into a contract for the growing or care of the poultry or eggs. The indemnity the Administrator may pay to such a party or parties shall be determined as follows:

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- (1) Divide the value of the contract the owner of the poultry or eggs entered into with another party for the growing and care of the poultry or eggs in dollars by the duration of the contract as it was signed prior to the H5/H7 LP AI outbreak in days;
- (2) Multiply this figure by the time in days between the date the other party began to provide services relating to the destroyed poultry or eggs under the contract and the date the poultry or eggs were destroyed due to H5/H7 LP AI.

(b) (1) If indemnity for the destroyed poultry or eggs is being provided for 100 percent of eligible costs under §56.3(b), the Administrator may pay contractors eligible for ~~compensation indemnity~~ under this section 100 percent of the indemnity amount determined in paragraph (a) of this section.

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(2) If indemnity for the destroyed poultry or eggs is being provided for 25 percent of eligible costs under §56.3(b), the Administrator may pay contractors eligible for ~~compensation indemnity~~ under this section 25 percent of the indemnity amount determined in paragraph (a) of this section.

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(c) If a contractor receiving indemnity under this section has received any payment under his or her contract from the owner of the poultry or eggs at the time the poultry or eggs are destroyed,

the amount of indemnity for which the contract grower is eligible will be reduced by the amount of the payment the contract grower has already received.

(d) If indemnity is paid to a contractor under this section, the owner of the poultry or eggs will be eligible to receive the difference between the indemnity paid to the growers and the total amount of indemnity that may be paid for the poultry or eggs.

(e) In the event that determination of indemnity to a party with which the owner of destroyed poultry or eggs has entered into a contract for the growing or care of the poultry or eggs using the method described in paragraph (a) of this section is determined to be impractical or inappropriate, APHIS may use any other method that the Administrator deems appropriate to make that determination.

§56.9 Claims not allowed

(a) The Department will not allow claims arising out of the destruction of poultry unless the poultry have been appraised as prescribed in this part and the owners have signed the appraisal form indicating agreement with the appraisal amount as required by §56.4(a)(1).

(b) The Department will not allow claims arising out of the destruction of poultry unless the owners have signed a written agreement with APHIS in which they agree that if they maintain poultry in the future on the premises used for poultry for which indemnity and/or compensation is paid, they will maintain the poultry in accordance with a plan set forth by the Cooperating State Agency and will not introduce poultry onto the premises until after the date specified by the Cooperating State Agency. Persons who do not maintain their poultry and premises in accordance with this written agreement will not be eligible to receive indemnity and/or compensation under this part.

(c) The Department will not allow claims arising out of the destruction of poultry unless the poultry have been moved or handled by the owner in accordance with an agreement for the control and eradication of H5/H7 LPAI and in accordance with part 56, for any progeny of any poultry unless the poultry have been moved or handled by the owner in accordance with an agreement for the control and eradication of H5/H7 LPAI and in accordance with part 56, or for any poultry that become or have become infected with or exposed to H5/H7 LPAI because of actions not in accordance with an agreement for the control and eradication of H5/H7 LPAI or a violation of this part.

§56.10 Initial State response and containment plan

(a) In order for poultry owners within a State to be eligible for indemnity and/or compensation for 100 percent of eligible costs under §56.3(b), the State in which the poultry participate in the Plan must have in place an initial State response and containment plan that has been approved by APHIS. The initial State response and containment plan must be developed by the Official State Agency. In States where the Official State Agency is different than the Cooperating State Agency, the Cooperating State Agency must also participate in the development of the plan. The plan must be administered by the Cooperating State Agency of the relevant State. This plan must include:

- (1) Provisions for a standing emergency disease management committee, regular meetings, and exercises, including coordination with any tribal governments that may be affected;
- (2) A minimum biosecurity plan followed by all poultry producers;
- (3) Provisions for adequate diagnostic resources;
- (4) Detailed, specific procedures for initial handling and investigation of suspected cases of H5/H7 LPAI;
- (5) Detailed, specific procedures for reporting test results to APHIS. These procedures must be developed after appropriate consultation with poultry producers in the State and

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must provide for the reporting only of confirmed cases of H5/H7 LPAI in accordance with §146.13 of this chapter;

- (6) Detailed, strict quarantine measures for presumptive and confirmed index cases;
- (7) Provisions for developing flock plans for infected and exposed flocks;
- (8) Detailed plans for disposal of infected flocks, including preexisting agreements with regulatory agencies and detailed plans for carcass disposal, disposal sites, and resources for conducting disposal, and detailed plans for disposal of materials that come into contact with poultry infected with or exposed to H5/H7 LPAI;
- (9) Detailed plans for cleaning and disinfection of premises, repopulation, and monitoring after repopulation;
- (10) Provisions for appropriate control/monitoring zones, contact surveys, and movement restrictions;
- (11) Provisions for monitoring activities in control zones;
- (12) If vaccination is considered as an option, a written plan for use in place with proper controls and provisions for APHIS approval of any use of vaccine;
- (13) Plans for H5/H7 LPAI-negative flocks that provide for quarantine, testing, and controlled marketing; and
- (14) Public awareness and education programs regarding avian influenza.

(b) If a State is designated a U.S. Avian Influenza Monitored State, Layers under §146.24(a) of this chapter or a U.S. Avian Influenza Monitored State, Turkeys under §146.44(a) of this chapter, it will lose that status during any outbreak of H5/H7 LPAI and for 90 days after the destruction and disposal of all infected or exposed birds and cleaning and disinfection of all affected premises are completed.

§56.11 H5/H7 LPAI Guidance Documents

The following LPAI Guidance Documents can be found on the NPIP website (www.poultryimprovement.org):

- a. Response, Communication, and Investigation of Avian Influenza in Domestic Poultry
- b. Procedures for Indemnity and Compensation Claims in Cases of H5/H7 Low Pathogenicity Avian Influenza in Poultry
- c. Development and Approval of Initial State Response and Containment Plans for H5/H7 Low Pathogenicity Avian Influenza (LPAI)
- d. Reporting Confirmed Findings of Low Pathogenicity Avian Influenza H5/H7 Subtypes to the World Organization for Animal Health (OIE) and to Trading Partners

Reason: USDA has proposed changes to the policy for compensation and indemnity for the H5/H7 LPAI program. This proposal serves to update the language in Part 56 to reflect the language used in USDA. Further, this proposal allows control marketed flocks to maintain their AI certifications and be slaughtered in commercial slaughter plants.

Sponsors: USDA-APHIS-Veterinary Services

Late Proposal No. 2

Delegates Combined

147.48 Approval conference recommendations by the Department

Proposals adopted by the official delegates will be recommended to the Department for incorporation into the provisions of NPIP. The Department reserves the right to approve or disapprove the recommendations of the conference as an integral part of its sponsorship of the National Poultry Improvement Plan. The Department will publish the recommendations in the Federal Register within fourteen months following the NPIP Biennial Conference, and the final rule will be published in the Federal Register within seventeen months of the Biennial Conference.

Reason: To establish a time frame for publication of the NPIP Biennial Conference proposed changes prior to the start of the following year's Biennial Conference.

Sponsor: Paul Wm. Brennan, Indiana State Poultry Association

Program Standards - Proposal No. 1 (Amended by TC)

Delegates: Combined

Standard A – Blood Testing Procedures

(5) Procedure for determining the status of flocks reacting to tests for *Mycoplasma gallisepticum*, *Mycoplasma synoviae*, and *Mycoplasma meleagridis*.

Procedures for isolation and identification of *Mycoplasma* may be found in A Laboratory Manual for the Isolation, Identification and Characterization of Avian Pathogens, published by the American Association of Avian Pathologists; Avian Mycoplasmosis (*Mycoplasma gallisepticum*), Manual of Diagnostic Tests and Vaccines for Terrestrial Animals, Fifth Ed., Office International des Epizooties, pp 842–855, 2004; and Program Standards, Standard B sections (6) and (7).

(a) Determining flock *Mycoplasma* status

(a) The status of a flock for *Mycoplasma* shall be determined according to the following criteria:

- (1) If the enzyme-labeled immunosorbent assay (ELISA), official molecular examination procedure, or serum plate test is negative, the flock qualifies for the classification for which it was tested.
- (2) If the ELISA or serum plate test is positive, the hemagglutination inhibition (HI) test or a molecular examination procedure shall be conducted: Provided, for the molecular examination procedure the appropriate antigen detection samples from a minimum of 30 ~~clinically affected~~ birds are tested. And Pprovided, for the HI test, that if more than 50 percent of the samples are positive for *M. gallisepticum*, *M. meleagridis*, or *M. synoviae*, the HI test shall be conducted on 10 percent of the positive samples or 25 positive samples, whichever is greater. HI titers of 1:40 or more may be interpreted as suspicious and appropriate antigen detection samples should be taken promptly (within 7 10 days of the original sampling) from 30 ~~clinically affected~~ birds and examined by an approved cultural technique individually, or pooled (up to 5 swabs per test) and used in a molecular examination procedure or in vivo bioassay.
- (3) If the in vivo bioassay, molecular examination procedure, or culture procedure is negative, the Official State Agency may qualify the flock for the classification for which it was tested. In the event of contaminated cultures, the molecular examination technique must be used to make a final determination.
- (4) If the in vivo bioassay, molecular examination procedure, or culture procedure is positive, the flock will be considered infected.

Reason: Clarification of the sampling required if molecular examination procedure is selected as the confirmatory test. As well, it can be logistically challenging if suspect sera must be sent over a weekend to obtain HI results and be able to collect swabs for PCR within 7 days on occasion.

Sponsor: Dr. Eric Jensen
Aviagen North America

Program Standards - Proposal No. 2

Delegates: Combined

Standard A—Blood Testing Procedures

(6) Standard test procedures for mycoplasma.⁵

The serum plate agglutination test and the enzyme-linked immunosorbent assay (ELISA) test should be considered basic screening tests for mycoplasma antibodies. Alternatively, an approved molecular examination procedure may be used alone or in conjunction with serology as a screening test for mycoplasma. The test selected will depend on preference, laboratory facilities, and availability of antigen. ~~The serological tests~~ determine flock status rather than individual bird status, since occasional reactions are nonspecific. Under normal circumstances, the rate of such nonspecific reactions is low. Nonspecific reactions may occasionally be high, particularly after the use of erysipelas bacterin in turkeys and where mycoplasma antibodies are present for closely related mycoplasma other than for the species being tested. The hemagglutination inhibition (HI) test is too cumbersome for routine screening use. Positive reactions are extremely accurate, however, and are useful in evaluating serum samples that react with the ELISA and/or plate antigens. The test should be conducted with 4 HA units. Titers of 1:80 or greater for both chicken and turkey sera are considered positive, while a 1:40 titer would be suspicious and additional tests should be required.

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Reason: Clarification that, in addition to SPA and ELISA, a molecular examination procedure may be used to screen flocks for mycoplasma.

Sponsor: Dr. Eric Jensen
Aviagen North America

Program Standards - Proposal No. 3 (Amended by TC)

Delegates: Combined

Standard A – Blood Testing Procedures

(8) Standard test procedures for avian influenza

(a) The agar gel immunodiffusion (AGID) test should be considered the basic screening test for antibodies to Type A influenza viruses. The AGID test is used to detect circulating antibodies to Type A influenza group-specific antigens, namely the ribonucleoprotein (RNP) and matrix (M) proteins. Therefore, this test will detect antibodies to all influenza A viruses, regardless of subtype. The AGID test can also be used as a group-specific test to identify isolates as Type A influenza viruses. The method used is similar to that described by Beard.⁶ The basis for the AGID test is the concurrent migration of antigen and antibodies toward each other through an agar gel matrix. When the antigen and specific antibodies come in contact, they combine to form a precipitate that is trapped in the gel matrix and produces a visible line. The precipitin line forms where the concentration of antigen and antibodies is optimum. Differences in the relative concentration of the antigen or antibodies will shift the location of the line towards the well with the lowest concentration or result in the absence of a precipitin line. Electrolyte concentration, pH, temperature, and other variables also affect precipitate formation.

(1) The testing procedure outlined in NVSL SOP (Avian Influenza Agar Gel Immunodiffusion Test to Detect Antibodies to Type A Influenza Virus) shall be followed. The approved testing procedure shall be posted to the NPIP website, and when procedural changes are made, they must be reviewed by the Technical Committee and approved by the General Conference Committee and notifications shall be sent to the NPIP Official State Agencies and the NPIP Authorized Laboratories. Document requests should be submitted to NVSL by e-mail at nvsl.dvl.avian@aphis.usda.gov.

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(1) *Materials needed.*

- (i) Refrigerator (4 °C).
- (ii) Freezer (-20 °C).
- (iii) Incubator or airtight container for room temperature (approximately 25 °C) incubations.
- (iv) Autoclave.
- (v) Hot plate/stirrer and magnetic stir bar (optional).
- (vi) Vacuum pump.
- (vii) Microscope illuminator or other appropriate light source for viewing results.
- (viii) Immunodiffusion template cutter, seven-well pattern (a center well surrounded by six evenly spaced wells). Wells are 5.3 mm in diameter and 2.4 mm apart.
- (ix) Top loading balance (capable of measuring 0.1 gm differences).
- (x) Pipetting device capable of delivering 50µl portions.
- (xi) Common laboratory supplies and glassware—Erlenmeyer flasks, graduated cylinders, pipettes, 100 × 15 mm or 60 × 15 mm petri dishes, flexible vacuum tubing, side-arm flask (500 mL or larger), and a 12- or 14-gauge blunt-ended cannula.

(2) *Reagents needed.*

- (i) Phosphate buffered saline (PBS), 0.01M, pH 7.2 (NVSL media #30054 or equivalent).
- (ii) Agarose (Type II Medium grade, Sigma Chemical Co. Cat.# A-6877 or equivalent).
- (iii) Avian influenza AGID antigen and positive control antiserum approved by the Department and the Official State Agency.
- (iv) Strong positive, weak positive, and negative control antisera approved by the Department and the Official State Agency (negative control antisera optional).

(3) *Preparing the avian influenza AGID agar.*

(i) Weigh 9 gm of agarose and 80 gm of NaCl and add to 1 liter of PBS (0.01 M, pH 7.2) in a 2-liter Erlenmeyer flask.

(ii) To mix the agar, either:

(A) Autoclave the mixture for 10 minutes and mix the contents by swirling after removing from the autoclave to ensure a homogeneous mixture of ingredients; or

(B) Dissolve the mixture by bringing to a boil on a hot plate using a magnetic stir bar to mix the contents in the flask while heating. After boiling, allow the agar to cool at room temperature (approximately 25°C) for 10 to 15 minutes before dispensing into petri plates.

(iii) Agar can be dispensed into small quantities (daily working volumes) and stored in airtight containers at 4°C for several weeks, and melted and dispensed into plates as needed.

Note: Do not use agar if microbial contamination or precipitate is observed.

(4) Performing the AGID—

(i) *Detection of serum antibodies.*

(A) Dispense 15 to 17 mL of melted agar into a 100 × 15 mm petri plate or 5 to 6 mL agar into a 60 × 15 mm petri plate using a 25 mL pipette. The agar thickness should be approximately 2.8 mm.

(B) Allow plates to cool in a relatively dust-free environment with the lids off to permit the escape of water vapor. The lids should be left off for at least 15 minutes, but not longer than 30 minutes, as electrolyte concentration of the agar may change due to evaporation and adversely affect formation of precipitin lines.

Note: Plates should be used within 24 hours after they are poured.

(C) Record the sample identification, reagent lot numbers, test date, and identification of personnel performing and reading the test.

(D) Using the template, cut the agar after it has hardened. Up to seven template patterns can be cut in a 100 × 15 mm plate and two patterns can be cut in a 60 × 15 mm plate.

(E) Remove the agar plugs by aspiration with a 12- to 14-gauge cannula connected to a side-arm flask with a piece of silicone or rubber tubing that is connected to a vacuum pump with tubing. Adjust the vacuum so that the agar surrounding the wells is not disturbed when removing the plugs.

(F) To prepare the wells, place 50 µl of avian influenza AGID antigen in the center well using a micropipette with an attached pipette tip. Place 50 µl AI AGID positive control antiserum in each of three alternate peripheral wells, and add 50 µl per well of test sera in the three remaining wells. This arrangement provides a positive control line on each side of the test serum, thus providing for the development of lines of identity on both sides of each test serum (see figure 1).

Note: A pattern can be included with positive, weak positive, and negative reference serum in the test sera wells to aid in the interpretation of results (see figure 2).

(G) Cover each plate after filling all wells and allow the plates to incubate for 24 hours at room temperature (approximately 25 °C) in a closed chamber to prevent evaporation. Humidity should be provided by placing a damp paper towel in the incubation chamber.

Note: Temperature changes during migration may lead to artifacts.

(ii) *Interpretation of test results.*

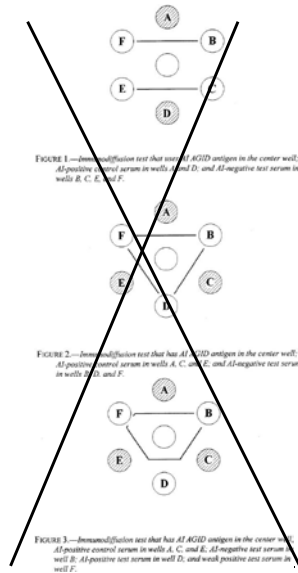
(A) Remove the lid and examine reactions from above by placing the plate(s) over a black background, and illuminate the plate with a light source directed at an angle from below. A microscope illuminator works well and allows for varying intensities of light and positions.

(B) The type of reaction will vary with the concentration of antibody in the sample being tested. The positive control serum line is the basis for reading the test. If the line is not distinct, the test is not valid and must be repeated. The following types of reactions are observed (see figure 3):

(1) *Negative reaction.* The control lines continue into the test sample well without bending or with a slight bend away from the antigen well and toward the positive control serum well.

(2) *Positive reaction.* The control lines join with, and form a continuous line (line of identity) with, the line between the test serum and antigen. The location of the line will depend on the concentration of antibodies in the test serum. Weakly positive samples may not produce a complete line between the antigen and test serum but may only cause the tip or end of the control line to bend inward toward the test well.

(3) *Non-specific lines.* These lines occasionally are observed between the antigen and test serum well. The control lines will pass through the non-specific line and continue on into the test serum well. The non-specific line does not form a continuous line with positive control lines.



Beard, C.W. Demonstration of type-specific influenza antibody in mammalian and avian sera by immunodiffusion. Bull. Wld. Hlth. Org. 42:779-785. 1970.

Reason: Due to inconsistencies noted between the 9CFR and other written procedures for AI AGID, NVSL conducted a method evaluation and discovered opportunities to optimize the assay. While the procedure as written in the 9CFR is not incorrect, the details included in the updated NVSL SOP-AV-0045 improve the overall accuracy and interpretation of the assay. Additionally, as advances in technology continue future updates may be needed.

Sponsor: Dr. Mia Torchetti, Mary Lea Killian, Terra Jenson
USDA APHIS National Veterinary Services Laboratory (NVSL)

Program Standards Proposal No. 4 (Amended by TC)

Delegates: Combined

Standard B - Bacteriological Examination Procedures as amended by the Salmonella Technical Committee

(1) Reserved

~~(1) — Laboratory procedure recommended for the bacteriological examination of egg-type and meat-type breeding flocks with salmonella enteritidis positive environments. Birds selected for bacteriological examination from egg-type and meat-type breeding flocks positive for Salmonella enteritidis after environmental monitoring should be examined as described in Section B(2)(a) of these Program Standards, with the following exceptions and modifications allowed due to the high number of birds required for examination:~~

~~(a) Except when visibly pathological tissues are present, direct culture, Section B(2)(a)(1) of these standards, may be omitted.~~

(2) **Laboratory procedure recommended for the bacteriological examination of Salmonella from birds**

(a) **For egg- and meat-type chickens, turkeys, waterfowl, exhibition poultry, and game birds**

~~All reactors to the pullorum-typhoid tests, up to 25 birds, and birds from Salmonella enteritidis (SE) positive environments should be cultured in accordance with both the direct enrichment (paragraph (a)(1)) and selective enrichment (paragraph (a)(2)) procedures described in this section: Provided, if there are more than four reactors to the pullorum-typhoid tests in the flock, a minimum of four reactors as provided for in 9 CFR 145.14(a)(6)(ii) shall be submitted to the authorized laboratory for bacteriological examination. Careful aseptic technique should be used when collecting all tissue samples.~~

~~For reactors to the pullorum-typhoid tests, if there are more than four reactors in a flock, a minimum of four reactors shall be submitted to the authorized laboratory; if the flock has four or fewer reactors all the reactors must be submitted [145.14(a)(6)(ii)]. The isolation of S. Enteritidis from U.S. S. Enteritidis Clean flocks will result in the submission of 60 live birds from a flock of 5,000 birds or more, or 30 live birds from a flock with fewer than 5,000 birds from multiplier egg-type chicken breeding flocks [145.23(d)(2)] or primary egg-type chicken breeding flocks [145.73(d)(2)] and 25 birds from primary meat-type chicken breeding flocks [145.83(e)(3)]. These birds should be cultured in accordance with both direct culture (paragraph (a)(1)) and selective enrichment (paragraph (a)(2)) procedures described in this section. Provided, if there are no grossly abnormal or diseased tissues present, direct culture may be omitted. Careful aseptic technique should be used when collecting all tissue samples.~~

(1) Direct culture (refer to illustration 1). Grossly abnormal or diseased liver, heart, pericardial sac, spleen, lung, kidney, peritoneum, gallbladder, oviduct, misshapen ova or testes, inflamed or unabsorbed yolk sac, and other visibly pathological tissues where purulent, necrotic, or proliferative lesions are seen (including cysts, abscesses, hypopyon, and inflamed serosal surfaces) should be sampled for direct culture using either flamed wire loops or sterile swabs. Since some strains may not dependably survive and grow in certain selective media,

inoculate non-selective plates (such as blood or nutrient agar) and selective plates (such as MacConkey [MAC] and brilliant green novobiocin [BGN] for suspect *S. pullorum* or *S. gallinarum* and MAC, BGN, and xylose- lysine-tergitol 4 [XLT 4] for SE). Refer to illustration 1 for recommended bacteriological recovery and identification procedures.⁷ Proceed immediately with collection of organs and tissues for selective enrichment culture.

(2) Selective enrichment culture (refer to illustration 1). Collect and culture organ samples separately from intestinal samples, with intestinal tissues collected last to prevent cross-contamination. Samples from the following organs or sites should be collected for culture in selective enrichment broth:

- (i) Heart (apex, pericardial sac, and contents if present);
- (ii) Liver (portions exhibiting lesions or, in grossly normal organs, the drained gallbladder and adjacent liver tissues);
- (iii) Ovary-Testes (entire inactive ovary or testes, but if ovary is active, include any atypical ova);
- (iv) Oviduct (if active, include any debris and dehydrated ova);
- (v) Kidneys and spleen; and
- (vi) Other visibly pathological sites where purulent, necrotic, or proliferative lesions are seen.

(3) From each bird, aseptically collect up to 10 to 15 grams of each organ or site listed in paragraph (a)(2) of this section. Mince, grind, or blend and place in a sterile plastic bag. All the organs or sites listed in paragraph (a)(2) of this section from the same bird may be pooled into one bag. Do not pool samples from more than one bird. Add sufficient tetrathionate enrichment broth to give a 1:10 (sample to enrichment) ratio. Incubate the sample at $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ ~~or 42.0°C~~ for 20 to 24 hours. Follow the procedure outlined in illustration 1 for the isolation and identification of *Salmonella*.

(4) From each bird, aseptically collect 10 to 15 grams of each of the following parts of the digestive tract: Crop wall, duodenum, jejunum (including remnant of yolk sac), both ceca, cecal tonsils, and rectum-cloaca. Mince, grind, or blend tissues and pool them into a sterile plastic bag. Do not pool tissues from different birds into the same sample. Add sufficient tetrathionate enrichment broth to give a 1:10 (sample to enrichment) ratio. Incubate the sample at $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ ~~or 42°C~~ for 20 to 24 hours. Follow the procedure outlined in illustration 1 for the isolation and identification of *Salmonella*.

(5) After selective enrichment, inoculate selective plates (such as MAC and BGN for *S. Pullorum* or *S. Gallinarum* and MAC, BGN, and XLT4) for SE). Incubate the plate at $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ for 20 to 24 hours. ~~Inoculate~~ Select three to five *Salmonella*-suspect colonies from plates. Inoculate each suspect colony individually into pairs of ~~into~~ triple sugar iron (TSI) and lysine iron agar (LIA) slants or equivalent method (i.e., inoculate TSI and LIA pair from one colony). Incubate slants at $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ for 20-24 hours. If there are no suspect colonies after 24 hours of incubation, incubate the plates an additional 24 hours before considering negative. Screen colonies by serological (i.e., serogroup) and biochemical procedures (e.g., the Analytical Profile Index for Enterobacteriaceae [API]) as shown in illustration 1.

(6) If the initial selective enrichment is negative [Section B(2)(a)(5)] for *Salmonella*, a delayed secondary enrichment (DSE) procedure is used. Leave the tetrathionate-enriched sample at room temperature for 5 to 7 days. Transfer 1 mL of the culture into a tube containing 10 mL of fresh tetrathionate enrichment

broth, incubate at $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ for 20 to 24 hours, and plate as in Section B(2)(a)(5).

(7) Serogroup all isolates identified as salmonellae and serotype all serogroup D1 isolates. ~~Phage type all SE isolates.~~

(3) Procedures for collection, isolation, and identification of Salmonella from house environmental samples, cloacal swabs, and hatchery samples.

a) For egg- and meat- type chickens, turkeys, waterfowl, exhibition poultry, and game birds.

(1) *Poultry house environmental samples.*

(ii) Drag swabs (DS).

(A) Preparation. DS may be purchased commercially or be user prepared. One suggested method of making the DS assemblies is as follows: A sterile gauze pad is folded in half and a 2-foot long (60cm) piece of twine is securely attached to the folded pad using a paper clip, staple, or similar device. A second sterile gauze pad is similarly fastened to a 5-foot (150 cm) long piece of twine. The shorter piece of twine is then tied to the longer piece producing a DS sample set of two swabs arranged in a Y-shaped configuration. Alternatively, two separate DS samplers may be prepared. The twine is wrapped around the swabs, and the swabs moistened with double-strength skim milk (DSSM) or BPW. The moistened swabs are placed in an instrument package. The sterilized swabs contained in the instrument pack may be frozen (to prevent drying) until use.

(B) Procedure. At the farm the thawed DS assemblies are unraveled and the ends of the twine held in gloved hands. The swabs are dragged across the environmental surfaces of the house for 15 minutes or the length of the house (down and back). One set of swabs (two individual pads) is dragged across the center of the house floor and another set of swabs (two individual pads) is dragged across the inside perimeter of the house floor. The four pads are individually placed in labeled, sterile bags. If necessary to prevent drying out, additional DSSM (evaporated skim milk) or BPW may be added to the bags. The bags should be protected from excessive heat and submitted as soon as possible to the authorized laboratory for testing. If samples are to be processed within 48 hours after collection at the farm, the drag swab assemblies may be stored in BPW. Samples to be processed after 48 hours and before 5 days, must be pre- moistened with DSSM. If the samples cannot be submitted to the laboratory the same day, they should be stored 2° - 48°C or placed in a cooler with ice or ice packs (do not freeze) for no more than 5 days before culturing.

(iii) Shoe cover swabs (Boot swabs).

Absorbable fabric shoe covers involve the exposure of the bottom surface of shoe covers to the surface of floor litter and slat areas. Wearing clean gloves, place the shoe covers over footwear that is only worn inside the poultry house. This can be footwear dedicated to the facility or disposable overshoes. Each pair of shoe covers should be worn while walking at a normal pace over a distance of 1,000 feet (305 meters). For flocks with fewer than 500 breeders, at least 1 pair of shoe covers should be worn to

sample the floor of the bird area. For flocks with 500 or more breeders, at least 2 pairs of shoe covers should be worn to sample the floor of the bird area. After sampling, place each shoe cover in a sterile container with 30 ml of double strength skim milk, unless pre-moistened swabs (BPW) are used. Seal the sterile containers and promptly refrigerate them at 2° to 4-8°C or place in a cooler with ice or ice packs. Do not freeze. If shoe cover swab samples are to be processed within 48 hours after collection, the shoe cover swab samples may be pre-moistened with BPW. Samples to be processed after 48 hours and before 5 days must be pre-moistened with DSSM. All samples are to be placed in a cooler with ice or ice packs for transport and refrigeration at 2° – 8° C in the period prior to the addition of the pre-enrichment broth. Samples should be stored at refrigerator temperatures of 2° to 4-8°C no more than 5 days before culturing.

(iv) Nest box or egg belt swabs as alternative sampling source

(A) Two sterile pre-moistened (ex. DSSM or BPW) gauze pads or sponges are swabbed inside approximately 10 percent of the nest boxes. Each swab or sponge is placed into a separate sterile bag and submitted to the authorized laboratory.

(B) Two sterile pre-moistened (ex. DSSM or BPW) gauze pads or sponges are used to swab the egg belts. At least 30 feet of belt material is swabbed with each swab. Each swab is placed into a separate sterile bag and submitted to the authorized laboratory.

(C) If nest box or egg belt swab samples are to be processed within 48 hours after collection, the nest box or egg belt swab samples may be pre-moistened with BPW. Samples to be processed after 48 hours and before 5 days must be pre-moistened with DSSM. All samples are to be placed in a cooler with ice or ice packs for transport and refrigeration at 2° – 8° C in the period prior to the addition of the pre-enrichment broth.

(2) Cloacal swabs. Cloacal swabs for bacteriological examination shall be taken from each bird in the flock or from a minimum of 500 birds in accordance with the procedure described in this section. A sterile cotton-tipped applicator or swab is inserted into the cloaca and rectum of the bird in such a manner to ensure the collection of fecal material. The applicator may be broken off into a sterile tube. The cloacal swabs may be combined in multiples of five or in combinations specified by the authorized laboratory.

(3) Hatchery samples. Hatchery-related samples, such as chick box papers, meconium, and fluff may be examined for the presence of Salmonella to indicate the transfer of Salmonella from parent to offspring.

(i) Chick box papers. Chick box paper samples may be collected by an authorized agent according to paragraph (a)(3)(i)(A) of this section or may be submitted directly to an authorized laboratory for testing according to paragraph (a)(3)(ii)(B) of this section. It is important to remove, with sanitized or gloved hands, the paper from the chick box before the box is placed in the brooding house.

(A) Instructions for sampling chick box papers. One chick box paper is collected for every 10 boxes of chicks placed in a house. With sanitized and gloved hands, lay out the papers on a clean, disinfected surface. Saturate a sterile gauze pad or sponge with DSSM or BPW and swab the surface of 5 chick box papers. The pad should be rubbed over approximately 75 percent of each paper

with sufficient pressure to remove any dried meconium. Addition of more DSSM or BPW may facilitate sampling. The process is repeated with a second swab and the other five chick box papers. Both swabs may be added to a single sterile, labeled plastic bag and submitted to the authorized laboratory. If chick box paper samples are to be processed within 48 hours after collection, the chick box paper samples may be pre-moistened with BPW. Samples to be processed after 48 hours and before 5 days must be pre-moistened with DSSM. All samples are to be placed in a cooler with ice or ice packs for transport and refrigeration at 2° – 8° C in the period prior to the addition of the pre-enrichment broth. Promptly refrigerate the Whirl-Pak bags containing the samples and transport them, on ice or otherwise refrigerated, to a laboratory to be cultured within 5 days of collection.

~~(ii) The Plan participant may send chick box papers directly to a laboratory, where samples may be collected as described in paragraph (a)(3)(i)(A) of this section. To send chick box papers directly to a laboratory:~~

~~(A) Collect 1 chick box paper for each 10 boxes of chicks placed in a house and place the chick papers immediately into large plastic bags and label and seal the bags.~~

~~(B) Place the plastic bags containing the chick box papers in a clean box and transport them within 48 hours to a laboratory. The plastic bags do not require refrigeration.~~

(B) Instructions for sending chick box papers directly to the laboratory. With sanitized or gloved hands, collect 1 chick box paper for each 10 boxes of chicks placed in a house. Place the chick papers immediately into large clean plastic bags, label and seal the bags. Transport them to the laboratory within 48 hours. The plastic bags do not require refrigeration.

~~(iii) Chick meconium. After collection, the container of meconium is mixed to obtain a uniform consistency. In the laboratory, a 25-gram sample will be removed for bacteriological examination.~~

~~(iv) Fluff. Samples of Fluff samples may be collected from the floor of the hatchery or from the tray following hatching. The fluff sample may be collected with sanitized or gloved hands by either swabbing the floor or tray with a pre-moistened gauze pad or sponge or by placing fluff material directly into a sterile bag.~~

b) Isolation and identification of Salmonella.

There are two enrichment procedures approved for the isolation of *Salmonella* from environmental samples as described in this section (See Illustration 2). Alternatively, approved rapid methods may be used to detect the presence of *Salmonella*. Provided, positive samples must be confirmed by culture which must then be isolated. The enriched sample used for the rapid assay should be transferred into MSR/V and follow the isolation and identification procedure in PS Standard B(3)(b)(1)(ii – vi). The culture process must be started within 24 hours of the positive screening test.

(1) Direct tetrathionate (TT) enrichment followed by Modified Semisolid Rappaport-Vassiliadis (MSRV) enrichment (Illustration 2).

(i) Fresh Tetrathionate enrichment broth is added to the sample to give a 1:10 (sample to enrichment) ratio. Incubate the samples at $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ or 42°C for 20 to 24 hours.

(ii) After incubation, transfer ~~approximately 100 microliters (3 drops)~~ of the enriched culture into (subsurface) an MSR plate at a 1:100 ratio of sample to enrichment. Incubate the plate right side up at 42°C for 24 hours.

(iii) Observe the MSR plate for growth migrating from the point of inoculation. If present, insert a sterile loop into the outer edge of the zone of growth and inoculate selective agar plates, such as BGN and XLT4.

(iv) If no zone of growth is present, incubate the MSR plate at 42°C for another 24 hours. Observe the MSR plate for growth migrating from the point of inoculation. If growth is present, insert a sterile loop into the outer edge of the zone of growth and inoculate selective agar plates, such as BGN and XLT4. If still no zone, insert the loop into the point of inoculation and inoculate selective agar plates. This ensures that weakly or non-motile strains of *Salmonella* will not be missed.

(v) Incubate the selective agar plates at $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ for 20 to 24 hours. Observe the plates for *Salmonella* suspect colonies. Screen three to five colonies by inoculating them individually into triple sugar iron agar (TSI) and lysine iron agar (LIA) slants or equivalent method. Incubate the slants at $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ for 20 to 24 hours. Screen the colonies by serological (i.e., serogroup) or biochemical (e.g. API) procedures as shown in Illustration 2.

(vi) Serogroup all isolates identified as *Salmonella* and serotype all serogroup D isolates. ~~Phage type one SE isolate per flock per submission.~~

(2) Pre-enrichment followed by selective enrichment. (Illustration 2.)

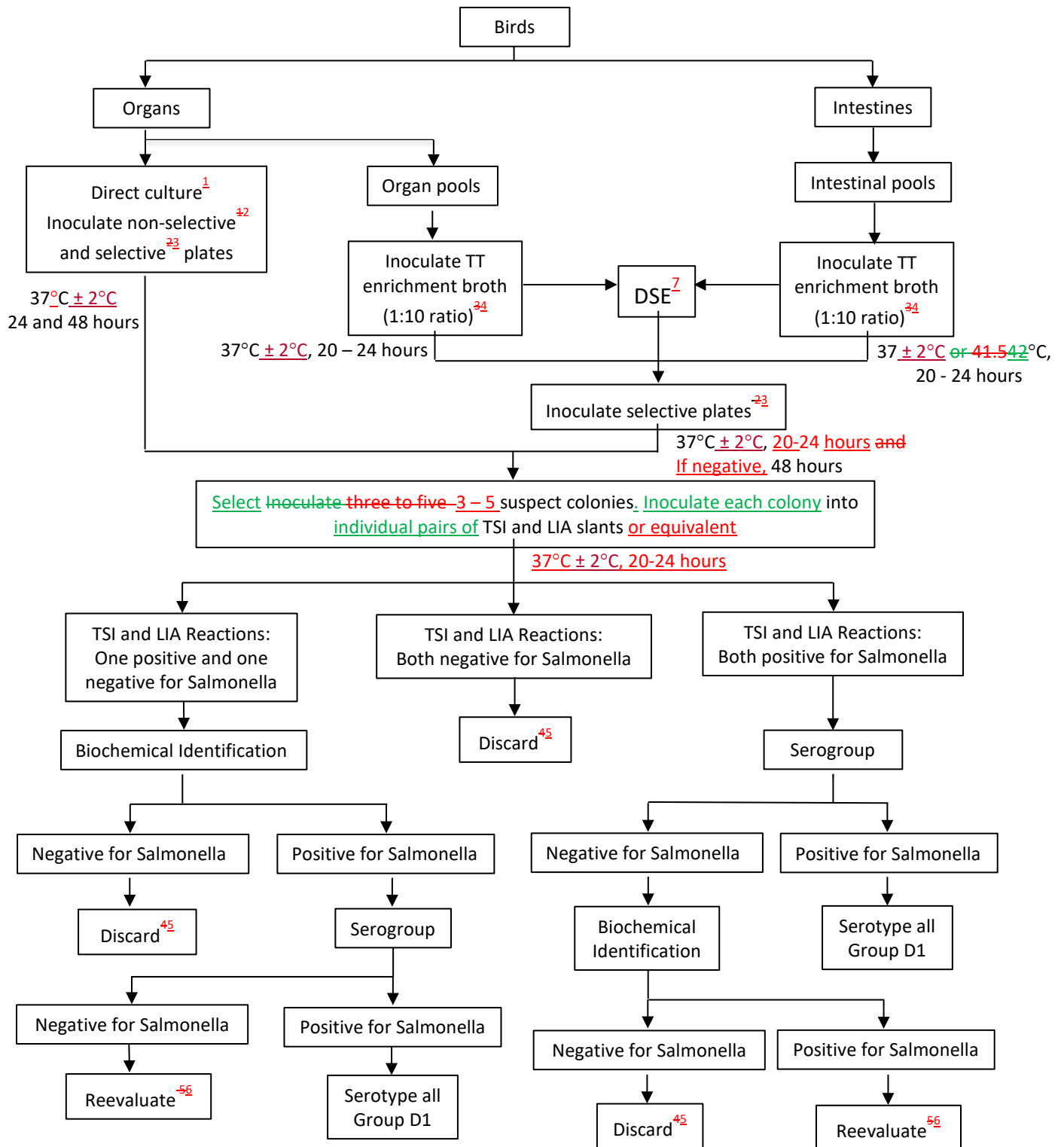
(i) Pre-enrichment broth (e.g. buffered peptone water, BPW) is added to the sample to give a 1:10 (sample to enrichment) ratio. Incubate the sample at $37^{\circ}\text{C} \pm 2^{\circ}\text{C}$ for 20 to 24 hours.

(ii) Transfer 1 ml of the pre-enriched sample into a tube containing 10 ml of tetrathionate enrichment broth and transfer 0.1 ml into either a tube containing 10 ml of Rappaport-Vassiliadis (RV) enrichment broth or into a MSR plate. Incubate at 42°C for 20 to 24 hours.

(iii) After incubation, inoculate the ~~tetrathionate~~ TT and RV enrichments onto separate selective agar plates, such as BGN and XLT4. If the MSR media was inoculated, then follow the steps in (1)(iii) and (1)(iv).

(iv) Screen the selective agar plates for *Salmonella* as described in (1)(v) and (1)(vi).

Illustration 1. Procedure for culturing Pullorum-Typhoid reactors and birds from SE positive environments.



¹. Direct plating is only required when there are grossly abnormal diseased tissues or organs present.

⁴². Non-selective plates, such as blood or nutrient agar.

²³. Selective plates, such as MacConkey (MAC) and Brilliant Green Novobiocin (BGN) for pullorum-typhoid reactors (Pullorum and Gallinarum are typically H₂S-negative after 24 hours incubation), and MacConkey MAC, BGN and/or xylose-lysine tertitol 4 (XLT 4) for birds from SE-positive environments.

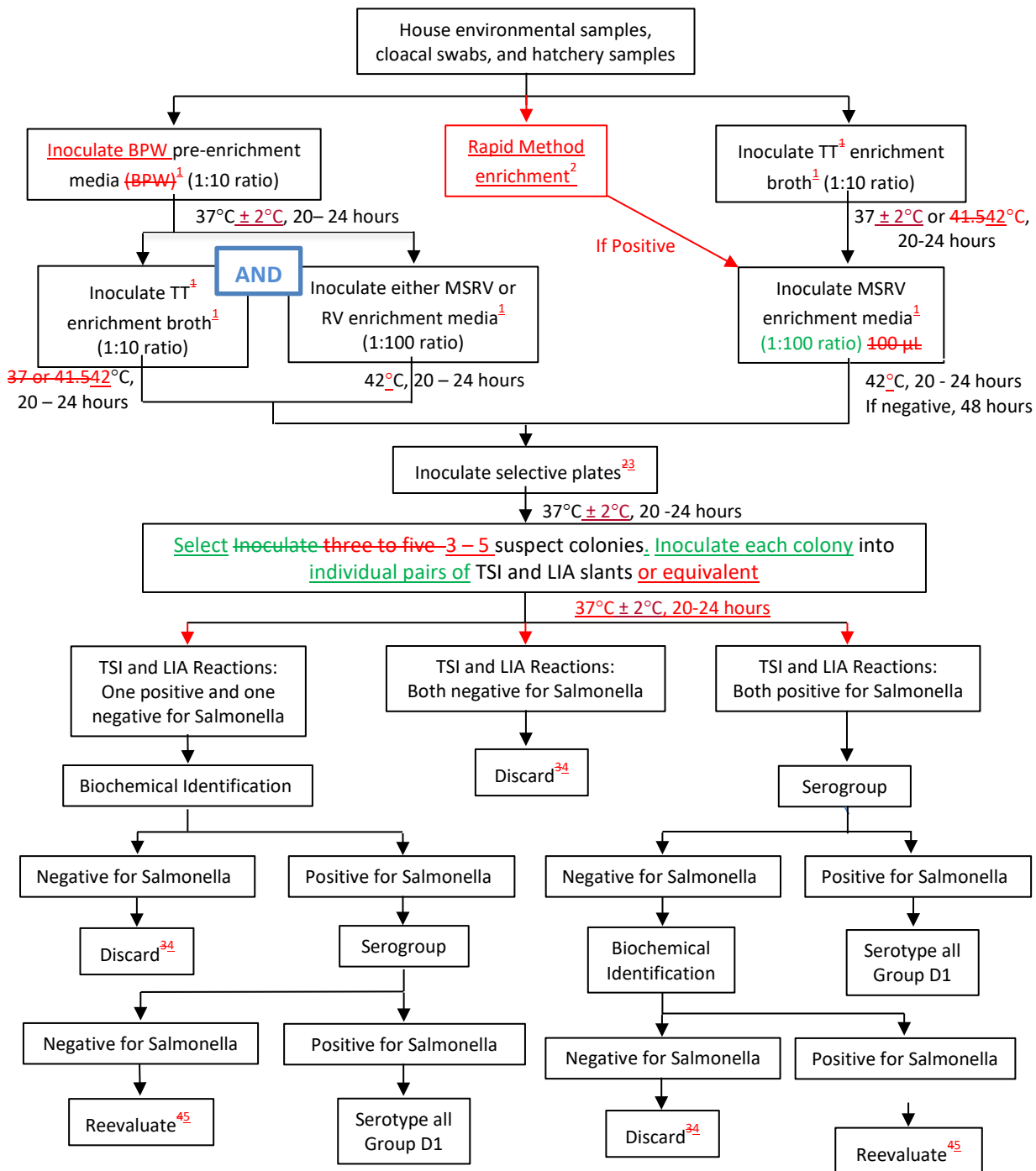
³⁴. Tetrathionate enrichment broth.

⁴⁵. Reevaluate if epidemiologic, necropsy or other information indicates the presence of an unusual strain of Salmonella.

⁵⁶. If biochemical identification and serogroup procedures results are inconclusive restreak original colony onto non-selective plating media to check for purity. Repeat biochemical and serogroup tests.

⁷. If initial selective enrichment is negative, use Delayed Secondary Enrichment (DSE): Hold TT-enriched sample at room temperature for 5-7 days, then transfer 1 mL of the sample into 10 mL TT. Incubate at 37°C ± 2°C for 20-24 hours.

Illustration 2. Two approved culture procedures for house environmental samples, cloacal swabs and hatchery samples



1. Buffered Peptone Water (BPW), Tetrathionate (TT) enrichment broth, e.g., Rappaport-Vassiliades (RV) or modified semisolid Rappaport-Vassiliades media (MSRV).

2. Refer to the manufacturer's protocols for using NPIP-approved rapid methods. All rapid methods are considered screening tests; therefore, samples giving positive results must be confirmed by NPIP culture methods.

23. Selective plates, such as Brilliant Green with Novobiocin (BGN) ~~or~~ and Xylose-Lysine Tergitol 4 (XLT 4).

34. Reevaluate if epidemiologic, necropsy, or other information indicates the presence of an unusual strain of Salmonella.

45. If biochemical identification and serogroup procedures are inconclusive, restreak original colony onto non-selective plating media to check for purity. Repeat biochemical and serology tests.

Reason: This proposal attempts to clean-up and clarify the isolation procedures for Salmonella and to ensure that Illustrations 1 and 2 agree with the written text in Standards B(2) and B(3). We removed the requirement for phage typing and clarified sampling of chick box papers. We widened the temperature range for refrigeration of samples from 2°-4°C to 2°-8°C to facilitate compliance with the standards set forth by laboratory accreditation bodies.

To allow for Buffered Peptone Water (BPW) to be used as an acceptable media to pre-moisten drag swabs, shoe cover, and chick liner paper samples for samples to be cultured within 48 hours of collection. To ensure that environmental samples are stored under refrigeration during transport and storage, added requirements for transporting environmental samples in a cooler with ice or ice packs for transport and refrigeration for storage. The coolers with ice or ice packs will avoid extreme temperatures and potential overgrowth of bacteria.

Sponsor: Dr. Doug Waltman, Georgia Poultry Laboratory Network
Dr. Carolyn Miller, Aviagen, Inc.

Program Standards - Proposal No. 5

Delegates: Combined

Standard B – Bacteriological Examination Procedures

~~(4) Procedure for bacteriological culturing of eggshells for colon bacilli organisms~~

~~Proper precautions to avoid environmental contamination of the samples during the collection and laboratory process, and proper handling of the samples following collection are essential. Each State Inspector involved in eggshell culture activities must receive instruction in the necessary sanitation procedures, sampling procedures, and sample handling by the authorized laboratory involved. The Official State Agency will maintain a record showing that the required instruction was given to each State Inspector.~~

- ~~(a) Sample selection. Forty eggs in the top flats of each of three randomly selected cases of sanitized eggs from each flock will be used for each sampling.~~
- ~~(b) Swab procedure. A 2.5 centimeter diameter circular area of the large end of each of the eggs will be rubbed with a sterile swab previously moistened with sterile lactose broth or other suitable liquid media provided by the authorized laboratory. One swab will be used for five eggs, and four swabs will be pooled to each sterile, capped tube provided by the authorized laboratory.~~
 - ~~(1) From the tube containing four swabs and lactose broth or other suitable media, 1 ml. will be transferred to 10 ml. lactose in a fermentation tube.~~
 - ~~(2) Incubate at 37 °C for 48 hours. The presence of acid, and gas in the amount of 10 percent or more after 24 and 48 hours of incubation, provides a presumptive conclusion of the presence of colon bacilli organisms.~~

Standard B – Bacteriological Examination Procedures

~~(5) Procedures to determine status and effectiveness of sanitation monitored program~~

~~The following monitoring procedures⁸ may be applied at the discretion of the Official State Agency:~~

- ~~(a) Monitor effectiveness of sanitation program~~
 - ~~(1) Culture the surface of cased eggs periodically for fecal contaminating organisms as described in Section B(4).~~
 - ~~(2) Culture a sample of dead in shell eggs periodically from each breeding flock for coliforms. Such eggs should also be cultured for the dependable recovery of salmonellae. Culturing for the dependable recovery of salmonellae should include the use of:~~
 - ~~(i) Preenrichment broths supplemented with 35 mg ferrous sulfate per 1,000 ml preenrichment to block iron binding, Salmonella inhibiting effects of egg conalbumin; and~~
 - ~~(ii) Tetrathionate selective enrichment broths, competitor controlling plating media (XLT4, BGN, etc.), delayed secondary enrichment procedures, and colony lift assays detailed in paragraph (a)(5) and illustration 2 of these Program Standards.~~

⁸ ~~Laboratory procedures for monitoring operations proposed here are described in the following two publications: Isolation and Identification of Avian Pathogens, American Association of Avian Pathologists, University of Pennsylvania, New Bolton Center, Kennett Square, Pennsylvania 19348-1692, 1980, and Culture Methods for the Detection of Animal Salmonellosis and Arizonosis, Iowa State University Press, Ames, Iowa 50010, 1976.~~

Reason: These procedures are historic (present in white book dated June 1985) in nature and are no longer used for “determining status and effectiveness of sanitation monitored program”.

Sponsor: Dr. Doug Waltman
Georgia Poultry Laboratory Network

Program Standards - Proposal No. 6

Delegates: Combined

Standard B – Bacteriological Examination Procedures

(8) Laboratory procedure recommended for the bacteriological examination of cull chicks and poults for *Salmonella*.

- (a) For cull chicks, from 25 randomly selected 1- to 5-day-old chicks that have not been placed in a brooding house, prepare 5 organ pools, 5 yolk pools, and 5 intestinal tissue pools as follows. For poults, from a sample of 10 poults that died within 10 days after hatching, prepare organ pools, yolk pools, and intestinal pools as follows:
 - (1) Organ pool: From each of five chicks or two poults, composite and mince 1- to 2-gram samples of heart, lung, liver, and spleen tissues. Include the proximal wall of the bursa of Fabricius for chicks only.
 - (2) Yolk pool: From each of five chicks or two poults, composite and mince 1- to 2-gram samples of the unabsorbed yolk sac or, if the yolk sac is essentially absent, the entire yolk stalk remnant.
 - (3) Intestinal pool: From each of five chicks or two poults, composite and mince approximately 0.5 cm² sections of the crop wall and 5-mm-long sections of the duodenum, cecum, and ileocecal junction.
- (b) Transfer each pool to tetrathionate selective enrichment broth (Hajna or Mueller-Kauffmann) at a ratio of 1 part tissue pool to 10 parts broth.
- (c) For cull chicks, repeat the steps in paragraphs (a) and (b) of this section for each 5-chick group until 25 chicks have been examined, producing a total of 15 pools (5 organ, 5 yolk, and 5 intestinal). For poults, repeat the steps in paragraphs (a) and (b) of this section for each two-poult group until all the poults in the sample have been examined.
- (d) Culture the tetrathionate pools as outlined for selective enrichment in illustration ~~2 1~~ of these Standards. Incubate the organ and yolk pools for 24 hours at 37 °C and the intestinal pools at ~~41-5~~ 42°C. Plate as described in illustration ~~2 1~~ of these Standards and examine after both 24 and 48 hours of incubation. Confirm suspect colonies as described. Further culture all *Salmonella*-negative tetrathionate broths by delayed secondary enrichment procedures described for ~~environmental, organ, and intestinal samples in illustration 2 1. A colony lift assay may also be used as a supplement to TSI and LI agar picks of suspect colonies.~~

Reason: This section currently is in Standard B, but is located after Mycoplasma methods. I would suggest moving this section into the section of Standard B that deals with *Salmonella*, for example before Standard B(4). Additionally, some modifications are proposed to clarify and update the procedure.

Sponsor: Dr. Doug Waltman
Georgia Poultry Laboratory Network

Program Standards - Proposal No. 7

Delegates: Combined

Standard C—Sanitation Procedures

(2) Hatching egg sanitation.

Hatching eggs should be collected from the nests at frequent intervals. ~~and, to aid in the prevention of contamination with disease-causing organisms, the~~ The following practices should be observed:

- (a) Cleaned and disinfected containers, such as egg flats, should be used in collecting the nest eggs for hatching and should be disinfected using products that are known to be effective against ~~Plan~~ disease agents, and/or are registered by the U.S. Environmental Protection Agency as effective against ~~Plan~~ disease agents. Egg handlers should thoroughly wash their hands with soap and water before and after egg collection. Clean outer garments should be worn.
- (b) Dirty eggs should not be used for hatching purposes and should be collected in a separate container from the nest eggs. Slightly soiled nest eggs may be gently dry cleaned by hand.
- (c) Hatching eggs should be stored in a designated egg room under conditions that will minimize egg sweating. The egg room walls, ceiling, floor, door, heater, and humidifier should be cleaned and disinfected after every egg pickup. Cleaning and disinfection procedures should be as outlined in Section C(4) of these Program Standards.
- (d) The egg processing area should be cleaned and disinfected daily.
- (e) Effective rodent and insect control programs should be implemented.
- (f) The egg processing building or area should be designed, located, and constructed of such materials as to ensure that proper egg sanitation procedures can be carried out, and that the building itself can be easily, effectively, and routinely sanitized.
- (g) All vehicles used for transporting hatching eggs ~~or chicks or poult~~ should be cleaned and disinfected after use. Cleaning and disinfection procedures should be as outlined in Section C(4).
- (h) Egg collection belts, tables, nest box pads and other egg collection equipment should be cleared of fecal material and managed on a regular basis to facilitate clean eggs.

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(3) Hatchery sanitation.

An effective program for the prevention and control of *Salmonella* and other infections should include the following measures:

- (a) An effective hatchery sanitation program should be designed and implemented.
- (b) The hatchery building should be arranged so that separate rooms are provided for each of the four operations: Egg receiving, incubation and hatching, chick/poult processing, and egg tray and hatching basket washing. Traffic and airflow patterns in the hatchery should be from clean areas to dirty areas (*i.e.*, from egg room to chick/poult processing rooms) and should avoid tracking from dirty areas back into clean areas.
- (c) The hatchery rooms, and tables, racks, and other equipment in them should be thoroughly cleaned and disinfected frequently. All hatchery wastes and offal should be burned appropriately managed and disposed of to prevent contamination of subsequent hatches. ~~or otherwise properly disposed of, and the containers~~ The equipment used to remove such materials should be cleaned and sanitized ~~after each use.~~
- (d) The hatching compartments of hatchers ~~incubators~~, including the hatching trays, should be thoroughly cleaned and disinfected after each hatch.
- (e) Only clean eggs should be used for hatching purposes.

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- (f) Only new or cleaned and disinfected egg cases or trays should be used for transportation of hatching eggs. Soiled egg case fillers should be destroyed.
- (g) Day-old chicks, poults, or other newly hatched poultry should be distributed in cleaned, or new boxes and new chick, or poult papers. All crates, lifting equipment, and vehicles used for transporting birds should be cleaned and disinfected after each use.

Reason: This proposal will update hatchery sanitation practices in Subpart C(2) Hatching egg sanitation and Subpart C(3) Hatchery sanitation that are currently employed by NPIP Participants. The proposed changes will allow flexibility for industry practices that are and have been commonly employed for many years.

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Dr. Katie Schlist, Forsman Farms, Howard Lake, MN
Dr. Dale Lauer, Minnesota Board of Animal Health, Willmar, MN

Program Standards - Proposal No. 8

Delegates: Combined

Standard C—Sanitation Procedures

(4) Cleaning and disinfecting

The following procedures are recommended:

(a) ~~In the p~~Poultry houses:

- (1) Remove all ~~live “escaped” and dead~~ birds from the building. Blow dust from equipment and other exposed surfaces. Empty the residual feed from the feed system and feed pans and remove it from the building. As appropriate, Disassemble feeding equipment and dump and scrape as needed to remove ~~any and all~~ feed cake and residue. Clean up spilled feed around the ~~tank~~ bulk feed bins and physically clean out ~~the tank~~ if possible. After dry cleaning of the inside of feed bins to remove any residual build-up of feed it may be beneficial to Rinse down and wash out the inside of the feed ~~tank~~ bins to decontaminate the surfaces and allow to completely dry.
- (2) The company and/or site specific biosecurity plan will detail the appropriate insect and rodent control.
- ~~(2)~~ (3) If litter is to be removed, ~~r~~Remove all litter and manure droppings to an isolated area where there is no opportunity for dissemination of any Plan disease agents that may be present. Housing where poultry infected with a mycoplasmal disease were kept should remain closed for 7 days before removal of the litter.
- ~~(3)~~ (4) When indicated for control of Plan disease agents, a wash down using clean water should be used, ~~avoiding untreated pond or stream water.~~ Wash down Washing the entire inside surfaces of the building and all the installed equipment such as curtains, ventilation ducts and openings, light traps and openings, fans, fan housings and shutters, feeding equipment, watering equipment, etc. shall be performed. Use high-appropriate pressure and high-volume of water spray (for example 200 pounds per square inch and 10 gallons per minute or more) to soak into and remove the dirt to decontaminate the building. Wash the walls, floors, and equipment with a soapy water solution. Rinse to remove soap. Pay specific attention to the area linking of side walls with building floors and or stem walls to remove all accumulated organic material. Make sure to close up any drain caps and doorways when building is not actively being worked on at all times during the cleaning process. Make sure any chemical cleaning and disinfecting agents used in the cleaning process are agents known to be effective against Plan disease agents and/or are registered by the Environmental Protection Agency as effective against Plan disease agents.
- (5) Perform any mechanical or physical maintenance on buildings and/or equipment necessary including patching up any wild bird or obvious rodent entry points.
- ~~(4)~~ (6) After washing is complete, ~~S~~spray with a disinfectant ~~which that~~ is known to be effective against Plan disease agents and/or is registered by the Environmental Protection Agency as effective against Plan disease agents, applying as germicidal, fungicidal, pseudomonocidal, and tuberculoecidal, in accordance with the specifications for use, as shown on the label of such disinfectant.
- (7) Make sure any building end pad areas are completely cleaned and free of organic material from the previous flock prior to adding new bedding or other supplies, birds, or equipment.

(b) In the hatchers and hatchery rooms:

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- (1) Use cleaning agents and sanitizers that are known to be effective against ~~Plan~~ disease agents and/or are registered by the U.S. Environmental Protection Agency as effective against Plan disease agents, as germicidal, fungicidal, pseudomonocidal, and tuberculocidal. Use manufacturer's recommended dilution rates. Remove loose organic debris ~~by sweeping, scraping, vacuuming, brushing, or scrubbing, or by hosing surface with high pressure water (for example 200 pounds per square inch and 10 gallons per minute or more)~~. Remove trays and all controls and fans for separate cleaning. Use hot water (minimum water temperature of 140 °F) for cleaning hatching trays and chick separator equipment. Thoroughly wet the ceiling, walls, and floors ~~with a stream of water~~, then scrub with a ~~hard-bristle~~ brush. Use a cleaner/sanitizer that can penetrate protein and fatty deposits. Allow the appropriate contact time per the manufacturer's recommendations ~~chemical to cling to treated surfaces at least 10 minutes before rinsing off~~. Manually scrub any remaining deposits of organic material until they are removed. Rinse until there is no longer any deposit on the walls, particularly near the fan opening, ~~and apply disinfectant~~. Use a clean and sanitized squeegee to remove excess water, working down from ceilings to walls to floors, ~~and being careful not to recontaminate cleaned areas~~. Apply disinfectant per the manufacturer's recommendations.
- (2) Replace the cleaned fans and controls. Replace the trays, ~~preferably still wet from cleaning~~, and bring the incubator to normal operating temperature.
- (3) The hatcher should be ~~fumigated (see Section C(5)) or otherwise~~ disinfected before transferring the eggs.
- (4) If the same machine is used for incubating and hatching, the entire machine should be cleaned after each hatch. A vacuum cleaner should be used to remove dust and down from the egg trays; then the entire machine should be vacuumed, mopped, and ~~fumigated (see Section C(5)) or otherwise~~ sanitized.

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(c) The egg and chick/poult delivery truck drivers and helpers should use the following good biosecurity practices while picking up eggs or delivering chicks or poults:

- (1) ~~Spray truck tires thoroughly with disinfectant before leaving the main road and entering the farm driveway.~~
- (2) ~~Put on sturdy, disposable plastic boots or clean rubber boots before getting out of the truck cab. Put on a clean smock or coveralls and a hairnet before entering the poultry house. Personnel that are entering egg rooms, or poultry ready facilities should take precautions, including washing of and or sanitation of hands, and wearing of premises specific clothing and footwear according to the company and/or site specific biosecurity plan.~~
- (3) After loading eggs or unloading chicks or poults, remove the dirty premises specific clothing and footwear (to leave at the facility), or smock or coveralls and place into a plastic garbage bag before loading in the truck. Be sure to keep clean clothing and footwear ~~coveralls~~ separate from dirty ones. Remove hairnet and disposable boots (if applicable) and discard at the farm.
- (4) ~~Re-enter the cab of the truck and remove boots before placing feet onto floorboards. Remove hairnet and leave with disposable boots on farm.~~
- (5) Sanitize hands using appropriate hand sanitizer.
- (6) ~~Re-enter the truck to~~ Return to the hatchery or go to the next farm and repeat the process.

Reason: This proposal will update poultry house and hatchery sanitation practices in Subpart C(4) Cleaning and disinfection that are commonly employed by NPIP Participants. The proposed changes will allow flexibility for some industry practices that are and have been commonly employed for many years.

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Dr. Dale Lauer, Minnesota Board of Animal Health, Willmar, MN

Program Standards Proposal No. 9

Delegates: Combined

Standard C – Sanitation Procedures

(9) Dealer sanitation

As applicable, a recommended program for control of *Salmonella* and other ~~Plan~~ disease agents:

(a) Hatching eggs

- (1) Accept hatching eggs from NPIP participants that follow the appropriate sanitation procedures outlined in Program Standards, Standards C – Sanitation Procedures.
- (2) Prior to movement refer to and follow the company and/or site specific biosecurity plan.

(b) Newly hatched poultry

- (1) Provide a pen location that can be cleaned and sanitized on a regular basis, in a manner acceptable to the Official State Agency.
- (2) Provide adequate clean bedding that is removed on a regular basis in a manner acceptable to the Official State Agency.
- ~~(3) Provide an adequate heat source with ventilation managed to optimize the well being of the newly hatched poultry.~~
- (4) Provide sufficient feed and water in containers that are cleaned and sanitized on a regular basis, in a manner acceptable to the Official State Agency.
- (5) Maintain a physical barrier to prevent the public from having direct contact with the newly hatched poultry.
- (6) Make available a wash station or hand sanitizer nearby the poultry display areas.
- (7) Provide information to customers about preventative public health measures regarding *Salmonella* in live poultry.

(c) Started poultry

- (1) Accept poultry from NPIP Participants that have followed the appropriate sanitation procedures detailed in Program Standards, Standards C – Sanitation Procedures.
- (2) Prior to movement refer to and follow the company and/or site specific biosecurity plan.

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Reason: This proposal will create a new Dealer sanitation standard that can be recommended, followed and used for the control of *Salmonella* and other program diseases.

Sponsor: Dr. Dale Lauer, Minnesota Board of Animal Health, Willmar, MN

Program Standards - Proposal No. 10

Delegates: Combined

Standard C—Sanitation Procedures

(1) Flock sanitation.

To aid in the maintenance of healthy flocks, the following procedures should be practiced:

- (a) ~~Baby poultry~~ Poultry should be started in a ~~clean brooder~~ house managed to reduce or eliminate exposure to Plan disease agents, and ~~Flocks should be housed~~ maintained in ~~constant physical~~ isolation from older birds and other animals. Personnel that are ~~in contact with older birds and other animals~~ crossing the Line of Separation (LOS) should take precautions, including ~~disinfection of footwear and change of outer clothing~~ washing and or sanitation of hands, and wearing premises specific clothing and footwear, ~~to prevent the introduction of infection through droppings that may adhere to the shoes, clothing, or hands.~~ (See Section C(4)(a).)
- (b) ~~Range used for growing young stock should not have been used for poultry the preceding year.~~ Where ~~broods~~ flocks of different ages must be kept on the same farm, there should be complete depopulation ~~of brooder houses and other premises~~ following ~~infection~~ contamination of such premises by any ~~contagious~~ Plan disease that causes the existence of a carrier population or a reservoir in the environment.
- (c) Poultry houses ~~should~~ should be screened and secured against free-flying ~~wild birds~~ with ~~An active rodent eradication campaign and insect vector eradication /control programs as defined in the company and/or site specific biosecurity plan.~~ is an essential part of the general sanitation program. The area adjacent to the poultry house ~~should~~ should be kept free from accumulated manure, rubbish, and unnecessary equipment. Vegetation surrounding all poultry housing shall be excluded from or minimized in amount for at least three feet distance to facilitate control of vermin. Animals not participating in the Plan should ~~not cross the Line of Separation (LOS).~~ never have access to poultry operations. Visitors should not be admitted to poultry areas, and authorized personnel should take the necessary precautions to prevent the introduction of Plan disease in accordance with the company and/or site specific biosecurity plan.
- (d) Poultry houses and equipment should be thoroughly cleaned and disinfected prior to use for a new lot of birds. (See Section C(4)(a).) ~~Feed and water containers should be situated where they cannot be contaminated by droppings and should be frequently cleaned and disinfected. Dropping boards or pits should be constructed so birds do not have access to the droppings.~~ Prior to the placement of a new flock, ~~the company and/or site specific biosecurity plan should detail the procedures in place to minimize the risk of~~ Plan disease introduction and transmission from premises sanitation, feed, replacement litter and water supplies.
- (e) ~~Replacement breeders shall be housed at the proper density consistent with the type of building and locality and which will allow the litter to be maintained in a dry condition. Frequent stirring of the litter may be necessary to reduce excess moisture and prevent surface accumulation of droppings. Slat or wire floors should be constructed so as to permit free passage of droppings and to prevent the birds from coming in contact with the droppings. Nesting areas should be kept clean and, where appropriate, filled with clean nesting material.~~ Management of ventilation systems should be done in a manner to optimize moisture removal and reduce excess moisture, dust and ammonia. Nesting areas should be kept clean, dry and free of fecal material.
- (f) When an outbreak of a Plan disease occurs in a flock, every effort should be made to identify the causative agent. ~~dead or sick birds should be taken, by private carrier, to a diagnostic~~

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laboratory for complete examination. All Salmonella cultures isolated should be typed serologically, as appropriate to determine specific control measures. ~~and complete records maintained by the laboratory as to types recovered from each flock within an area. Records on isolations and serological types should be made available to Official State Agencies or other animal disease control regulatory agencies in the respective States for followup of foci of infection. Such information is necessary for the development of an effective Salmonella control program.~~

- (g) ~~Introduction~~ Placement of started or mature birds should be ~~avoided~~ managed to reduce the possible hazard of introducing introduction of Plan disease agents. ~~If~~ When birds are to be placed introduced, the health status of ~~both the flock and the newly placed introduced~~ birds should be evaluated with recent test results for applicable Plan disease agents prior to movement.
- (h) ~~In rearing broiler or replacement~~ all poultry stock, a sound and adequate immunization program, as advised by a poultry health professional, should be adopted. ~~Since different geographic areas may require certain specific recommendations, the program recommended by the State experiment station or other State agencies should be followed.~~
- (i) ~~Feed, pelleted by heat process,~~ should be fed to all age groups produced and treated to prevent transmission of Plan disease agents by heating or approved chemical treatment. ~~Proper feed pelleting procedures can destroy many disease producing organisms contaminating feedstuffs.~~

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Reason: This proposal will update flock sanitation practices in Subpart C (1), flock sanitation measures commonly employed by NPIP Participants. The proposed changes will allow flexibility for industry practices that are and have been used for many years.

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Dr. Katie Schlist, Forsman Farms, Howard Lake, MN
Dr. Dale Lauer, Minnesota Board of Animal Health, Willmar, MN

Program Standards - Proposal No. 11

Delegates: Combined

Standard D—Molecular Examination Procedures

(7) Approved tests

The following diagnostic test kits that are not licensed by the Service (e.g., bacteriological culturing kits) are approved for use in the NPIP:

1. Rapid Chek©Select TMSalmonella Test Kit, Romer Labs, Inc., Newark, DE 19713.
2. ADIAFOOD Rapid Pathogen Detection System for *Salmonella* spp., AES Chemunex Canada. Laval, QC (Canada) H7L4S3.
3. DuPont Qualicon BAX Polymerase Chain Reaction (PCR)-based assay for *Salmonella* 1 and 2 DuPont Qualicon, Wilmington, DE 19810.
4. Applied Biosystems TaqMan® *Salmonella* Enteritidis Real-Time PCR assay for the detection of *Salmonella* Enteritidis. Life Technologies Corporation. Foster City, CA 94404.
5. IDEXX MG/MS RT-PCR.
6. MicroSEQ Salmonella Species Detection Kit, Life Technologies Corporation, Austin, TX.
7. ANSR Salmonella Test, Neogen Corporation, Lansing, MI 48912.
8. Reveal 2.0 Group D1 Salmonella (Including SE) Kit, Neogen, Neogen Corporation, Lansing, MI 48912.
9. DNable Salmonella Detection Kit, EnviroLogix, Inc., Portland, Maine 04103.
10. Bactotype MG/MS Kit, QIAGEN, Germantown, Maryland, 20874.
11. IDEXX RealPCR MG DNA reagents-IDEXX Laboratories, Inc. Westbrook, ME 04092.
12. IDEXX RealPCR MS DNA reagents-IDEXX Laboratories, Inc. Westbrook, ME 04092.
13. IDEXX RealPCR MG-MS Multiplex DNA reagents-IDEXX Laboratories, Inc. Westbrook, ME 04092.
14. Poultry Check MP MS-MG Test Kit-Biovet, Inc. St. Hyacinthe, Quebec J2S 8W2 Canada.
15. Thermo Fisher Scientific MG/MS Reagents-Thermo Fisher Scientific, Life Sciences Solutions, Austin, TX 78744.
16. Qiagen *mericon* ® Salmonella spp. real-time PCR kit-Qiagen, Germantown, MD 20874.
17. IDEXX RealPCR Salmonella DNA spp. DNA reagents- IDEXX Laboratories, Inc. Westbrook, ME 04092.
18. ~~BioChek Salmonella Species PCR-BioChek, 3 Southgate Rd, Scarborough, ME 04074.~~
19. ~~BioChek MgMs qPCR test BioChek, 3 Southgate Rd, Scarborough, ME 04074.~~

Program Standards Proposal No. 12

Delegates: Combined

Standard E – Biosecurity Principles

Based on the flock size as stated in the 9 CFR 53.10, and including breeding flock premises with at least 5000 birds, the following minimum management practices and principles are designed to prevent the introduction and spread of infectious diseases.

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(14) Auditing

Auditing of the biosecurity principles is based on flock size as outlined in 9 CFR 53.10, and shall include breeding flock premises with at least 5000 birds. Audits shall be conducted at least once every two years or a sufficient number of times during that period by the Official State Agency to ensure the participant is in compliance. Each audit shall require the biosecurity plan's training materials, documentation of implementation of the NPIP Biosecurity Principles, corrective actions taken, and the Biosecurity Coordinator's annual review to be audited for completeness and compliance with the NPIP Biosecurity Principles. An audit summary report containing satisfactory and unsatisfactory audits will be provided to the NPIP National Office by the OSAs.

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Those participants who failed the initial document audit conducted by the NPIP OSA may elect to have a check audit performed by a team appointed by National NPIP Office including: an APHIS poultry subject matter expert, the OSA, and a licensed, accredited poultry veterinarian familiar with that type of operation. If these participants seek to be reinstated as being in compliance with the Biosecurity Principles by the NPIP OSA, they must demonstrate that corrective actions were taken following the audit by the team appointed by NPIP.

Reason: There are several reasons supporting these changes to the program standards. The original intention of the biosecurity principles was to increase the minimum biosecurity requirements for all commercial industry, including breeding complexes. The current language, referencing section 53.10 of the 9 CFR, unintentionally exempts all breeding complexes from the auditing requirement. While most companies have no issues complying across the board with the audits, some OSAs have already reported push back on the auditing requirement from a few companies, thus prompting the need to revisit the language. If we are going to hold the poultry industry to a specific standard, especially when indemnity funds can potentially become involved, then all aspects of commercial production should be held to that standard equally. Additionally, as recent HPAI outbreaks have demonstrated, there is a larger risk for long-lived birds to contract Avian Influenza, thus it's counterintuitive to only be auditing the biosecurity program of broiler complexes, and not the parent stock. All birds are at risk for contracting avian diseases, therefore all aspects of commercial industry should adhere to the same standard, and would benefit from these biosecurity principles. Finally, one would expect the breeding stock of commercial broilers and turkeys to have higher biosecurity standards than their progeny as typical industry practice, thus this change should not create an increased burden.

Sponsor: Dr. Melissa Yates
Arkansas Livestock and Poultry Commission

****Pre-Conference Edition****

Proposed Changes Booklet
to be considered at the
NPIP 44th Biennial Conference



COMPARTMENTALIZATION



FRANKLIN MARRIOTT COOL SPRINGS HOTEL
JUNE 26-28, 2018

****Pre-Conference Edition****

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**COMPARTMENTALIZATION FOR
PROTECTION AGAINST Avian
Influenza and Newcastle DISEASE IN
PRIMARY POULTRY BREEDING
COMPANIES IN THE UNITED STATES
OF AMERICA**

Specifications For:



United States Department of Agriculture ~ National Poultry Improvement Plan

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Additional Information

[9 CFR Part 145 Subpart G—Special Provisions for Primary Egg-Type Chicken Breeding Flocks and Products](#)

[9 CFR Part 145 Subpart H—Special Provisions for Primary Meat-Type Chicken Breeding Flocks and Products](#)

[9 CFR Part 145 Subpart D—Special Provisions for Turkey Breeding Flocks and Products](#)

[NPIP Program Standards Document](#)

[GSA FY Per-Diem Allowance](#)

[EPA Registered and Licensed Disinfectants](#)

Historical Background

The USDA-APHIS-National Poultry Improvement Plan (NPIP) is a disease surveillance and control program for the U.S. poultry industry. The NPIP was established to help control existing diseases incompatible with the growth and development of a modern poultry industry. APHIS added the avian influenza (AI) programs for breeding chickens and breeding turkeys to the NPIP in the 1990s. Prior to this time, only vertically transmitted diseases (*Salmonella Pullorum*, *Salmonella Gallinarum*, *Salmonella Enteritidis*, *Mycoplasma gallisepticum*, *Mycoplasma synoviae*, and *Mycoplasma meleagridis*) were included in the NPIP. However, when the poultry industry began to export large quantities of poultry genetic stock and poultry meat and eggs, major U.S. trading partners wanted assurances that the poultry and poultry products originated from breeding flocks free of AI. H5/H7 AI monitoring programs for commercial table-egg layers, broilers, and meat turkeys were added to the NPIP in 2006.

Today, the NPIP continues to provide assurance that poultry and poultry products originating in the United States are free of AI. Compliance with NPIP standards and subsequent Federal endorsement is required for interstate and international sale and distribution of commercial poultry breeding stock.

Introduction

Regionalization is a procedure a country may implement to manage animal populations confined to a distinct geographical region within its territory for the purpose of disease control and international trade. In the event of a disease occurrence within a specific region, compartmentalization may become an option to maintain trade.

Compartmentalization is a procedure a country may implement to define and manage animal subpopulations of distinct health status and common biosecurity program within its territory, in accordance with the guidelines in the World Organization for Animal Health (OIE) Terrestrial Animal Health Code (hereinafter "Code"), for the purpose of disease control and international trade. Concepts of regionalization and compartmentalization are not mutually exclusive.

A compartment may be established with respect to a specific disease or diseases. A compartment should be clearly defined, indicating the location of all its components, including establishments as well as related functional units (such as feedmills, slaughter houses, rendering plants, etc.), their interrelationships, and their contribution to an epidemiological separation between the animals in a compartment and subpopulations with a different health status. The definition of a compartment may revolve around disease-specific epidemiological factors, animal production systems, biosecurity practices, infrastructural factors and surveillance. (Code, Chapter 4.4. – Application of Compartmentalization).

The current control and surveillance programs for participants for AI in the United States are AI Clean and H5/H7 AI Clean and can be found in the Title 9, Code of Federal Regulations (9 CFR) 145.43(g) (turkey breeding flocks), 145.73(f) (primary egg-type chicken breeding flocks), and 145.83(g) (primary meat-type chicken breeding flocks). The regulations at 9 CFR 145.45, 9 CFR 145.74, and 9 CFR 145.84 provide the basis for compartmentalization of poultry primary breeding companies.

The U.S. Avian Influenza [and Newcastle Disease](#) Clean Compartment and program is intended to allow the primary egg-type chicken (9 CFR 145.74(a)) and primary meat-type chicken (9 CFR 145.84(a)) breeding-hatchery industry to demonstrate the existence and implementation of a program approved by the Official State Agency (OSA) and APHIS to establish a compartment consisting of a primary breeding-hatchery company free of H5/H7 AI, ~~also referred to as notifiable avian influenza (NAI)~~ [and Newcastle disease](#). This compartment protects the defined subpopulation and avoids the introduction and spread of ~~NAI~~ [and ND](#) within that subpopulation by prohibiting contact with other commercial poultry operations, other domestic and wild birds, and other intensive animal operations.

The U.S. H5/H7 Avian Influenza [and Newcastle Disease](#) Clean Compartment program is intended to allow the primary turkey (9 CFR 145.45(a)) breeding-hatchery industry to demonstrate the existence and implementation of a program approved by the OSA and APHIS to establish a compartment consisting of a primary breeding-hatchery company free of H5/H7 AI (~~NAI~~) [and Newcastle disease](#). This compartment protects the defined subpopulation and avoids the introduction and spread of ~~NAI~~ [and ND](#) within that subpopulation by prohibiting contact with other commercial poultry operations, other domestic and wild birds, and other intensive animal operations.

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Compartment Oversight

APHIS Veterinary Services National Import Export Services (NIES) will provide technical advice regarding international animal health standards and export risk mitigation to compartment program managers and participants. NIES also advocates for compartmentalization participants to build relationships with animal health and regulatory counterparts in other countries, explaining the program to foreign officials and developing bilateral and multilateral agreements with trading partners to accept imports of poultry from compartment participants.

The primary breeder company will define the compartment with respect to [NAI and ND](#). Specifically, the company will use a comprehensive biosecurity program to define the compartment as a subpopulation of poultry with an [NAI and ND](#) health status separate from birds and poultry outside the compartment. The OSA and APHIS must first approve all documentation submitted by the company to substantiate the defined compartment as adequate to qualify for epidemiological separation from other potential sources of [NAI and ND](#) infection.

Compartment Requirements

1. A participant in good standing with the NPIP in ~~one~~ two of the following programs:
 - [U.S. Newcastle Disease Clean Program for Turkey Breeding Flocks \(9 CFR 145.43\)](#).
 - U.S. H5/H7 Avian Influenza Clean Program for Turkey Breeding Flocks (**9 CFR 145.453**).
 - [U.S. Newcastle Disease Clean Program for Primary Egg-Type Chicken Breeding Flocks \(9 CFR 145.73\)](#).
 - U.S. Avian Influenza Clean Program for Primary Egg-Type Chicken Breeding Flocks (**9 CFR 145.743**).
 - [U.S. Newcastle Disease Clean Program for Primary Meat-Type Chicken Breeding Flocks \(9 CFR 145.83\)](#).
 - U.S. Avian Influenza Clean Program for Primary Meat-Type Chicken Breeding Flocks (**9 CFR 145.843**).
2. Compliant with all of the management procedures, physical requirements, and protocols found in this document, the *Code of Federal Regulations*, and the NPIP Program Standards document.
 - [NPIP Provisions](#)
 - [Program Standards document](#)
3. Located in a State or States with an APHIS-approved Initial State Response and Containment Plan (**9 CFR 56.10**).
4. Perform routine surveillance of all flocks within the compartment in a NPIP-authorized laboratory certified to test for AI and ND.
5. [Flocks within the compartment may be vaccinated with a USDA licensed Newcastle disease vaccine or may be unvaccinated for Newcastle disease. All flocks have a routine serological monitoring program in place to monitor antibody response or freedom from ND if flocks are unvaccinated.](#)
 - **For unvaccinated flocks:**
 - [It is a primary breeding flock in which a minimum of 30 birds have been tested negative for ND using an approved test when more than 4 months of age. To retain this classification:](#)
 - [A sample of at least 30 birds must be tested negative at intervals of 90 days; OR](#)
 - [A sample of fewer than 30 birds may be tested, and found negative, at any one time if all pens are equally represented and a total of 30 birds is tested within each 90-day period; AND](#)
 - [During each 90-day period, all primary spent fowl, up to a maximum of 30, must be tested negative to ND within 21 days prior to movement to slaughter.](#)
 - **For vaccinated flocks:**
 - [It is a primary breeding flock that has been vaccinated with licensed vaccines, as described in §113.329, manufactured with low-virulence live strains during early stages of development up to grow-out, and killed vaccines as final vaccination prior to onset of egg production; AND](#)
 - [The flock has been monitored for antibody response using approved serological tests as described in §145.14 and the results are compatible with immunological response against ND vaccination; AND](#)

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- Testing must include:
 - a minimum of 30 birds when birds are more than 4 months of age and not longer than every 90 days thereafter.

5.6. All companies participating in the compartment must be able to provide the following general management protocols (GMP) on request:

GMP 1. Biosecurity training for employees, contract staff, and visitors.

GMP 2. Biosecurity compliance agreement for employees, contract staff, and visitors.

GMP 3. Biosecurity risk assessment for each component of the compartment.

GMP 4. Cleaning, sanitation, and control of vehicles prior to entering biosecure areas.

GMP 5. General physical traits of each compartment component.

GMP 6. Detailed diagrammatic description for movement of people, vehicles, equipment, birds, and eggs between all components inside and outside the compartment.

GMP 7. Company Emergency Response Plan.

GMP 8. Veterinary Health Plan.

GMP 9. ND Vaccination Program if applicable.

GMP 10. ND Serological Monitoring Program for ND vaccinated or unvaccinated flocks.

Compartment Application Process

To apply initially as a compartment, a company should complete and submit Application Form A: Compartment Registration and Application Form B: Component Registration. After Application Form A is reviewed and signed by the OSA and approved by the NPIP National Office, Application Form B will be reviewed. Once Application Form B has been reviewed and signed by the OSA and approved by the NPIP, an auditor is assigned. The auditor will assess and inspect all components. If all components pass inspection, NPIP will notify the company of the compartment certification and the list of certified components within the compartment. The company will also receive an official U.S. Avian Influenza [and Newcastle Disease](#) Clean Compartment certificate. For initial registration, each component within the compartment will be inspected by a certified auditor. Re-certification of components must take place every year, and the compartment is subject to audits of components specified by APHIS.

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Compartment Auditing Process

Auditing and oversight of compartments is a key element of the program. NIES will oversee the auditing process. After approval of the documentation submitted, a certified auditor assigned by the NPIP office will conduct an initial audit and inspection of both the office and field sites. Every component within the compartment will be subject to this audit. The compartment will only be approved after successful completion of the initial inspection and audit. All hatcheries, feedmills, and egg depots in approved compartments will be audited annually, and 25 percent of the farm components will be subject to annual audits. NIES will conduct a Compartmentalization Service Review every 4 years, examining all aspects of the program.

The auditing process ensures a successful compartment. For the companies involved, the process includes submission of an application, both office and field audits conducted by a certified auditor, NPIP reviews, recognition and approval of each component within the compartment, and re-qualification. Use of certified auditors ensures a successful process. A certified auditor is one who has met the requirements listed below:

- Must attend and successfully complete an official USDA-NPIP Auditor Compartment Training Course prior to conducting any audits, and become recertified at least once every 4 years thereafter.
- Must operate and conduct oneself with the highest code of ethics and must not have a conflict of interest with any of the companies which are compartmentalized or seeking compartment certification.
- Must be a U.S. licensed and accredited veterinarian who is board certified by the American College of Poultry Veterinarians (ACPV) and meets contract requirements set forth by APHIS, or must be a Federal Veterinary Medical Officer (VMO), preferably one with poultry experience.

The purpose of the NPIP Auditor Compartment Training Course is threefold: To familiarize the auditors with the contents of this document as well as the official audit checklist of items and equip them to perform audits accurately and consistently, including conducting mock audits at farm, hatchery, feedmill, egg depot, and office sites; to expose auditors to the primary breeder industry and continually educate auditors on pertinent operational activities and important updates in technology within the poultry industry; and to emphasize the code of ethics in operating as a certified auditor for the U.S. Avian Influenza [and Newcastle Disease](#) Clean Compartment Program. All auditors must pass an examination at the end of the Auditor Compartment Training Course to earn certified status.

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Compartment Suspension

The auditing process may highlight the need for certain components within certified compartments to correct deficiencies that could compromise the integrity of the certified compartment. If a component is found to have a minor noncompliance, the issue will be listed within the audit as requiring corrective action and the company given time deemed appropriate by the auditor to correct the problem. The auditor will revisit the component after the specified time to verify that the problem is fixed. If the company fails to correct the problem within the given time, the entire certified compartment will be suspended. If a minor noncompliance is found, documented, and not fixed within the specified time during the initial audit for a component seeking certification within the prospective compartment, that component within the prospective compartment will not be granted certification and must wait 30 days before re-applying using Application Form B.

If at any time a component within a certified compartment is found to have a major noncompliance, the entire compartment will be suspended immediately. Examples of major noncompliances include: 1) loss of NPIP U.S. Avian Influenza Clean status (for meat-type and egg-type breeders) or loss of the NPIP U.S. H5/H7 Avian Influenza Clean status (for turkey breeders) by failure to adequately test or by National Veterinary Services Laboratory (NVSL)-confirmed detection of HPAI in the certified compartment; 2) failure to renew certification on time; 3) failure to satisfactorily remediate and apply appropriate, effective corrective measures to any documented minor non-compliance offenses; 4) [loss of NPIP U.S. Newcastle Disease Clean status by failure to adequately test or by confirmed detection of ND in the certified compartment.](#)

To regain certified compartment status after a compartment suspension, the entire suspended compartment, including each component within that compartment, must wait 30 days and then re-apply using Application Form A and Application Form B.

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Definitions

Animal and Plant Health Inspection Service: The Animal and Plant Health Inspection Service of the U.S. Department of Agriculture.

Authorized laboratory: Laboratory that meets the requirements of 9 CFR 147.52.

Avian influenza: An infection of poultry caused by any influenza A virus of the H5 or H7 subtypes or by any influenza A virus with an intravenous pathogenicity index (IVPI) greater than 1.2 (or as an alternative at least 75 percent mortality).

Auditor: An individual who has successfully met all requirements and is certified to conduct audits for U.S. AI [and ND](#) Clean and U.S. H5/H7 AI [and ND](#) Clean Compartments.

Biosecure zone: Zone of the compartment premises to which high biosecurity standards apply for the disease of concern. All personnel must undergo a whole body shower and change of clothing and footwear prior to entering the biosecure zone. A biosecure zone barrier must define the limits of the biosecure zone. The biosecure zone may include multi-age and multi-building premises in which personnel, visitors, and contractors follow all company-established procedures.

Biosecure zone barrier: Contains all or portions of the external walls of buildings or geographic structures that discourage human and animal traffic. Permanent structures that may consist of, at minimum height, 4-foot chain link fences that form the perimeter of and totally enclose the biosecure zone are adequate.

Chicks: Young poultry less than 72 hours from hatch.

Classification: A designation earned by participation in a Plan program.

Company-established protocols/procedures/policies: Written guidelines developed and implemented by companies to maintain applicable NPIP classification for AI programs and to meet U.S. AI Clean compartmentalization requirements.

Compartmentalization: A procedure which may be implemented by a country to define and manage animal subpopulations of distinct health status within its territory, in accordance with the recommendations in the OIE Terrestrial Animal Health Code (the Code), for the purpose of disease control and/or international trade.

Component: Any farm, feedmill, hatchery, or egg depot that will be included in a compartment.

Contractor: Third-party agent who performs a specific task or service for a compartment company. These agents are obligated to meet biosecurity requirements specified by the compartment company.

Controlled access zone: Area surrounding the biosecure zone which only authorized personnel or vehicles may enter. Unauthorized personnel, vehicle traffic, and livestock are not permitted within the controlled access zone. A gate is required and signage indicating that unauthorized entry is prohibited must be posted at the entrance to this zone.

Department: The United States Department of Agriculture.

Egg Depot: Temporary egg storage and holding facility.

Equivalent requirements: Requirements which are equal to or exceed the program, conditions, criteria, or classifications with which they are compared.

Farm: Area of land and associated buildings dedicated to housing and rearing poultry breeding stock.

Feedmill: Facility for manufacturing, storing, and distributing feed.

Flock: (1) As applied to breeding: All poultry of one kind of mating (breed and variety or combination of stocks) and of one classification on one farm; (2) As applied to disease control: All of the poultry on one farm, except that any group of poultry which is segregated from another group and has been so segregated for a period of at least 21 days may be considered as a separate flock.

Hatching egg: Fertilized poultry egg.

High Pathogenicity Avian Influenza (HPNAI)-virus: Virus having an intravenous pathogenicity index in 6-week-old chickens greater than 1.2 or, as an alternative, causes at least 75 percent mortality in 4-to 8-week-old chickens infected intravenously. H5 and H7 viruses which do not have an intravenous pathogenicity index of greater than 1.2 or that cause less than 75 percent mortality in an intravenous lethality test should be sequenced to determine whether multiple basic amino acids are present at the cleavage site of the hemagglutinin molecule (HA0); if the amino acid motif is similar to that observed for other high pathogenicity AI isolates, the isolate being tested should be considered as highly pathogenic high pathogenicity AI virus.

Hatchery: Facility where eggs are temporarily stored, incubated, hatched, and distributed.

High-risk period: When AI is reported in a State or within a 30-mile radius of a compartment facility. The high-risk period ends when any control zones are released.

Livestock: Farm animals (such as cows, horses, sheep, goats, pigs, etc.) kept, raised, and used by people.

Livestock fence: Permanent structure serving as a barrier to restrict access of livestock to a compartment facility.

Low Pathogenicity Notifiable Avian Influenza (LPNAI): All influenza A viruses of H5 and H7 subtype that are not HPNAI viruses.

Low risk period: When AI is not in the State or within a 30-mile radius of a compartment facility.

Multi-age premises: Premises where birds are of different ages present.

Multi-building premises: Premises where there is more than one house. Multi-building premises may also be operated as multi-age premises.

National Poultry Improvement Plan: A voluntary Federal disease control program for the poultry

industry in the United States. Established in the early 1930's to provide a cooperative industry, State, and Federal program through which new diagnostic technology can be effectively applied to improve poultry and poultry products throughout the country.

Newcastle Disease: [Newcastle disease is defined by OIE for reporting an outbreak of ND as an infection of poultry caused by a virus of avian paramyxovirus serotype 1 \(APMV-1\) that meets one of the following criteria for virulence: a. The virus has an intracerebral pathogenicity index \(ICPI\) in day-old chicks \(Gallus gallus\) of 0.7 or greater. Or b. Multiple basic amino acids have been demonstrated in the virus \(either directly or by deduction\) at the C-terminus of the F2 protein and phenylalanine at residue 117, which is the N-terminus of the F1 protein. The term 'multiple basic amino acids' refers to at least three arginine or lysine residues between residues 113 and 116. Failure to demonstrate the characteristic pattern of amino acid residues as described above would require characterization of the isolated virus by an ICPI test.](#)

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NPIP Program Standards: A document that contains tests and sanitation procedures approved by the Administrator in accordance with 9 CFR 147.53. This document may be obtained from the [NPIP website](#) or by writing to the National Poultry Improvement Plan, APHIS, USDA, 1506 Klondike Road, Suite 101, Conyers, GA 30094.

Official State Agency: The State authority recognized by the Department to cooperate in administering the Plan.

Official Veterinarian: Veterinarian employed or contracted by the Department.

Plan: The provisions of the National Poultry Improvement Plan.

Poultry: Domesticated fowl, including chickens, turkeys, ostriches, emus, rheas, cassowaries, waterfowl, and game birds, except doves and pigeons, which are bred for the primary purpose of producing eggs or meat.

Poults: Newly hatched turkeys.

Primary breeding flock: A flock composed of one or more generations maintained for the purpose of establishing, continuing, or improving parent lines.

Primary egg-type chicken breeding flocks: Foundation flocks composed of pedigree, great-grandparent, and grandparent stock developed for egg production and maintained for the principal purpose of producing multiplier breeding chicks used to produce table egg layers.

Primary meat-type chicken breeding flocks: Foundation flocks composed of pedigree, great-grandparent, and grandparent stock developed for meat production and maintained for the principal purpose of producing multiplier breeding chicks used to produce commercial broilers.

Primary spent fowl: Domesticated poultry that were in production of hatching eggs and have been removed from such production.

Primary turkey breeding flocks: Foundation flocks composed of pedigree, great-grandparent, and grandparent stock developed for meat production and maintained for the principal purpose of producing multiplier breeding poults used to produce commercial turkeys.

Regionalization: Recognition of geographical zones of a country that can be identified and characterized by their level of risk for specific diseases. These zones can cover entire countries or parts of countries. Adjacent zones of different countries having similar risk characteristics can be combined into international regions. The region must be clearly and effectively delineated by natural, artificial, or legal boundaries. The region must have a common control policy for the specific disease. There must be a uniform, effective system of epidemiological surveillance throughout the region.

Sanitize: To treat with a product which is registered and licensed by the Environmental Protection Agency (EPA) for the disease of concern in accordance with the specifications for use as shown on the label of each product.

Service: The Animal and Plant Health Inspection Service of the Department.

Shower: Process of cleansing, which includes first removing personal clothing and shoes in a designated (dirty) area, then washing with soap one's whole body and hair thoroughly under a stream of water, and then donning clean, company-provided, premises-specific clothing and footwear, in the biosecure zone (clean area). This process must follow the compartment company established policies.

Started chickens: Young chickens (chicks, pullets, cockerels, capons) which have been fed and watered and are less than 6 months of age.

State: Any State or U.S. territory, including the District of Columbia.

State Inspector: Any person employed or authorized under 9 CFR 145.11(b) to perform functions under that part.

Stock: The progeny of a specific breeding combination within a species of poultry. These breeding combinations may include pure strains, strain crosses, breed crosses, or combinations thereof.

Strain: Exclusive group of birds bred with a certain emphasis on specific traits.

Succeeding flock: A flock brought onto premises during the 12 months following removal of a flock.

Visitor: Individual who enters a premises who is not employed or contracted by the compartment company to work at those premises as his or her principal work location.

Acronyms

- **AI**-Avian Influenza
- **AIV**-Avian Influenza Virus
- **APHIS**-Animal and Plant Health Inspection Service
- **CFR**-Code of Federal Regulations
- **EDMP**-Egg Depot Management Protocols
- **EPA**-Environmental Protection Agency
- **FMP**-Farm Management Protocols
- **FMMP**-Feedmill Management Protocols
- **GMP** - General Management Protocols
- **HMP**-Hatchery Management Protocols
- **HRP**-High-Risk Period
- **HPAI**-Highly Pathogenic Avian Influenza
- **LPAI**-Low Pathogenicity Avian Influenza
- **LRP**-Low Risk Period
- ~~**NAI**-Notifiable Avian Influenza~~
- **NIES**-National Import Export Services
- **NPIP**-National Poultry Improvement Plan
- **NVSL**-National Veterinary Services Laboratory
- [ND](#)-[Newcastle Disease](#)
- [NDV](#)-[Newcastle Disease Virus](#)
- **OSA**-Official State Agency
- **PSD**-Program Standards Document
- **ISRCP**-Initial State Response and Containment Plan
- **OIE**-World Organization for Animal Health
- **USDA**-United States Department of Agriculture
- **VS**-Veterinary Services

Farm Design, Physical Requirements, and Management Procedures

For each requirement and procedure in this section, written biosecurity protocols must be on record for periods of low risk and high risk (when applicable). Training of all affected personnel implementing these protocols must be documented. Compliance with these protocols must be recorded.

Physical Requirements

- Farm must be separated from livestock (if present) by a livestock fence.
- Signs prohibiting unauthorized entry must be posted at the entrances to the controlled access zone and the biosecure zone to exclude unauthorized personnel and vehicles.
- Farm must be designed and built to deter and prevent entry of wildlife, pests, and companion animals.
- Each entrance to the biosecure zone should be locked or controlled at all times.
- A biosecure zone barrier must surround the biosecure zone of each farm to exclude unauthorized personnel and vehicles.
- Buildings within the biosecure zone must be constructed of materials that are durable and moistureproof and that can withstand routine cleaning and disinfection.
- Egg holding rooms must have barriers in place to prevent unauthorized entry.

Management Procedures

- Authorized personnel may enter the controlled access zone and biosecure zone after meeting company-established sanitation procedures.
- Authorized vehicles may enter the controlled access zone after meeting company established sanitation procedures. There may be dedicated vehicles that do not leave the controlled access zone.
- Authorized vehicles may enter the biosecure zone after meeting company-established procedures for cleaning and sanitizing the interior and exterior of the vehicle.
- Authorized personnel must follow company protocols and procedures and meet all biosecurity requirements for employment or contractual agreement before entry into the biosecure zone:
 - ✦ Company employees (and household members) and contract staff cannot own any birds.
 - ✦ Company employees and contract staff should avoid contact with birds outside the compartment and/or must comply with company policies related to downtime and quarantine.
 - ✦ Company employees and contract staff must receive annual, documented biosecurity training.
- All visitors must meet a minimum 24-hour downtime from contact with non- compartment birds (including a shower and change of clothing) and be authorized by following company-established procedures. All visitors must sign a declaration stating date of last bird contact.
- A whole-body shower and a change of clothing and footwear are required to enter the biosecure zone.
- All personnel and visitors must follow company-established policy regarding personal items and food.
- All personnel and visitors entering the biosecure zone must log in.
- Procedures must be in place to prevent entry from the egg room into the biosecure zone.

- Supplies and goods coming onto the farm should undergo company-established sanitation procedures. Tools and equipment must undergo company-established cleaning and disinfection procedures.
- Programs for vermin, wild birds, insects, and rodent control must be in place.
- Vegetation must be properly maintained according to company-established protocols.
- Surface water must not be used for any purpose. Treated well or municipal water must be used.
- Bedding materials must be obtained from a company-approved supplier. Suitable storage of bedding materials should prevent access from pests and wild birds.
- Any feed spills must be removed following company-established procedures.
- Daily mortality, biological waste, and cull eggs must be disposed of according to the company's biosecurity plan and in compliance with local environmental regulations.
- Record keeping and health program:
 - ✦ Daily production and mortality records must be kept according to company-established policies.
 - ✦ Unexplained increases in mortality and other clinical signs of disease must be investigated in compliance with the company veterinary health plan.
- All hatching eggs/chicks/poults and bird movement:
 - ✦ Hatching eggs should be sanitized with an EPA-approved disinfectant prior to delivery at a compartment facility.
 - ✦ All vehicles, equipment, and personnel involved in moving hatching eggs/chicks/poults within the compartment must comply with company-established sanitation and biosecurity procedures.
 - ✦ Hatching egg/chick/poult transport personnel must be trained and meet company-established biosecurity protocols. Drivers must wear company-provided clothing and footwear.
 - ✦ Records tracing the origin and production dates of all hatching eggs/chicks/poults must be kept.
- Bird movement within the compartment:
 - ✦ When birds are moved between premises within the compartment, a flock must test AI negative within 21 days prior to movement. Day-old chicks/poults must be derived from NPIP AI Clean Program source flocks.
 - ✦ [Birds must originate from flocks that were vaccinated for ND using licensed vaccines and compliant with a program to evaluate serological response to ND vaccination OR if unvaccinated, flocks have tested negative to ND.](#)
- Bird movement into the compartment:
 - ✦ Day old chicks/poults originating outside the compartment must be derived from a source flock that has tested negative for AI within 21 days of shipment. A minimum of 30 samples per source flock must be tested using an approved NPIP assay. The source flocks must participate in a national AI plan equivalent to the NPIP.
 - ✦ Pullets, cockerels, and adult birds originating outside the compartment must have tested negative for AI within 21 days of shipment. A minimum of 30 samples per flock must be tested by serology and 15 samples by antigen detection. Flocks must be inspected by an official veterinarian or designee within 30 days of movement.
 - ✦ [Birds must originate from flocks that were vaccinated for ND using licensed vaccines and compliant with a program to evaluate serological response to ND vaccination OR if unvaccinated, flocks have tested negative to ND.](#)
- Flock depletion and house sanitation:
 - ✦ Flocks to be depleted must test AI negative within 21 days prior to movement.

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- ◇ [Flocks to be depleted must be inspected by an accredited veterinarian within 21 days prior to flock depletion.](#)
- ◇ The removal of birds and litter must follow company-established biosecurity and sanitation procedures. Cleaning and disinfection of the houses following depletion must adhere to company-established procedures.
- ◇ Depletion of multi-age and/or multi-building premises requires appropriate company-established procedures to ensure biosecurity.
- Restocking of farm:
 - ◇ Introduction of new bedding material and bird restocking will only be allowed after trained company employees ensure that cleaning and disinfection have been performed to meet the company-established procedures.
- [Vaccination and serological monitoring programs for Newcastle disease:](#)
 - ◇ [Unvaccinated Flocks:](#)
 - [It is a primary breeding flock in which a minimum of 30 birds have been tested negative for ND using an approved test when more than 4 months of age. To retain this classification:](#)
 - [A sample of at least 30 birds must be tested negative at intervals of 90 days; OR](#)
 - [A sample of fewer than 30 birds may be tested, and found negative, at any one time if all pens are equally represented and a total of 30 birds is tested within each 90-day period; AND](#)
 - [During each 90-day period, all primary spent fowl, up to a maximum of 30, must be tested negative to ND within 21 days prior to movement to slaughter.](#)
 - ◇ [Vaccinated Flocks:](#)
 - [It is a primary breeding flock that has been vaccinated with licensed vaccines, as described in §113.329, manufactured with low-virulence live strains during early stages of development up to grow-out, and killed vaccines as final vaccination prior to onset of egg production; AND](#)
 - [The flock has been monitored for antibody response using approved serological tests as described in §145.14 and the results are compatible with immunological response against ND vaccination; AND](#)
 - [Testing must include:](#)
 - [A minimum of 30 birds when birds are more than 4 months of age and not longer than every 90 days thereafter.](#)

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Required Farm Design, Physical Requirements, and Management Protocols (FMP)

- FMP 1.** Site plan for each farm in the compartment which shows the physical characteristics of the component.
- FMP 2.** Farm specifications: Fencing, signage, and construction.
- FMP 3.** Farm biosecurity.
- FMP 4.** Entry of staff, visitors, and vehicles into the controlled access zone.
- FMP 5.** Entry of staff and visitors into the biosecure areas (shower, login).
- FMP 6.** Production and mortality monitoring records.
- FMP 7.** Entry of supplies, bedding, and equipment into biosecure areas.
- FMP 8.** Farm depletion, sanitation, and restocking.
- FMP 9.** Movement of birds and eggs into the compartment.

FMP 10. Movement of birds and eggs within the compartment.

FMP 11. Movement of birds and eggs out of the compartment.

FMP 12. Pest and wildlife management and control.

FMP 13. Mortality and biological waste disposal.

Feedmill Design, Physical Requirements, and Management Procedures

For each requirement and procedure in this section, written biosecurity protocols must be on record for periods of low risk and high risk (when applicable). Training of all affected personnel implementing these protocols must be documented. Compliance with these protocols must be recorded.

Physical Requirements

- Feedmill must be separated from livestock (if present) by a livestock fence.
- Signs prohibiting unauthorized entry must be posted at the entrance to the controlled access zone to exclude unauthorized personnel and vehicles.
- Feedmill must have a gate at the entrance of the controlled access zone.
- Feedmill must be designed and built to deter and prevent entry of wildlife, pests, and companion animals.
- Feedmill must be constructed of materials that are durable and moistureproof and that can withstand routine cleaning and disinfection.

Management Procedures

- Authorized personnel and vehicles may enter the controlled access zone after meeting company-established sanitation procedures.
- Authorized personnel must follow company protocols, procedures, and meet all biosecurity requirements for employment or contractual agreement before entry into the feedmill:
 - ✦ Company employees (and household members) and contract staff cannot own any birds.
 - ✦ Company employees and contract staff should avoid contact with birds outside the compartment and/or must comply with company policies related to downtime and quarantine.
 - ✦ Company employees and contract staff must receive annual, documented biosecurity training.
- All visitors must meet company-established procedures before entering the feedmill.
- All visitors must sign a declaration stating date of last bird contact.
- All personnel and visitors must follow company-established policy regarding personal items and food.
- All personnel and visitors entering the controlled access zone must log in.
- Programs for vermin, wild birds, insects, and rodent control must be in place.
- Vegetation must be properly maintained according to company-established protocols.
- Surface water must not be used for any purpose. Treated well or municipal water must be used.
- Finished feed must undergo a company-established treatment procedure prior to storage and distribution.
- Spills of any feed ingredient and/or finished feed must be removed following company-established procedures.
- All feedmills must have company-established protocols for separation of raw ingredients and finished feed.
- All feedmills must have company-established protocols for cleaning and disinfection.
- Feed delivery vehicles and personnel must comply with company-established biosecurity and sanitation policies for compartment and non-compartment premises deliveries.

- If a contract feedmill is used by the company, it must meet all physical requirements and management and manufacturing protocols listed above. A signed contract, which includes these details, must be available for inspection. Feed truck drivers and vehicles should be dedicated to the compartment. However, if feed is delivered to a non-compartment component, the vehicle and driver must undergo company-established cleaning and sanitation protocols before new delivery of feed into the compartment premises.

Required Feedmill Design, Physical Requirements, and Management Protocols (FMMP)

FMMP 1. Site plan for each feedmill in the compartment which shows the physical characteristics of the component.

FMMP 2. Feedmill specifications: Signage and construction.

FMMP 3. Feedmill biosecurity.

FMMP 4. Entry of staff and visitors into the controlled access zone (login).

FMMP 5. Manufacturing of feed.

FMMP 6. Separation and storage of raw ingredients and finished feed.

FMMP 7. Delivery of feed: Vehicles and drivers.

FMMP 8. Pest and wildlife management and control.

FMMP 9. Feed ingredient/finished feed spillage cleanup.

Hatchery Design, Physical Requirements, and Management Procedures

For each requirement and procedure in this section, written biosecurity protocols must be on record for periods of low risk and high risk (when applicable). Training of all affected personnel implementing these protocols must be documented. Compliance with these protocols must be recorded.

Physical Requirements

- Hatchery must be separated from livestock (if present) by a livestock fence.
- Signs prohibiting unauthorized entry must be posted at the entrances to the controlled access zone and the biosecure zone to exclude unauthorized personnel and vehicles.
- Hatchery must have a gate at the entrance of the controlled access zone. Hatchery office and egg/chick/poult loading docks may be considered part of the controlled access zone. The remainder of the hatchery is considered the biosecure zone.
- Hatchery must be designed and built to deter and prevent entry of wildlife, pests, and companion animals.
- A biosecure zone barrier must surround the biosecure zone of the hatchery to exclude unauthorized personnel and vehicles.
- Each entrance to the biosecure zone should be locked or controlled at all times.
- Hatchery must be constructed of materials that are durable and moistureproof and can withstand routine cleaning and disinfection.
- Receiving/shipment dock should be an enclosed area.
- Receiving/holding rooms must have barriers to prevent unauthorized entry into the biosecure zone.

Management Procedures

- Authorized personnel may enter the controlled access zone and biosecure zone after meeting company-established sanitation procedures.
- Authorized vehicles may enter the controlled access zone after meeting company-established sanitation procedures.
- Authorized vehicles can enter the biosecure zone only after meeting company-established cleaning and sanitizing procedures for the interior and exterior of the vehicle.
- Authorized personnel must follow company protocols and procedures and meet all biosecurity requirements for employment or contractual agreement before entry into the biosecure zone:
 - ✧ Company employees (and household members) and contract staff cannot own any birds.
 - ✧ Company employees and contract staff should avoid contact with birds outside the compartment and/or must comply with company policies related to downtime and quarantine.
 - ✧ Company employees and contract staff must receive annual, documented biosecurity training.
- All visitors must meet a minimum 24-hour downtime from contact with non- compartment birds (including a shower and change of clothing) and be authorized by following company-established procedures.
- All visitors must sign a declaration stating date of last bird contact.
- A whole-body shower and a change of clothing and footwear are required to enter the biosecure zone.

- All personnel and visitors must follow company-established policy regarding personal items and food.
- All personnel and visitors entering the biosecure zone must log in.
- Procedures must be in place to prevent entry from the egg receiving area into the biosecure zone.
- Supplies and goods coming into the hatchery should undergo company-established sanitation procedures. Tools and equipment must undergo company-established cleaning and disinfection procedures.
- Programs for vermin, wild birds, insects, and rodent control must be in place.
- Vegetation must be properly maintained according to company-established protocols.
- Surface water must not be used for any purpose. Treated well or municipal water must be used.
- Biological waste, hatchery residue, and cull eggs must be disposed of according to the company's biosecurity plan and in compliance with local environmental regulations.
- All hatcheries must have company-established protocols for cleaning and disinfection.
- Hatchery egg and chick/poult identification and traceability records:
 - ✦ Records tracing the origin and production dates of all hatching eggs and chicks/poults in the hatchery must be kept.
- All hatching egg/chick/poult movement:
 - ✦ Hatching eggs should be sanitized with an EPA-approved disinfectant prior to delivery at a compartment facility.
 - ✦ All vehicles, equipment, and personnel involved in moving hatching eggs/chicks/poults within the compartment must comply with company-established sanitation and biosecurity procedures.
 - ✦ Hatching egg/chick/poult transport personnel must be trained and meet company-established biosecurity protocols. Drivers must wear company-provided clothing and footwear.
 - ✦ Records tracing the origin and production dates of all hatching eggs/chicks/poults must be kept.
- Hatching egg movement into the compartment:
 - ✦ Hatching eggs originating outside the compartment must be derived from a source flock that has tested negative for AI within 21 days of shipment. A minimum of 30 samples per source flock must be tested using an approved NPIP assay. The source flocks must participate in a national AI plan equivalent to the NPIP.
 - ✦ Hatching eggs must originate from flocks that were vaccinated for ND using licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If source flocks were not vaccinated for ND, they must test negative to ND.
- Hatching egg movement within the compartment:
 - ✦ When hatching eggs are moved between premises within the compartment, they must be derived from NPIP AI Clean Program source flocks.
 - ✦ Hatching eggs must originate from flocks that were vaccinated for ND using licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If source flocks were not vaccinated for ND, they must test negative to ND.
- Day-old chick/poult movement within the compartment:
 - ✦ Day-old chicks/poults must be derived from NPIP AI Clean Program compartment source flocks or otherwise qualified flocks that have equivalent requirements to be brought in from outside the compartment.

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- ◇ Reusable chick/poult boxes used to deliver day-old chicks/poults must be cleaned and disinfected on return to the hatchery.
- ◇ [Birds must originate from flocks that were vaccinated for ND using licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If source flocks were not vaccinated for ND, they must test negative to ND.](#)
- Hatching egg/chick/poult movement out of the compartment:
 - ◇ Any reusable equipment which returns to the hatchery must be cleaned and disinfected.

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Required Hatchery Design, Physical Requirements, and Management Protocols (HMP)

HMP 1. Site plan for each hatchery in the compartment which shows the physical characteristics of the component.

HMP 2. Hatchery specifications: Fencing, signage, and construction.

HMP 3. Hatchery biosecurity.

HMP 4. Entry of staff and visitors into the biosecure areas (shower and login).

HMP 5. Entry of supplies and equipment into biosecure areas.

HMP 6. Entry of staff, visitors, and vehicles into the controlled access zone.

HMP 7. Chick/poult delivery: Washing and disinfection (vehicle, personnel, boxes).

HMP 8. Hatchery egg and chick/poult identification and traceability records.

HMP 9. Hatching egg movement into the compartment.

HMP 10. Chick/poult and hatching egg movement within the compartment.

HMP 11. Chick/poult and hatching egg movement out of the compartment.

HMP 12. Hatchery sanitation.

HMP 13. Hatching egg sanitation.

HMP 14. Pest and wildlife management and control.

HMP 15. Chick/poult delivery and hatching egg pickup.

Egg Depot Design, Physical Requirements, and Management Procedures

For each requirement and procedure in this section, written biosecurity protocols must be on record for periods of low risk and high risk (when applicable). Training of all affected personnel implementing these protocols must be documented. Compliance with these protocols must be recorded.

Physical Requirements

- Egg depot must be separated from livestock (if present) by a livestock fence.
- Signs prohibiting unauthorized entry must be posted at the entrance to the controlled access zone to exclude unauthorized personnel and vehicles.
- Egg depot must have a gate at the entrance of the controlled access zone.
- Egg depot must be designed and built to deter and prevent entry of wildlife, pests, and companion animals.
- Egg depot must be constructed of materials that are durable and moistureproof and that can withstand routine cleaning and disinfection.
- Egg receiving/shipment dock should be an enclosed area.
- Egg receiving/holding rooms must have barriers in place to prevent unauthorized entry.

Management Procedures

- Authorized personnel and vehicles may enter the controlled access zone after meeting company-established sanitation procedures.
- Authorized personnel must follow company protocols and procedures and meet all biosecurity requirements for employment or contractual agreement before entry into the egg depot:
 - ✦ Company employees (and household members) and contract staff cannot own any birds.
 - ✦ Company employees and contract staff should avoid contact with birds outside the compartment and/or must comply with company policies related to downtime and quarantine.
 - ✦ Company employees and contract staff must receive annual, documented biosecurity training.
- All visitors must meet company-established procedures before entering the egg depot. All visitors must sign a declaration stating date of last bird contact.
- All personnel and visitors must follow company-established policy regarding personal items and food.
- All personnel and visitors entering the controlled access zone must log in.
- Procedures and barriers must be in place to prevent entry from the egg receiving area into the egg depot.
- Programs for vermin, wild birds, insects, and rodent control must be in place.
- Vegetation must be properly maintained according to company-established protocols.
- Surface water must not be used for any purpose. Treated well or municipal water must be used.
- Biological waste, egg depot residue, and cull eggs must be disposed of according to the company's biosecurity plan and in compliance with local environmental regulations.
- All egg depots must have company-established protocols for cleaning and disinfection.
- Egg depot identification and traceability records:
 - Records tracing the origin of all hatching eggs and production dates in the egg depot must be kept.

- All hatching egg movement:
 - ✦ Hatching eggs must be sanitized with an EPA-approved disinfectant prior to delivery at a compartment facility.
 - ✦ All vehicles, equipment, and personnel involved in moving hatching eggs/chicks/poults within the compartment must comply with company-established sanitation and biosecurity procedures.
 - ✦ Hatching egg transport personnel must be trained and meet company-established biosecurity protocols. Drivers must wear company-provided clothing and footwear.
 - ✦ Records tracing the origin and production dates of all hatching eggs must be kept.
- Hatching egg movement into the compartment:
 - ✦ Hatching eggs originating outside the compartment must be derived from a source flock that has tested negative for AI within 21 days of shipment. A minimum of 30 samples per source flock must be tested using an approved NPIP assay. The source flocks must participate in a national AI plan equivalent to the NPIP.
 - ✦ [Hatching eggs must originate from flocks that were vaccinated for ND using licensed ND vaccines and compliant with a program to evaluate serological response to ND vaccination. If source flocks were not vaccinated for ND, they must test negative to ND.](#)
- Hatching egg movement within the compartment:
 - ✦ Egg receiving/shipment dock must undergo routine company-established cleaning and disinfection procedures.
 - ✦ When hatching eggs are moved between premises within the compartment they must be derived from NPIP AI Clean Program source flocks.
 - ✦ [Hatching eggs must originate from flocks that were vaccinated for ND using licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If source flocks were not vaccinated for ND, they must test negative to ND.](#)
- Hatching egg movement out of the compartment:
 - ✦ Any reusable equipment returning to the egg depot must be cleaned and disinfected.

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Required Egg Depot Design, Physical Requirements, and Management Protocols (EDMP)

- EDMP 1. Site plan for each egg depot in the compartment which shows the physical characteristics of the component.
- EDMP 2. Egg depot specifications: Fencing, signage, and construction.
- EDMP 3. Egg depot biosecurity.
- EDMP 4. Entry of staff and visitors into the egg depot (login, change of clothing).
- EDMP 5. Chick/poult delivery: Washing and disinfection (vehicle, personnel, boxes)
- EDMP 6. Hatchery egg identification and traceability records.
- EDMP 7. Egg movement into the compartment.
- EDMP 8. Egg movement within the compartment.
- EDMP 9. Egg movement outside the compartment.
- EDMP 10. Egg depot sanitation.
- EDMP 11. Hatching egg sanitation.
- EDMP 12. Pest and wildlife management and control.
- EDMP 13. Egg pickup and delivery.

Required High-Risk Period Biosecurity and Management Protocols (HRP)

- HRP 1.** Non-essential visitation and delivery policy.
- HRP 2.** Regional poultry industry meeting attendance.
- HRP 3.** Enhanced communication system for company employees, contract growers, and suppliers.
- HRP 4.** 48-hour testing prior to movement/depletion of poultry.
- HRP 5.** Alternate transport and service vehicle driving routes.
- HRP 6.** Reduced vehicle movement and non-essential parking policy.
- HRP 7.** Enhanced vehicle cleaning and disinfection.
- HRP 8.** Use, cleaning, and disinfection of tools and equipment.
- HRP 9.** Increased downtime after contact with non-compartment birds.
- HRP 10.** Bird hunting policy for employees and contract growers.
- HRP 11.** Reporting of increased mortality and egg production drops by veterinarians and live production.
- HRP 12.** 48-hour testing prior to moving litter/manure from premises with birds present.
- HRP 13.** Controlled access zone entry.



Appendix A: NPIP Avian Influenza and Newcastle Disease Compartmentalization Application Forms



Thank you for your application for NPIP Avian Influenza and Newcastle Disease compartmentalization. Below are the next steps to expect after completion of your Compartmentalization Application Forms. “You” refers to the person listed in the contact section of the application form.

Term Definitions:

Registered/Registration: A compartment or component that has had initial applications approved.

Certified/Certification: A compartment or component that has been through the audit process and successfully passed.

FIRST TIME APPLICANTS:

If you have completed and submitted **Application Form A (Compartment Registration)**, the NPIP Office will review your application form. If the NPIP determines that your application is satisfactory, you will be approved for compartment registration. However, you will not have any components within the compartment, so you will be asked to complete **Application Form B**, which shows in detail each component you intend to add to the registered compartment for which you are seeking certification.

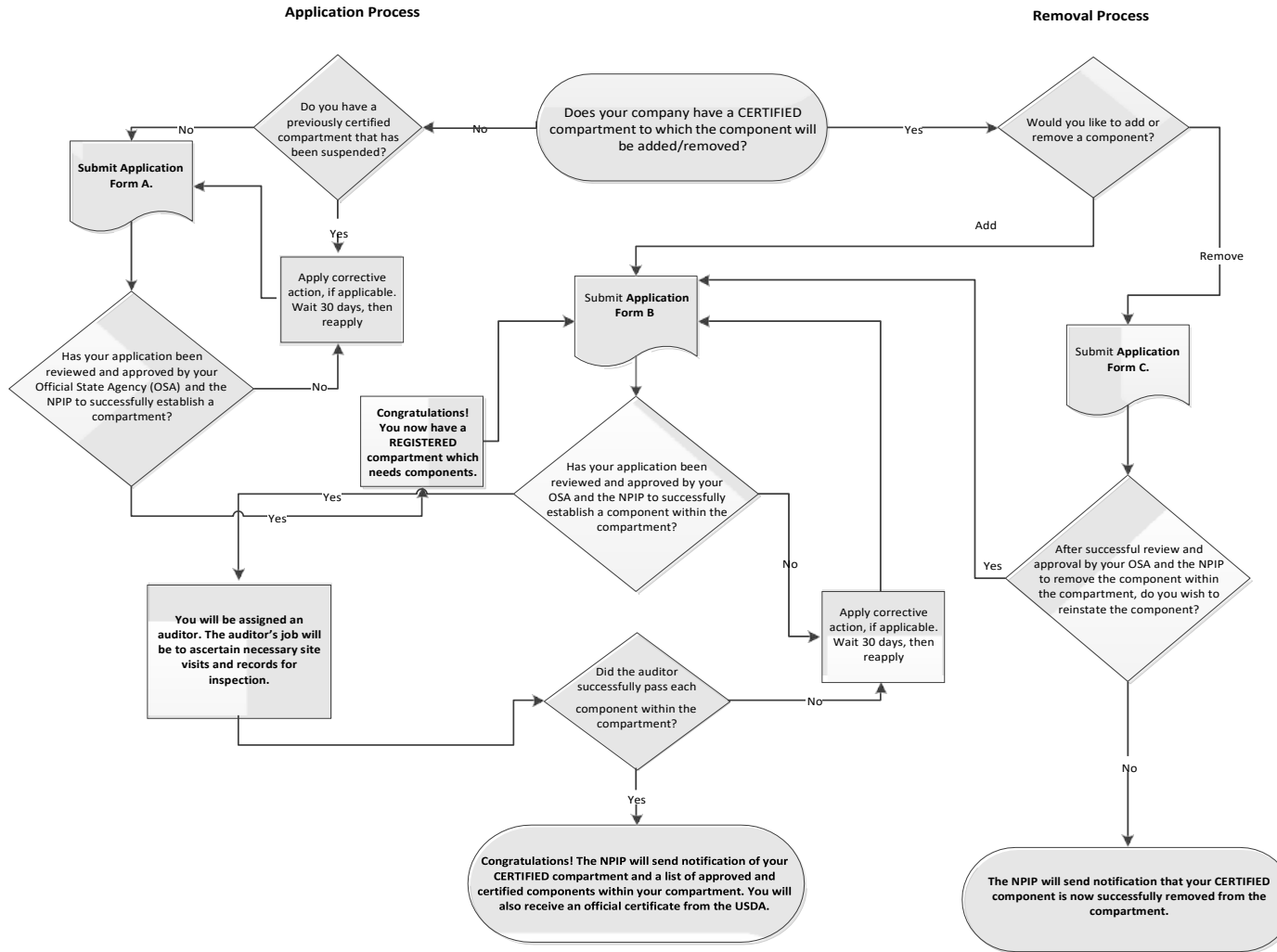
Once you have completed and submitted **Application Form B (Component Registration)**, the NPIP Office will review your application form. If the NPIP determines that your application is satisfactory, it will assign an *Auditor*. The auditor will contact you and will request a meeting to set up details for reviewing in more depth each piece of documented information as listed in the *Prerequisites* section of **Application Form A** and **Application Form B**. This initial audit contact may include, but will not necessarily be limited to, a phone interview, document sharing in person or via a secure connection, and a site visit. **Note that for initial compartment registration, all components require a site inspection.* Only after an approved Application Form A and an approved Application Form B, which REGISTER both the compartment and the component, as well as a passing score from the auditor, will both the registered compartment and the registered components within the compartment become CERTIFIED.

CERTIFIED COMPARTMENT USERS:

If your compartment with the USDA has been successfully certified and you wish to add a new component to your compartment, reinstate a previously removed component to the compartment, or recertify your previously suspended component of the compartment, complete **Application Form B**. If you have a certified compartment with the USDA and wish to remove a certified component from your compartment, complete **Application Form C (Component Removal)**.

The same procedure listed above for first-time users will apply for the addition of a new component, after successful completion of **Application Form B**. If, after you have completed and submitted **Application Form C**, the NPIP Office determines that your application is satisfactory, you will be sent notification of removal of the desired component. If you wish to reinstate the removed component, you will need to complete **Application Form B**.

Appendix B: Application and Removal Processes





Appendix C: Application Form A: U.S. Avian Influenza and Newcastle Disease Clean Compartment Registration



Instructions: Step 1: Applicants, please complete Sections A and B and certify application with signature on pg. 3. Step 2: Send Form A to the OSA which completes Section C and signs. Step 3: OSA returns form to NPIP. Note: If you are using Form A to comply with recertification requirements and none of the information in Sections A or B has changed since initially applying, please complete only Section A and proceed to Step 2. Disclaimer: This form may be simultaneously submitted with Application Form B: Component Registration. However, Application Form B will not be reviewed until Application Form A has been reviewed and approved.

A: Background Information. *To be completed by company seeking certification.*

Name of Company	
Company Mailing Address	
Name of Contact	
Telephone Number	
Alternate Telephone Number	
Fax Number	
Email Address	
NPIP Classification	U.S. AI Clean <input type="checkbox"/> U.S. H5/H7 AI Clean <input type="checkbox"/> U.S. ND Clean <input type="checkbox"/>
Breed/Type of Poultry	
NPIP Classification Seeking	
Compartment Mailing Address	
Compartment Location (List States Involved)	
Name of Compartment	
Anticipated Type of Components (F, M, H, and E) to add within Compartment	Farm <input type="checkbox"/> Feedmill <input type="checkbox"/> Hatchery <input type="checkbox"/> Egg Depot <input type="checkbox"/>

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B: Prerequisites. *To be completed by company seeking certification.*

*To be eligible for certification as a compartment, all of the protocols listed below and supporting documents must be available and ready for presentation to the compartmentalization auditors. Refer to the **Compartmentalization for Protection Against Avian Influenza and Newcastle Disease in Primary Poultry Breeding Companies in the United States of America; Specifications for Management Procedures, Physical Requirements and Protocols** for more details.*

Please place a check mark by the answer that applies.

General Management Protocols		
For each component, have you met all of the required specifications for management procedures and physical requirements; do you have the necessary protocols and documentation as specified in the Compartmentalization for Protection Against Avian Influenza and Newcastle Disease in Primary Poultry Breeding Companies in the U.S.A. and further, do you have documentation outlining the following items?	Yes	No
Biosecurity training for employees, contract staff, and visitors		
Biosecurity compliance agreement for employees, contract staff, and visitors		
Biosecurity risk assessment for each component of the compartment		
Cleaning, sanitation, and control of vehicles prior to entering biosecure areas		
General physical traits of each compartment component (Farms, Feedmills, Hatcheries, Egg Depots and Offices), including physical address with GPS location		
Detailed diagrammatic description for movement of people, vehicles, equipment, birds, and eggs between all components inside and outside the compartment		
Company Emergency Response Plan		
Veterinary Health Plan		
ND Vaccination Program		
ND Testing Program for ND, vaccinated or unvaccinated flocks		

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C. Questionnaire. To be completed by each Official State Agency

Please place a check mark by the answer that applies.

	Yes	No
Is the company seeking certification in the U.S. H5/H7 Avian Influenza and Newcastle Disease Clean Compartment program a participant in good standing with the NPIP: U.S. H5/H7 Avian Influenza Clean and Newcastle Disease Clean Programs for Turkey Breeding Flocks?		
Is the company seeking certification in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program a participant in good standing with the NPIP: U.S. Avian Influenza Clean and Newcastle Disease Clean Programs for Primary Egg-Type Chicken Breeding Flocks?		
Is the company seeking certification in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program a participant in good standing with the NPIP: U.S. Avian Influenza Clean and Newcastle Disease Clean Programs for Primary Meat-Type Chicken Breeding Flocks?		
Within the company, are all operations seeking certification as components within the registered compartment in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program (for egg- type chicken breeding flocks and meat-type chicken breeding flocks) or the U.S. H5/H7 Avian Influenza and Newcastle Disease Clean Compartment (for turkey breeding flocks) located in a State which has an APHIS-approved Initial State Response and Containment Plan?		
Does the company seeking certification in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program perform routine surveillance of all flocks within the compartment in an NPIP-authorized laboratory which is certified to test for AI and ND ?		

CERTIFICATION OF OFFICIAL STATE AGENCY or AGENCIES

I DO HEREBY CERTIFY THAT ALL STATEMENTS MADE BY ME IN THIS APPLICATION ARE TRUE AND CORRECT TO THE BEST OF MY KNOWLEDGE, INFORMATION AND BELIEF; FURTHER, I UNDERSTAND THAT IN THE EVENT I HAVE KNOWINGLY AND WILLFULLY MADE ANY FALSE STATEMENTS, I WILL BE LIABLE FOR PUNISHMENT IN ACCORDANCE WITH ALL APPLICABLE LAWS AND STATUTES.

State: _____ State: _____
Signature: _____ Signature: _____
Date: _____ Date: _____

State: _____ State: _____
Signature: _____ Signature: _____
Date: _____ Date: _____

CERTIFICATION OF APPLICANT

I DO HEREBY CERTIFY THAT ALL STATEMENTS MADE BY ME IN THIS APPLICATION ARE TRUE AND CORRECT TO THE BEST OF MY KNOWLEDGE, INFORMATION, AND BELIEF AND I HAVE OBTAINED ALL NECESSARY OFFICIAL STATE AGENCIES' CERTIFICATION IN C ABOVE. FURTHER, I UNDERSTAND THAT IN THE EVENT I HAVE KNOWINGLY AND WILLFULLY MADE ANY FALSE STATEMENTS, I WILL BE LIABLE FOR PUNISHMENT IN ACCORDANCE WITH ALL APPLICABLE LAWS AND STATUTES.

Signature: _____
Date: _____

Application

A complete application must be sent to:

The National Poultry Improvement Plan
1506 Klondike Road,
Suite 101
USDA-APHIS-VS
Conyers, GA 30094

~~Denise.L.Brinson@aphis.usda.gov~~ ~~Denise.L.Heard@aphis.usda.gov~~

with cc to

~~Elena.L.Behnke@aphis.usda.gov~~ ~~Christina.Lindsev@aphis.usda.gov~~

For Department Use Only

Date

Received: _____ Reviewer: _____

Check Here if Registration Approval Granted:

Check Here if Registration Approval Denied:

Signature: _____

If Denied, List Reasons:

Please note that registration approval does not mean that the component is certified. Only after a successful registration using this form, a successful registration of components using Application Form B, and a successful audit can the compartment become fully certified.



Appendix D: Application Form B: U.S. Avian Influenza and Newcastle Disease Clean Compartment Component Registration



Instructions: Step 1: Applicants, please complete Sections A-E and certify application with signature on pg. 6. Step 2: Send the form to the OSA which completes Section F and signs. Step 3: OSA returns form to NPIP. Note: If you are using Form B to comply with recertification requirements and none of the information in Sections A-E has changed since initially applying, please complete only Section A and proceed to Step 2. Disclaimer: For initial Compartment and Component registration, this form may be simultaneously submitted with Application Form A: Compartment Registration for initial registration. However, Application Form B will not be reviewed until Application Form A has been reviewed and approved.

A: Background Information. To be completed by company seeking certification.

To be considered for approval as a new component within a certified compartment, the following must be completed.

Name of Company	
Company Mailing Address	
Name of Contact	
Telephone Number	
Alternate Telephone Number	
Fax Number	
Email Address	
NPIP Classification	U.S. AI Clean <input type="checkbox"/> U.S. H5/H7 AI Clean <input type="checkbox"/> U.S. ND Clean <input type="checkbox"/>
Breed/Type of Poultry	
NPIP Classification Seeking	
Compartment Mailing Address	
Compartment Location (List States Involved)	
Name of Compartment	
Anticipated Type of Components (F, M, H, and E) to add within Compartment	Farm <input type="checkbox"/> Feedmill <input type="checkbox"/> Hatchery <input type="checkbox"/> Egg Depot <input type="checkbox"/>
Total Number of Components Seeking Certification (sum of total numbers listed in sections B-E below)	

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Questionnaire. *To be completed by company seeking certification.*

Please place a check mark by the answer that applies.

	YES	NO
U.S. Avian Influenza and Newcastle Disease Compartment Registration Form (Application Form A) submitted. This form contains the components to be added within the new compartment.		
New facility within previously certified compartment.		
Requalification of components within certified compartment due.		
Components previously removed from certified compartment and now seeking reinstatement within certified compartment.		

B. Prerequisites for Farms (F). *To be completed by company seeking certification.*

To be considered for approval as a component in a certified compartment, you must first provide the following information.

Total number of farm premises seeking approval (Please list number). _____

List farm names (and associated NPIP numbers) seeking approval in box provided below. Separate farms by use of a semicolon. Example: ChickaD, 13-3223; Hollow Oak 1, 12-1392; Hollow Oak 2, 12-1293. This example includes three separate farms and three separate NPIP numbers [or EMRS Premises Identification Numbers](#).

Note: Supporting documents for Statements 1 and 2 below must be submitted with this application for [each farm](#). Please refer to the [Compartmentalization for Protection Against Avian Influenza and Newcastle Disease in Primary Poultry Breeding Companies in the United States of America](#); [Specifications for Management Procedures, Physical Requirements, and Protocols for verification of statement 3](#).

Farm Design, Physical Requirements, and Management Protocols	YES	NO
Statement 1: FMP 1: <i>Site plan for each farm in the component which shows characteristics of the component.</i> I hereby certify that I have attached to this application a site plan for each farm seeking to be added as a component within the compartment.		
Statement 2: FMP 2: <i>Farm specifications, including fencing, signage, and construction. (Note that farm specifications include the physical address of each farm along with GPS coordinates.)</i>		

I hereby certify that I have attached to this application the applicable farm specifications for each farm seeking to be added as a component within the compartment.		
Statement 3: FMP3-FMP13: <i>Written documentation must be shown to the assigned auditor on request.</i> I hereby certify that written documentation for each of the Farm Management Protocols 3-13 is on file as accurate and complete to my knowledge and will be provided to the assigned auditor on request.		

C. Prerequisites for Feedmills (M). *To be completed by the company seeking certification.*

To be considered for approval as a component in a certified compartment, you must first provide the following information.

Total number of feedmill premises seeking approval (Please list number). _____

List feedmill names seeking approval in box provided below. Separate feedmills by use of a semicolon. Example: Feedmille 1; Jones & Parks; Willow Mill. This example includes three separate feedmills.

Note: Supporting documents for Statements 1 and 2 below must be submitted with this application for each feedmill. Please refer to the [Compartmentalization for Protection Against Avian Influenza and Newcastle Disease in Primary Poultry Breeding Companies in the United States of America](#); [Specifications for Management Procedures, Physical Requirements, and Protocols](#) for verification of statement 3.

Feedmill Design, Physical Requirements, and Management Protocols	YES	NO
Statement 1: FMMP 1: <i>Site plan for each feedmill in the component which shows characteristics of the component.</i> I hereby certify that I have attached to this application a site plan for each feedmill seeking to be added as a component within the compartment.		
Statement 2: FMMP 2: <i>Feedmill specifications, including signage and construction. (Note that feedmill specifications include the physical address of each feedmill along with GPS coordinates.)</i> I hereby certify that I have attached to this application the applicable feedmill specifications for each feedmill seeking to be added as a component within the compartment.		
Statement 3: FMMP3-FMMP9: <i>Written documentation must be shown to the assigned auditor on request.</i>		

I hereby certify that written documentation for each of the Feedmill Management Protocols 3-9 is on file as accurate and complete to my knowledge and will be provided to the assigned auditor on request.		
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D. Prerequisites for Hatcheries (H). *To be completed by company seeking certification.*

To be considered for approval as a component in a certified compartment, you must first provide the following information.

Total number of hatchery premises seeking approval (Please list number). _____

List hatchery names (and associated NPIP numbers) seeking approval in box provided below. Separate hatcheries by use of a semicolon. Example: Chickadee, Inc. -15-1425; Grandparent Line-65-1293. This example includes two separate hatcheries with two separate NPIP numbers.

Note: Supporting documents for Statements 1 and 2 below must be submitted with this application for each hatchery. Please refer to the [Compartmentalization for Protection Against Avian Influenza and Newcastle Disease in Primary Poultry Breeding Companies in the United States of America](#); Specifications for Management Procedures, Physical Requirements, and Protocols for verification of statement 3.

Hatchery Design, Physical Requirements, and Management Protocols	YES	NO
Statement 1: HMP 1: <i>Site plan for each hatchery in the component which shows characteristics of the component.</i> I hereby certify that I have attached to this application a site plan for each hatchery seeking to be added as a component within the compartment.		
Statement 2: HMP 2: <i>Hatchery specifications, including fencing, signage, and construction. (Note that hatchery specifications include the physical address of each hatchery along with GPS coordinates.)</i> I hereby certify that I have attached to this application the applicable hatchery specifications for each hatchery seeking to be added as a component within the compartment.		
Statement 3: HMP3-HMP15: <i>Written documentation must be shown to the assigned auditor on request.</i> I hereby certify that written documentation for each of the Hatchery Management Protocols 3-15 is on file as accurate and complete to my knowledge and will be provided to the assigned auditor on request.		

E. Prerequisites for Egg Depots (E). *To be completed by company seeking certification.*

To be considered for approval as a component in a certified compartment, you must first provide the following information.

Total number of egg depot premises seeking approval (Please list number). _____

List egg depot names seeking approval in box provided below. Separate egg depots by use of a semicolon. Example: Clayton 1, 2, and 3; Heart Storage. This example includes two separate egg depots. [Alternatively, Egg Depot location may be identified with NPIP number +/- EMRS premises identification number.](#)

Note: Supporting documents for Statements 1 and 2 below must be submitted with this application for each egg depot. Please refer to the **Compartmentalization for Protection Against Avian Influenza and Newcastle Disease in Primary Poultry Breeding Companies in the United States of America**; **Specifications for Management Procedures, Physical Requirements, and Protocols** for verification of statement 3.

Egg Depot Design, Physical Requirements, and Management Protocols	YES	NO
Statement 1: EDMP 1: Site plan for each hatchery in the component which shows characteristics of the component. I hereby certify that I have attached to this application a site plan for each egg depot seeking to be added as a component within the compartment.		
Statement 2: EDMP 2: Hatchery specifications, including fencing, signage, and construction. (Note that egg depot specifications include the physical address of each egg depot along with GPS coordinates.) I hereby certify that I have attached to this application the applicable egg depot specifications for each hatchery seeking to be added as a component within the compartment.		
Statement 3: EDMP3-EDMP12: Written documentation must be shown to the assigned auditor on request. I hereby certify that written documentation for each of the Egg Depot Management Protocols 3-12 is on file as accurate and complete to my knowledge and will be provided to the assigned auditor on request.		

F. Verification. To be completed by each Official State Agency.

Please place a check mark by the answer that applies.

	YES	NO
Is the company seeking certification in the U.S. H5/H7 Avian Influenza and Newcastle Disease Clean Compartment program a participant in good standing with the NPIP U.S. H5/H7Avian Influenza Clean and Newcastle Disease Clean Programs for Turkey Breeding Flocks?		

Is the company seeking certification in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program a participant in good standing with the NPIP U.S. Avian Influenza Clean and Newcastle Disease Clean Programs for Primary Egg-Type Chicken Breeding Flocks?		
Is the company seeking certification in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program a participant in good standing with the NPIP U.S. Avian Influenza Clean and Newcastle Disease Clean Programs for Primary Meat-Type Chicken Breeding Flocks?		
Within the company, are all operations seeking certification as components within the registered compartment participating in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program (for egg- type chicken breeding flocks and meat-type chicken breeding flocks) or the U.S. H5/H7 Avian Influenza and Newcastle Disease Clean Compartment program (for turkey breeding flocks) located in a State which has an APHIS-approved Initial State Response and Containment Plan?		

CERTIFICATION OF OFFICIAL STATE AGENCY or AGENCIES

I DO HEREBY CERTIFY THAT ALL STATEMENTS MADE BY ME IN THIS APPLICATION ARE TRUE AND CORRECT TO THE BEST OF MY KNOWLEDGE, INFORMATION AND BELIEF. FURTHER, I UNDERSTAND THAT IN THE EVENT I HAVE KNOWINGLY AND WILLFULLY MADE ANY FALSE STATEMENTS, I WILL BE LIABLE FOR PUNISHMENT IN ACCORDANCE WITH ALL APPLICABLE LAWS AND STATUTES.

State: _____ State: _____
 Signature: _____ Signature: _____
 Date: _____ Date: _____

State: _____ State: _____
 Signature: _____ Signature: _____
 Date: _____ Date: _____

CERTIFICATION OF APPLICANT

I DO HEREBY CERTIFY THAT ALL STATEMENTS MADE BY ME IN THIS APPLICATION ARE TRUE AND CORRECT TO THE BEST OF MY KNOWLEDGE, INFORMATION, AND BELIEF AND I HAVE OBTAINED ALL NECESSARY OFFICIAL STATE AGENCIES' CERTIFICATION IN C ABOVE. FURTHER, I UNDERSTAND THAT IN THE EVENT I HAVE KNOWINGLY AND WILLFULLY MADE ANY FALSE STATEMENTS, I WILL BE LIABLE FOR PUNISHMENT IN ACCORDANCE WITH ALL APPLICABLE LAWS AND STATUTES.

Signature: _____
 Date: _____

Application

A complete application must be sent to:

The National Poultry Improvement Plan
1506 Klondike Road,
Suite 101
USDA-APHIS-VS
Conyers, GA 30094

Denise.L.Brinson@aphis.usda.gov Denise.L.Heard@aphis.usda.gov
with cc to
Elena.L.Behnke@aphis.usda.gov Christina.Lindsey@aphis.usda.gov

For Department Use Only

Date _____
Received: _____ Reviewer: _____

Check Here if Registration Approval Granted:

Check Here if Registration Approval Denied:

Signature: _____

For Components Denied, if Any, List Reasons:

Please note that registration approval for components does not mean the components are certified. Only after an auditor's review and successful passing can a component become certified.



Appendix E: Application Form C: U.S. Avian Influenza and Newcastle Disease Clean Compartment Component Removal



Instructions: Applicants please complete Sections A and B and certify with signature. Then send the form to the OSA which completes Section C and signs. OSA returns form to NPIP.

A: BACKGROUND INFORMATION. *To be completed by company seeking removal of a component within a certified compartment. Please note that once a component has been successfully removed, it will no longer function as part of the compartment. Adding the component back to the compartment will require recertification using Application Form B.*

Name of Company	
Company Mailing Address	
Name of Contact	
Telephone Number	
Alternate Telephone Number	
Fax Number	
Email Address	
NPIP Classification	U.S. AI Clean <input type="checkbox"/> U.S. H5/H7 AI Clean <input type="checkbox"/> U.S. ND Clean <input type="checkbox"/>
Breed/Type of Poultry	
NPIP Classification Seeking	
Compartment Mailing Address	
Compartment Location (List States Involved)	
Name of Compartment	
Type of Components (F, M, H, and E) to add within Compartment	Farm <input type="checkbox"/> Feedmill <input type="checkbox"/> Hatchery <input type="checkbox"/> Egg Depot <input type="checkbox"/>

Deleted: v

B. Reason for Removal. *To be completed by company seeking component removal. To be eligible for removal as a compartment, a justification for removal and a detailed description of how the component removal will affect the rest of the compartment must be provided. Please use the box below. (Note: If component removal will not affect the compartment, please check here .)*

C. Verification. To be completed by each Official State Agency. Please place a check mark by the answer that applies.

	YES	NO
Is the component of the compartment part of a company that is a participant in the U.S. H5/H7 Avian Influenza and Newcastle Disease Clean Compartment program and in good standing with the NPIP: U.S. H5/H7 Avian Influenza Clean and Newcastle Disease Clean Programs for Turkey Breeding Flocks?		
Is the component of the compartment part of a company that is a participant in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program and in good standing with the NPIP: U.S. Avian Influenza Clean and Newcastle Disease Clean Programs for Primary Egg-Type Chicken Breeding Flocks?		
Is the component of the compartment part of a company that is a participant in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program and in good standing with the NPIP: U.S. Avian Influenza Clean and Newcastle Disease Clean Programs for Primary Meat-Type Chicken Breeding Flocks?		
Within the company, are all other operations certified as components within the registered compartment part of the U.S. Avian Influenza and Newcastle Disease Clean Compartment program (for egg-type chicken breeding flocks and meat-type chicken breeding flocks) or the U.S. H5/H7 Avian Influenza and Newcastle Disease Clean Compartment (for turkey breeding flocks) and located in a State which has an APHIS-approved Initial State Response and Containment Plan?		

CERTIFICATION OF OFFICIAL STATE AGENCY or AGENCIES

I DO HEREBY CERTIFY THAT ALL STATEMENTS MADE BY ME IN THIS APPLICATION ARE TRUE AND CORRECT TO THE BEST OF MY KNOWLEDGE, INFORMATION AND BELIEF. FURTHER, I UNDERSTAND THAT IN THE EVENT I HAVE KNOWINGLY AND WILLFULLY MADE ANY FALSE STATEMENTS, I WILL BE LIABLE FOR PUNISHMENT IN ACCORDANCE WITH ALL APPLICABLE LAWS AND STATUTES.

State: _____ State: _____
 Signature: _____ Signature: _____
 Date: _____ Date: _____

State: _____ State: _____
 Signature: _____ Signature: _____
 Date: _____ Date: _____

CERTIFICATION OF APPLICANT

I DO HEREBY CERTIFY THAT ALL STATEMENTS MADE BY ME IN THIS APPLICATION ARE TRUE AND CORRECT TO THE BEST OF MY KNOWLEDGE, INFORMATION, AND BELIEF, AND I HAVE OBTAINED ALL NECESSARY OFFICIAL STATE AGENCIES' CERTIFICATION IN C ABOVE. FURTHER, I UNDERSTAND THAT IN THE EVENT I HAVE KNOWINGLY AND WILLFULLY MADE ANY FALSE STATEMENTS, I WILL BE LIABLE FOR PUNISHMENT IN ACCORDANCE WITH ALL APPLICABLE LAWS AND STATUTES. FURTHER, I CERTIFY THAT BY COMPLETION OF THIS FORM, THIS COMPONENT OF THE COMPARTMENT WILL HAVE TO GO THROUGH THE RE- APPLICATION PROCESS TO BE FORMALLY RECOGNIZED AS A CERTIFIED COMPONENT.

Signature: _____

Date: _____

Application

A complete application must be sent to:

The National Poultry Improvement Plan
1506 Klondike Road,
Suite 101
USDA-APHIS-VS
Conyers, GA 30094

Denise.L.Brinson@aphis.usda.gov Denise.L.Heard@aphis.usda.gov

with cc to

Elena.L.Behnke@aphis.usda.gov Christina.Lindsey@aphis.usda.gov

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Date Received: _____ Reviewer: _____

Check Here if Approval Granted for Removal of Component:

Check Here if Approval Denied for Removal of Component:

Signature: _____

If Denied, List Reasons:



Appendix F: Auditor Application for USDA-APHIS-VS-NPIP AI Clean and Newcastle Disease Compartment Program



Instructions: Applicants, please complete sections A, B, and C and sign and date application. Applicant must have a qualified sponsor complete Section D and attach a letter of recommendation. Completed application must be submitted to the NPIP. After NPIP review, each applicant will receive notice of approval or denial.

A. Background Information. *To be completed by candidate seeking auditor certification.*

Personal Information

Name of Applicant:	
Business Address (Physical Location with City, State, and Zip):	
Home Address (Physical Location with City, State, and Zip):	
Telephone Number:	
Alternate Phone Number:	
Fax Number:	
Email Address:	

Qualifications

Colleges/Institutions where degrees earned:	
Major (Minor):	
Degrees:	
Veterinary License Number:	
Are you a United States Citizen?	<input type="checkbox"/> Yes <input type="checkbox"/> No
Are you a Federal VMO?	<input type="checkbox"/> Yes <input type="checkbox"/> No
Are you accredited by the NVAP?	<input type="checkbox"/> Yes <input type="checkbox"/> No
Are you a member in good standing with the American College of Poultry Veterinarians?	<input type="checkbox"/> Yes <input type="checkbox"/> No
Estimated years of poultry experience (not including school-related experiences)	<input type="checkbox"/> <1 <input type="checkbox"/> 1-5 <input type="checkbox"/> 5-10 <input type="checkbox"/> >10

B. REASON FOR INTEREST. *To be completed by candidate seeking auditor certification.*

To be eligible for admission into the auditor training workshop course, you must briefly describe your interest in the program and the qualifications you possess that you feel will allow you to become a successful auditor.

C. Affidavit. *To be completed by candidate seeking auditor certification. Please place a check mark by the answer that applies.*

	YES	NO
I certify that I do not own birds of any avian species, whether for meat, eggs, sale, resale, pet, hobby, or otherwise.		
I certify that I have not been employed by, nor do I have spouse, relative, or household member employed by or in contractual relations with any of the companies that belong to the U.S. AI and ND Clean Compartment Program.		
I certify that I will uphold the U.S. veterinarian's oath in all interactions, which states: Being admitted to the profession of veterinary medicine, I solemnly swear to use my scientific knowledge and skills for the benefit of society through the protection of animal health and welfare, the prevention and relief of animal suffering, the conservation of animal resources, the promotion of public health, and the advancement of medical knowledge. I will practice my profession conscientiously, with dignity, and in keeping with the principles of veterinary medical ethics. I accept as a lifelong obligation the continual improvement of my professional knowledge and competence.		
I certify that I have never been convicted of a felony.		
I certify that I have never had my veterinary license revoked by any State board of veterinary medicine.		

Deleted: v

D. Verification. *To be completed by sponsor.*

To be considered as a certified auditor for the training course, a qualified sponsor must write a letter of recommendation to attach with this form, describing the candidate's interest in and dedication to the field of poultry medicine. Qualified sponsors must not be related to the candidate but may be a supervisor, colleague, former professor, or otherwise qualified individual within the field of poultry veterinary medicine.

List relationship to candidate: _____

I have known the candidate for _____ years.

I certify that I have attached to this application a letter of recommendation. Yes No

Name of Sponsor: _____

Signature of Sponsor: _____

Date: _____

CERTIFICATION OF APPLICANT

I DO HEREBY CERTIFY THAT ALL STATEMENTS MADE BY ME IN THIS APPLICATION ARE TRUE AND CORRECT TO THE BEST OF MY KNOWLEDGE, INFORMATION, AND BELIEF; FURTHER, I UNDERSTAND THAT IN THE EVENT I HAVE KNOWINGLY AND WILLFULLY MADE ANY FALSE STATEMENTS, I WILL BE LIABLE FOR PUNISHMENT IN ACCORDANCE WITH ALL APPLICABLE LAWS AND STATUTES. FURTHER, I PLEDGE TO ABIDE BY ALL THE CODES SET FORTH BY EACH COMPANY AND AGREE TO HONOR THE CODE OF CONFIDENTIALITY. I ALSO UNDERSTAND THAT APPROVAL OF THIS APPLICATION WILL ALLOW MY ENROLLMENT IN THE AUDITOR TRAINING COURSE; HOWEVER, I WILL STILL NEED TO SUCCESSFULLY ATTEND AND PASS THE COURSE EXAMINATION TO BECOME A CERTIFIED AUDITOR. ADDITIONALLY, I UNDERSTAND THAT, IF SUCCESSFUL, I WILL NEED TO ENROLL IN CONTINUING EDUCATION EVERY 4 YEARS THEREAFTER.

Signature: _____

Date: _____

Application

A complete application must be sent to:

The National Poultry Improvement Plan
1506 Klondike Road,
Suite 101
USDA-APHIS-VS
Conyers, GA 30094

Denise.L.Brinson@aphis.usda.gov Denise.L.Heard@aphis.usda.gov

with cc to

Elena.L.Behnke@aphis.usda.gov Christina.Lindsey@aphis.usda.gov

For Department Use Only

Date

Received: _____ Reviewer: _____

Approval Granted for Candidate to Attend Workshop:

Approval Denied for Candidate to Attend Workshop:

Anticipated Date of Next Workshop: _____

Signature: _____

If Denied, List Reasons:



Appendix G: NPIP Avian Influenza and Newcastle Disease Compartmentalization Auditor Information and Frequently Asked Questions



Thank you for your interest in the U.S. Avian Influenza and Newcastle Disease Clean Compartment program and for your interest in becoming a certified auditor. Please review the following frequently asked questions.

Q1: Am I qualified to be an auditor?

Certified auditors must meet the following qualifications to be considered for the program:

- Auditors must attend and successfully complete an official USDA-NPIP Auditor Compartment Training Course prior to conducting any audits, and become recertified at least once every 4 years thereafter.
- Auditors must operate and conduct themselves with the highest code of ethics and must not have a conflict of interest with any of the companies which are compartmentalized or seeking compartment certification.
- Auditors must be U.S. licensed and accredited veterinarians who are board certified by the American College of Poultry Veterinarians (ACPV) and meet contract requirements and code of conduct confidentiality standards set forth by APHIS; ~~OR~~ auditors must be Federal Veterinary Medical Officers (VMOs), preferably with poultry experience, who meet the same code of conduct confidentiality standards.

Q2: Do I have to attend a workshop to be a certified auditor?

Yes. Auditors must initially attend and successfully complete an official NPIP Auditor Compartment Training Course, which includes passing a course examination, to become certified to conduct audits. To maintain status, auditors must become recertified by enrolling in the course at least once every 4 years and will be issued a continuing education certificate.

Q3: How do I register for the NPIP Auditor Compartment Training Course?

Auditor applications must be submitted to the NPIP office. Your eligibility will be reviewed, and if you are a successful candidate, you will be invited to register for the next course. Non-Federal VMOs will be responsible for ~~paying the \$500 registration fee and~~ all travel costs associated with the course.

Q4: What is the purpose of the NPIP Auditor Compartment Training Course?

The purpose of the NPIP Auditor Compartment Training Course is threefold:

- To familiarize auditors with the contents of the Management Guidelines and Protocols as well as the official audit checklist of items, and equip them to perform audits accurately and consistently, including conducting mock audits at farm, hatchery, feedmill, egg depot, and office sites.

- To expose auditors to the primary breeder industry and continually educate auditors on pertinent operational activities and important updates in technology within the poultry industry.
- To emphasize the code of ethics in operating as a certified auditor for the U.S. Avian Influenza [and Newcastle Disease](#) Clean Compartment Program.

Q5: What's in it for me?

This opportunity is for any veterinarian who meets the requirements and is hard-working, honest, and willing to learn. As an auditor, you will have the ability to expand your network within the poultry primary breeding industry and develop your skills both as an auditor and as a poultry scientist. Auditors will be given the chance to interact with a very specialized segment of the industry.

Appendix H: Compartmentalization Audit Checklist: Office

	Compartmentalization AUDIT CHECKLIST: Protocols, Procedures, and Requirements			Legend			
OFFICE	<i>Disclaimer: This checklist is to be used at the initial inspection and subsequent reinspection at each individual component within a compartment.</i>			E: Egg Depot F: Farm H: Hatchery M: Feedmill O: Office			
Company Name:	Instructions: Please answer yes/no for each answer. If the item is not applicable, mark NA. Comments are highly useful and should be made when appropriate. Note: Categories marked with triple asterisks (***) are considered part of the AI <u>and ND</u> Clean Programs. Any noncompliances found in one of these categories are considered major, resulting in automatic compartment failure. Noncompliances in categories with no asterisks are considered minor and corrective action should be requested.						
Company Address:							
Date of Inspection:							
Requirements/Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
		Location	Method				
Biosecurity	<i>Biosecurity Training</i>	O	Employees, contract staff, and visitors are trained in biosecurity.				
	<i>Biosecurity Compliance</i>	O	Employees, contract staff, and visitors have a signed compliance agreement.				
	<i>In Periods of High Risk</i>	O	All nonessential visitors and deliveries are prohibited during periods of high risk.				
		O	The company reviews and updates bird hunting activities from employees and contract growers during periods of high risk.				
		O	Employees and contract staff will not attend nonessential regional poultry industry meetings during periods of high risk.				
	<i>Biosecurity Risk Assessment</i>	O	The company has a risk assessment for each component of the compartment (farms, hatchery, feedmill, and egg depot).				
	<i>Biosecurity Vehicular and Equipment Control</i>	O	The company has a written policy for the cleaning, disinfection, and control of all vehicles and equipment before moving into the controlled access and/or the biosecure zone.				

Deleted: [v](#)

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
		Location	Method				
Physical Traits and Diagrams of All Components	<i>Physical Traits</i>	<input type="radio"/>	The company provides general physical traits of all components of the compartment (farms, hatchery, feedmill, and egg depot).				
	<i>Diagrammatic Descriptions</i>	<input type="radio"/>	The company has detailed diagrammatic descriptions for movement of people, vehicles, equipment, birds, and eggs between all components inside and outside the compartment.				
Response Plans	<i>Emergency Response Plan</i>	<input type="radio"/>	The company has a written emergency response plan in conjunction with the ISRCP.				
	<i>In Periods of High Risk</i>	<input type="radio"/>	The company increases communication to company employees, contract growers, and suppliers to promote increased awareness and emphasize essential biosecurity practices during periods of high risk.				
		<input type="radio"/>	Employees increase downtime from last non-compartment bird contact by 24 hours during periods of high risk.				
		<input type="radio"/>	The veterinary and production teams are on high alert and investigate immediately any reported incidents of increased mortality or egg production drops during periods of high risk.				
	<i>Veterinary Health Plan</i>	<input type="radio"/>	The company has a written veterinary health plan.				
	<u>ND Vaccination and Monitoring Program</u>	<u><input type="radio"/></u>	<u>The company has a ND vaccination program.</u>				
		<u><input type="radio"/></u>	<u>The company has a monitoring program for ND vaccinated flocks.</u>				
<u><input type="radio"/></u>		<u>The company has a monitoring program for ND unvaccinated flocks.</u>					

- Deleted:
- Deleted:
- Deleted:
- Deleted:

Appendix I: Compartmentalization Audit Checklist: Farm

FARM	Compartmentalization AUDIT CHECKLIST: Protocols, Procedures, and Requirements				Legend E: Egg Depot F: Farm H: Hatchery M: Feedmill O: Office		
Company Name:		Instructions: Please answer yes/no for each answer. If the item is not applicable, mark NA. Comments are highly useful and should be made when appropriate. Note: Categories marked with triple asterisks (***) are considered part of the AI and ND Clean Programs. Any noncompliances found in one of these categories are considered major, resulting in automatic compartment failure. Noncompliances in categories with no asterisks are considered minor and corrective action should be requested.					
Company Address:							
Date of Inspection:							
Requirements/Procedures	Criteria	Location	Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Physical Requirements		Location	Method				
Perimeter	<i>Livestock Exclusion</i>	F	When livestock are present, they are separated from the controlled access zone by a fence.				
	<i>Vehicular and Personnel Exclusion</i>	F	A biosecure zone barrier surrounds the biosecure zone.				
Unauthorized Entry	<i>Signage</i>	F	Signs indicating unauthorized entry of persons and/or vehicles is prohibited are posted at the entrance to the controlled access zone.				
		F	Signs indicating unauthorized entry of persons and/or vehicles is prohibited are posted at the entrance to the biosecure zone.				
	<i>Barriers</i>	F	Egg holding rooms have barriers in place to prevent unauthorized entry.				

Deleted: [v](#)

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Physical Requirements		Location	Method				
Construction	<i>Wildlife and Pests</i>	F	Buildings are designed and built to deter and prevent entry of wildlife, pests, and companion animals.				
		F	Water systems are designed and built to deter and prevent entry of wildlife and pests.				
	<i>Biosecure Zone</i>	F	Each entrance to the biosecure zone is locked or controlled at all times.				
	<i>Materials</i>	F	Buildings within the biosecure zones are constructed of durable, moisture proof materials and are able to withstand routine cleaning and disinfection.				
Management Procedures							
Biosecurity	<i>Controlled Access Zone Policy</i>	F,O	Authorized personnel and vehicles only enter the controlled access zone after meeting company-established sanitation procedures.				
		F	Dedicated vehicles, if in use, do not leave the controlled access zone.				
		F	Egg holding rooms have procedures in place to prevent unauthorized entry.				
		F	Farm is managed to deter wildlife and pests from the controlled access zone.				
	<i>In Periods of High Risk</i>	F,O	Entrance to the controlled access zone is locked or controlled at all times during periods of high risk.				
	<i>Biosecure Zone Policy</i>	F	Authorized personnel only enter the biosecure zone after meeting company-established sanitation procedures.				
		F	A whole-body shower and change of clothing and footwear occurs before entry into the biosecure zone.				
		F	Before entering the biosecure zone, all vehicles follow company-established cleaning and sanitizing procedures on the interior and exterior of the vehicle.				

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Biosecurity	<i>Biosecure Zone Policy</i>	F	All personnel and visitors entering the biosecure zone must log in.				
		F,O	If the egg storage room is not in the biosecure zone, procedures and barriers are in place to prevent entry from the egg room into the biosecure zone.				
		O	In the case of a multi-age/multi-building biosecure zone, company-established procedures must be documented for transit between buildings.				
		F	Farm is managed to deter wildlife and pests from the biosecure zone.				
	<i>Company Employees and Contract Staff</i>	O	Authorized personnel follow company protocols and procedures and meet all biosecurity requirements for employment or contractual agreement before entry into biosecure zone.				
		O	Company employees (and household members) and contract staff do not own birds.				
		O	Company employees and contract staff avoid contact with birds outside the compartment and/or comply with company policies related to downtime and quarantine.				
		O	Company employees and contract staff receive annual biosecurity training.				
		O	Company employees and contract staff agree to follow company-established policy regarding personal items and food.				
		O	<u>Farm company employees, farm contract staff and farm visitors are trained in company-established biosecurity procedures.</u>				
	<i>Visitors</i>	O	All visitors meet a minimum 24-hour downtime from contact with non-compartment birds (including shower and change of clothing) or as specified by company-established visitor requirements.				
O		All visitors agree to comply with company-established biosecurity procedures.					

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Biosecurity	Visitors	O	All visitors sign a declaration stating date of last bird contact.				
		F	All visitors agree to follow company-established policy regarding personal items and food.				
	In Periods of High Risk	F,O	All non-essential visitors and deliveries are prohibited during periods of high risk.				
Transportation	Supplies and Equipment	F,O	Supplies and goods coming into the biosecure zone are from approved suppliers only and have undergone company-established cleaning and disinfection procedures.				
		O	Bedding materials coming into the biosecure zone are obtained only from company-approved suppliers.				
		O	Tools and equipment coming into the biosecure zone undergo company-approved cleaning and disinfection procedures.				
	In Periods of High Risk	F,O	Only essential tools and equipment that have undergone enhanced company-approved cleaning and disinfection procedures move into the biosecure zone during periods of high risk.				
	All Hatching Egg/Chick/ Poult and Bird Movement	F, O, H, E	Company-established sanitation and biosecurity procedures apply for vehicles, equipment, authorized egg pickup personnel, and personnel involved in moving flocks.				
		F, O, H, E	Hatching eggs are sanitized with an approved disinfectant at the farm, hatchery or egg depot.				
		O	Egg/chick/poult transport personnel agree to meet company-established biosecurity procedures for delivery.				
		O	Drivers wear company-provided clothing and footwear.				
		O	Records tracing the origin and production dates of all hatching eggs/chicks/poults are kept.				

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Transportation	Bird Movement Within Compartment	O	***Flocks test negative to AI within 21 days prior to movement.				
		<u>O</u>	***Birds must originate from flocks that were vaccinated for ND using USDA licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If unvaccinated flocks, they must test negative to ND.				Deleted: <u>v</u>
		O	***Day-old chicks/poults are derived from a NPIP AI Clean source flock.				Deleted: <u>v</u>
	In Periods of High Risk	O	***Flocks test negative to AI and ND via RT-PCR 48 hours prior to movement during periods of high risk.				Deleted: <u>v</u>
	Bird Movement Into the Compartment	O	***Source flocks participate in NPIP AI Clean or equivalent program.				
		O	***Pullets, cockerels, and adult birds originating outside the compartment have tested negative to AI (30 samples per flock via serology and 15 samples per flock via antigen detection) within 21 days of shipment.				
		<u>O</u>	***Birds must originate from flocks that were vaccinated for ND using USDA licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If unvaccinated flocks, they must test negative to ND.				Deleted: <u>v</u>
		O	***Flocks are inspected by an official veterinarian or designated individual within 30 days prior to movement.				Deleted: <u>v</u>

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Transportation	In Periods of High Risk	O	***Pullets, cockerels, and adult birds originating outside the compartment have 30 samples tested negative to <u>AI and ND</u> via RT-PCR 48 hours prior to shipment during periods of high risk.				
		O	***Birds must originate from flocks that were vaccinated for ND using USDA licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If unvaccinated flocks, they must test negative to ND.				
		O	Driving routes for all authorized transport and service vehicles are reviewed and alternate routes are predetermined to avoid any areas with other poultry or migratory birds that could present a potential risk during periods of high risk.				
		F,O	Vehicular traffic is reduced to only critical components when necessary during periods of high risk.				
		F,O	The vehicle non-essential parking perimeter is increased during periods of high risk.				
		O	Vehicles undergo enhanced cleaning and disinfection during periods of high risk.				

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Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Husbandry	<i>Wildlife and Pests</i>	O	Pest control procedures are documented and recorded.				
	<i>Insect Control</i>	O	Insect control procedures are documented and recorded.				
	<i>Vegetation Control</i>	O	Vegetation control procedures are documented and recorded.				
	<i>Water</i>	O	If surface water is present, company has procedures in place to deter waterfowl and wild birds.				
		F	Surface water is not used for drinking water or for any purpose.				
		F	Only treated well water or municipal water is used at the farm.				
	<i>Feed</i>	F,O	Feed spills are removed following company-established procedures.				
	<i>Mortality and Biologic Waste Disposal</i>	F,O	Daily mortality is disposed of according to company's biosecurity plan and local environmental regulations.				
		F,O	Biologic waste and cull eggs are disposed of according to the company's biosecurity plan and in compliance with local environmental regulations.				
	<i>Bedding Materials</i>	F	Bedding materials are stored to prevent access from pests and wild birds.				
F,O		Bedding material restocking is allowed only after a trained company employee ensures that cleaning and disinfection of the facility has been performed according to company standards.					
O		The company has biosecurity and sanitation procedures for removal of litter.					

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Husbandry	Flock Depletion and House Sanitation	O	Flocks to be depleted test AI negative within 21 days prior to movement.				
		O	Flocks are inspected by an accredited veterinarian within 21 days prior to flock depletion.				
		O	The company has biosecurity and EPA-approved sanitation procedures for depletion of flocks.				
		O	The company has biosecurity and sanitation procedures for house cleaning and disinfection after flock depletion.				
	In Periods of High Risk	O	Premises where birds are present must test negative for AI and ND via RT-PCR 48 hours prior to movement of litter or manure during periods of high risk.				
Bird Health	Veterinary Health Plan	F,O	Daily production and mortality records are kept according to company-established policy.				
		F,O	The company has a veterinary health plan which outlines procedures to investigate unexplained increases in mortality according to company-established procedures.				

Appendix J: Compartmentalization Audit Checklist: Feedmill

FEEDMILL	Compartmentalization AUDIT CHECKLIST: Protocols, Procedures, and Requirements				Legend E: Egg Depot F: Farm H: Hatchery M: Feedmill O: Office		
Company Name:		<p style="font-size: small; margin: 0;">Disclaimer: This checklist is to be used at the initial inspection and subsequent reinspection at each individual component within a compartment.</p> <p style="font-size: small; margin: 0;">Instructions: Please answer yes/no for each answer. If the item is not applicable, mark NA. Comments are highly useful and should be made when appropriate.</p> <p style="font-size: small; margin: 0;">Note: Categories marked with triple asterisks (***) are considered part of the AI and ND Clean Programs. Any noncompliances found in one of these categories are considered major, resulting in automatic compartment failure. Noncompliances in categories with no asterisks are considered minor and corrective action should be requested.</p>					
Company Address:							
Date of Inspection:							
Requirements/Procedures	Criteria	Location	Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Physical Requirements		Location	Method				
Perimeter	<i>Livestock Exclusion</i>	M	When livestock are present, they are separated from the feedmill by a fence.				
Unauthorized Entry	<i>Signage</i>	M	Signs indicating unauthorized entry of persons and/or vehicles is prohibited are posted at the entrance to the controlled access zone.				
	<i>Barriers</i>	M	The feedmill has a gate at its entrance to the controlled access zone.				
Construction	<i>Wildlife and Pests</i>	M	Buildings are designed and built to deter and prevent entry of wildlife, pests, and companion animals.				
		M	Water systems are designed and built to deter and prevent entry of wildlife and pests.				
	<i>Materials</i>	M	Feedmill is constructed of durable, moistureproof materials that are able to withstand routine cleaning and disinfection.				

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Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Biosecurity	<i>Controlled Access Zone Policy</i>	M,O	Authorized personnel and vehicles only enter the controlled access zone after meeting company established sanitation procedures.				
		M	All personnel and visitors entering the controlled access zone must log in.				
	<i>Company Employees and Contract Staff</i>	O	Authorized personnel follow company-established protocols and procedures and meet all biosecurity requirements for employment or contractual agreement before entry into feedmill.				
		O	Company employees (and household members) and contract staff do not own birds.				
		O	Company employees and contract staff avoid contact with birds outside the compartment and/or comply with company policies related to downtime and quarantine.				
		O	Company employees and contract staff receive annual biosecurity training.				
		O	Company employees and contract staff agree to follow company-established policy regarding personal items and food.				
		<u>O</u>	<u>Feedmill company employees, feedmill contract staff and feedmill visitors are trained in company-established biosecurity procedures.</u>				
		<i>Visitors</i>	O	All visitors agree to follow company-established policy regarding personal items and food.			
	O		All visitors sign a declaration stating date of last bird contact.				
	O		All visitors follow company-established biosecurity procedures.				
	<i>In Periods of High Risk</i>	O	All non-essential visitors and deliveries are prohibited during periods of high risk.				
		O	Driving routes for all authorized transport and service vehicles are reviewed and alternate routes are predetermined to avoid any areas with other poultry or migratory birds that could present a potential risk during periods of high risk.				

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Biosecurity	<i>In Periods of High Risk</i>	M,O	Vehicular traffic is reduced to only critical components when necessary during periods of high risk.				
		M,O	The vehicle non-essential parking perimeter is increased during periods of high risk.				
		O	Vehicles undergo enhanced cleaning and disinfection during periods of high risk.				
Husbandry	<i>Wildlife and Pests</i>	O	Wildlife and pest control procedures are documented and recorded.				
	<i>Insect Control</i>	O	Insect control procedures are documented and recorded.				
	<i>Vegetation Control</i>	O	Vegetation control procedures are documented and recorded.				
	<i>Cleaning and Disinfection</i>	O	Feedmill has company-established protocols for cleaning and disinfection.				
	<i>Water</i>	O	If surface water is present, company has procedures in place to deter waterfowl and wild birds.				
		O	Surface water is not used at the feedmill for any purpose.				
O		Only treated well water or municipal water is used at the feedmill.					
Finished Feed and Delivery	<i>Finished Feed</i>	M,O	Finished feed undergoes established treatment procedures prior to storage and distribution.				
		M,O	The company has established procedures for separating raw feed ingredients from finished feed.				
		M,O	Feed ingredients and/or finished feed spills are removed following company-established procedures.				
		M,O	The company has established procedures for feed delivery.				
		O	The company has established procedures and documentation for compartment and non-compartment feed delivery.				
	<i>Delivery Vehicles</i>	O	The company has established EPA-approved sanitation and biosecurity procedures for feed delivery vehicles.				

Appendix K: Compartmentalization Audit Checklist: Hatchery

	Compartmentalization AUDIT CHECKLIST: Protocols, Procedures, and Requirements				Legend E: Egg Depot F: Farm H: Hatchery M: Feedmill O: Office		
HATCHERY	<i>Disclaimer: This checklist is to be used at the initial inspection and subsequent reinspection at each individual component within a compartment.</i>						
Company Name:	Instructions: Please answer yes/no for each answer. If the item is not applicable, mark NA. Comments are highly useful and should be made when appropriate. Note: Categories marked with triple asterisks (***) are considered part of the AI and ND Clean Programs . Any noncompliances found in one of these categories are considered major, resulting in automatic compartment failure. Noncompliances in categories with no asterisks are considered minor and corrective action should be requested.						
Company Address:							
Date of Inspection:							
Requirements/Procedures	Criteria	Location	Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Physical Requirements		Location	Method				
Perimeter	<i>Livestock Exclusion</i>	H	When livestock are present, they are separated from the hatchery by a fence.				
	<i>Vehicular and Personnel Exclusion</i>	H	A biosecure zone barrier surrounds the biosecure zone.				
Unauthorized Entry	<i>Signage</i>	H	Signs indicating unauthorized entry of persons and/or vehicles is prohibited are posted at the entrance to the controlled access zone.				
		H	Signs indicating unauthorized entry of persons and/or vehicles is prohibited are posted at the entrance to the biosecure zone.				
	<i>Barriers</i>	H	Egg holding rooms have barriers in place to prevent unauthorized entry.				
		H	Receiving/shipment dock is an enclosed area that undergoes company-established cleaning and disinfection procedures.				

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Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Physical Requirements		Location	Method				
Construction	<i>Wildlife and Pests</i>	H	Buildings and water systems are designed to deter and prevent entry of wildlife, pests, and companion animals.				
	<i>Biosecure Zone</i>	H	Each entrance to the biosecure zone is locked or controlled at all times.				
		H	Egg holding, setting, hatching, chick/poult processing, chick/poult transfer, and vaccine rooms are part of the biosecure zone.				
		H	Barriers are in place to prevent entry from the egg room into the biosecure zone.				
	<i>Controlled Access Zone</i>	H	Hatchery office, egg and chick/poult loading dock can be a part of the controlled access zone.				
	<i>Materials</i>	H	The biosecure zones of the hatchery are constructed of durable, moistureproof materials that are able to withstand routine cleaning and disinfection.				
		H	Hatchery is designed and built to deter and prevent entry of wildlife and pests.				
		H	Egg holding rooms are constructed of durable materials to exclude wildlife and pests.				
Management Procedures							
Biosecurity	<i>Controlled Access Zone Policy</i>	H,O	Authorized personnel and vehicles only enter the controlled access zone after meeting company-established sanitation procedures.				
	<i>Biosecure Zone Policy</i>	H,O	Authorized personnel only enter the biosecure zone after meeting company-established sanitation procedures.				
		H,O	Authorized vehicles only enter the biosecure zone after meeting company established cleaning and sanitizing procedures for the interior and exterior of the vehicle.				
		H,O	A whole-body shower and change of clothing and footwear occurs before entry into the biosecure zone.				

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Biosecurity	<i>Biosecure Zone Policy</i>	H	All personnel and visitors entering the biosecure zone must log in.				
		H,O	Procedures are in place to prevent entry from the egg room into the biosecure zone.				
	<i>Company Employees and Contract Staff</i>	O	Authorized personnel follow company protocols and procedures and meet all biosecurity requirements for employment or contractual agreement before entry into biosecure zone.				
		O	Company employees (and household members) and contract staff do not own birds.				
		O	Company employees and contract staff avoid contact with birds outside the compartment and/or comply with company policies related to downtime and quarantine.				
		O	Company employees and contract staff receive annual biosecurity training.				
		O	Company employees and contract staff agree to follow company-established policy regarding personal items and food.				
		O	Drivers wear company-approved clothing and footwear.				
		<u>O</u>	<u>Hatchery company employees, hatchery contract staff and hatchery visitors are trained in company-established biosecurity procedures.</u>				
		<i>Visitors</i>	O	All visitors meet a minimum 24-hour downtime from contact with non-compartment birds (including shower and change of clothes or as specified by company-established visitor requirements).			
	O		All visitors sign a declaration stating date of last bird contact.				
	O		All visitors follow company-established biosecurity procedures.				
	O		All visitors agree to follow company-established policy regarding personal items and food.				
	<i>In Periods of High Risk</i>	O	All non-essential visitors and deliveries are prohibited during periods of high risk.				

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Transportation	<i>Supplies and Equipment</i>	O	Supplies and goods coming into the biosecure zone undergo approved, company-established cleaning and disinfection procedures.				
		H,O	Tools and equipment undergo company-approved cleaning and disinfection procedures.				
		H,O	Reusable equipment that returns to the hatchery is cleaned and disinfected.				
	<i>In Periods of High Risk</i>	H,O	Only essential tools and equipment that have undergone enhanced company-approved cleaning and disinfection procedures move into the biosecure zone during periods of high risk.				
	<i>All Hatching Egg/Chick /Poult Movement</i>	H,O,F,E	Hatching eggs are sanitized with an approved disinfectant at the farm, hatchery, or egg depot.				
		H,O,F,E	Company-established sanitation and biosecurity procedures apply for vehicles, equipment, and personnel involved in moving hatching eggs and chicks/poults.				
		H,O,F,E	Records tracing the origin and production dates of all hatching eggs and day-old chicks/poults are maintained.				
	<i>Hatching Egg Movement Into the Compartment</i>	O	***Hatching eggs are derived from a source flock in which 30 samples have tested negative to AI using an approved NPIP assay within 21 days of the shipment.				
		<u>O</u>	***Hatching eggs are derived from a source flock that was vaccinated for ND using USDA licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If unvaccinated, source flocks must test negative to ND.				
		O	***Source flocks participate in NPIP AI Clean or equivalent program.				
<i>Hatching Egg Movement Within the Compartment</i>	O	***Hatching eggs that are moved between premises within the compartment are derived from source flocks that participate in the NPIP AI Clean or equivalent program.					
	<u>O</u>	***Hatching eggs must originate from flocks that were vaccinated for ND using USDA licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If unvaccinated, source flocks must test negative to ND.					

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Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Transportation	Day-Old Chick/Poult Movement Within the Compartment	O	***Source flocks participate in NPIP AI Clean or equivalent program.				
		O	***Birds must originate from flocks that were vaccinated for ND using USDA licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If unvaccinated, source flocks must test negative to ND.				
	Hatching Egg and Day-Old Chick/Poult Movement Outside the Compartment	O	The company has established biosecurity procedures for all equipment that returns to the hatchery from outside the compartment to be cleaned and disinfected.				
		In Periods of High Risk	O	Driving routes for all authorized transport and service vehicles are reviewed and alternate routes are predetermined to avoid any areas with other poultry or migratory birds that could present a potential risk during periods of high risk.			
			H,O	Vehicular traffic is reduced to only critical components when necessary during periods of high risk.			
			H,O	The vehicle nonessential parking perimeter is increased during periods of high risk.			
			O	Vehicles undergo enhanced cleaning and disinfection during periods of high risk.			

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Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Husbandry	<i>Wildlife and Pests</i>	O	Wildlife and pest control procedures are documented and recorded.				
	<i>Insect Control</i>	O	Insect control procedures are documented and recorded.				
	<i>Vegetation Control</i>	O	Vegetation control procedures are documented and recorded.				
	<i>Cleaning and Disinfection</i>	H,O	Hatchery has company-established protocols for cleaning and disinfection.				
	<i>Water</i>	O	Surface water is not used in the hatchery for any purpose.				
		O	Only treated well water or municipal water is used in the hatchery.				
<i>Waste Removal</i>	H	All biologic waste, hatchery residue, and cull eggs are disposed according to company biosecurity plan and in compliance with local environmental regulations.					

Appendix L: Compartmentalization Audit Checklist: Egg Depot

	Compartmentalization AUDIT CHECKLIST: Protocols, Procedures, and Requirements			Legend E: Egg Depot F: Farm H: Hatchery M: Feedmill O: Office			
EGG DEPOT	<i>Disclaimer: This checklist is to be used at the initial inspection and subsequent reinspection at each individual component within a compartment.</i>						
Company Name:	Instructions: Please answer yes/no for each answer. If the item is not applicable, mark NA. Comments are highly useful and should be made when appropriate. Note: Categories marked with triple asterisks (***) are considered part of the AI <u>and ND</u> Clean Programs. Any noncompliances found in one of these categories are considered major, resulting in automatic compartment failure. Noncompliances in categories with no asterisks are considered minor and corrective action should be requested.						
Company Address:							
Date of Inspection:							
Requirements/Procedures	Criteria	Location	Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Physical Requirements		Location	Method				
Perimeter	<i>Livestock Exclusion</i>	E	When livestock are present, they are separated from the egg depot by a fence.				
Unauthorized Entry	<i>Signage</i>	E	Signs indicating unauthorized entry of persons and/or vehicles is prohibited are posted at the entrance to the controlled access zone.				
	<i>Barriers</i>	E	Egg depot has a gate at its entrance of the controlled access zone.				
E		Egg holding rooms have barriers in place to prevent unauthorized entry.					
Construction	<i>Wildlife and Pests</i>	E	Egg depot is designed and built to deter and prevent entry of wildlife, pests, and companion animals.				
		E	Egg holding rooms are constructed of durable materials to exclude wildlife and pests.				
		E	Egg depot is constructed of durable, moistureproof materials that are able to withstand routine cleaning and disinfection.				
	<i>Materials</i>	E	Egg receiving/shipment dock is should be an enclosed area that is part of the controlled access zone.				

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Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action	
Management Procedures		Location	Method					
Biosecurity	<i>Controlled Access Zone Policy</i>	E,O	Authorized personnel and vehicles enter the controlled access zone after meeting company established sanitation procedures.					
		E	All personnel and visitors entering the egg depot must log in.					
		E	Egg depot has procedures in place to prevent entry from the egg receiving area into the egg depot.					
	<i>Company Employees and Contract Staff</i>	O	Authorized personnel follow company protocols and procedures and meet all biosecurity requirements for employment or contractual agreement before entry into the egg depot.					
		O	Company employees (and household members) and contract staff do not own birds.					
		O	Company employees and contract staff agree to avoid contact with birds outside the compartment and/or agree to comply with company policies related to downtime and quarantine.					
		O	Company employees and contract staff receive annual documented biosecurity training.					
		O	Drivers agree to wear company-provided clothing and footwear.					
		O	Company employees and contract staff agree to follow company-established policy regarding personal items and food.					
		<u>O</u>	<u>Egg depot company employees, egg depot contract staff and egg depot visitors are trained in company-established biosecurity procedures.</u>					
	<i>Visitors</i>	O	All visitors meet a minimum 24-hour downtime from contact with non-compartment birds (including shower and change of clothes) or as specified by company-established visitor requirements.					
		O	All visitors follow company-established biosecurity procedures.					
		O	All visitors sign a declaration stating date of last bird contact.					
		O	All visitors agree to follow company-established protocols regarding personal items and food.					

Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Biosecurity	<i>In Periods of High Risk</i>	O	All non-essential visitors and deliveries are prohibited during periods of high risk.				
	<i>All Hatching Egg Movement</i>	E,O,H,F	Hatching eggs are sanitized with an approved disinfectant at the farm, hatchery, or egg depot.				
		O	Records tracing the origin and production dates of all hatching eggs are kept.				
		E,O,H,F	Company-established sanitation and biosecurity procedures apply for vehicles, equipment, and personnel involved in moving hatching eggs.				
	E,O	Reusable equipment that returns to the egg depot is cleaned and disinfected.					
Transportation	<i>Hatching Egg Movement Into the Compartment</i>	O	***Source flocks participate in NPIP AI Clean or equivalent program.				
		<u>O</u>	***Hatching eggs must originate from flocks that were vaccinated for ND using USDA licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If unvaccinated, source flocks must test negative to ND.				Deleted: <u>Y</u>
		O	***Hatching eggs are derived from a source flock in which 30 samples have tested negative to AI using an approved NPIP assay within 21 days of the shipment.				
		O	Records which trace the origin and production dates of all hatching eggs are kept.				
	<i>Hatching Egg Movement Within the Compartment</i>	O	The company has established biosecurity procedures for vehicles, equipment, and personnel transporting hatching eggs.				
		O	***Source flocks participate in NPIP AI Clean or equivalent program.				
		<u>O</u>	***Hatching eggs must originate from flocks that were vaccinated for ND using USDA licensed vaccines and compliant with a program to evaluate serological response to ND vaccination. If unvaccinated, source flocks must test negative to ND.				Deleted: <u>Y</u>
	O	Egg receiving/shipment dock undergoes routine company-established cleaning and disinfection procedures.					

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Requirements/ Procedures	Criteria		Verification Method	Records (Protocols and/or Training) (Y/N/NA)	Training (Y/N/NA)	Compliance (Y/N/NA)	Additional Comments / Recommended Corrective Action
Management Procedures		Location	Method				
Transportation	<i>Hatching Egg Movement Out of the Compartment</i>	O	The company has established biosecurity procedures for all equipment that returns to the egg depot from outside the compartment to be cleaned and disinfected.				
	<i>In Periods of High Risk</i>	O	Driving routes for all authorized transport and service vehicles are reviewed and alternate routes are predetermined to avoid any areas with other poultry or migratory birds that could present a potential risk during periods of high risk.				
		E,O	Vehicular traffic is reduced to only critical components when necessary during periods of high risk.				
		E,O	The vehicle non-essential parking perimeter is increased during periods of high risk.				
		O	Vehicles undergo enhanced cleaning and disinfection during periods of high risk.				
Husbandry	<i>Wildlife and Pests</i>	O	Wildlife and pest control procedures are documented and recorded.				
	<i>Insect Control</i>	O	Insect control procedures are documented and recorded.				
	<i>Vegetation Control</i>	O	Vegetation control procedures are documented and recorded.				
	<i>Water</i>	E,O	Surface water is not used in the egg depot for any purpose.				
		E,O	Only treated water or municipal water is used in the egg depot.				
	<i>Cleaning and Disinfection</i>	E,O	Egg depot has company established protocols for cleaning and disinfection.				
<i>Waste Removal</i>	E,O	All biologic waste, egg depot residue, and cull eggs are disposed according to company biosecurity plan and in compliance with local environmental regulations.					

Delegates: 145 D, G, H

Program Standard F: Compartmentalization for Protection against Avian Influenza in Primary Poultry Breeding Companies in the United States of America. ***NOTE: Each Compartmentalization proposal is color-coded in the text and will receive a separate vote***

Program Standards - Proposal No. 13

Reason: The basis for these proposals is for the addition of Newcastle Disease (ND), caused by virulent APMV-1, to the Compartmentalization for Protection against Avian Influenza in Primary Poultry Breeding Companies in the United States of America program. Though the United States are free of ND, this addition is intended to ensure trading partners that the primary breeding-hatchery industry conducts a program for the prevention, detection and control of Newcastle Disease. As well, a compartment that includes AI and ND aligns with compartment programs established in other countries. Through routine surveillance of each participating breeding flock, the presence of virulent APMV-1 in primary breeding poultry flocks is being surveilled. As well, additional considerations and testing parameters are outlined in the event of ND and a heightened risk period.

Sponsor: Dr. Kate Hayes, Aviagen

Program Standards - Proposal No. 14

Reason: To have Egg Depot audit checklist criteria be consistent and match the wording on page 27 of Subpart F-Compartmentalization which states that "Physical Requirements: Egg receiving/shipment dock should be an enclosed area."

Sponsor: Melissa Phillips, Cobb-Vantress, Inc.

Program Standards – Proposal No. 15

Reason: In the original checklist, certain categories were open to interpretation for completion, and being required to provide proof in the form of a protocol or training did not make sense for each verification method. Some of the protocol boxes for those categories which are not applicable in any circumstance are now shaded so that a response Y, N or NA will no longer be required. Additionally, the protocol column is expanded for clarification, and the training column is now replaced with a single verification method in each section regarding biosecurity.

Sponsor: Dr. Elena Behnke, Aviagen, Inc.

Program Standards - Proposal No. 16

Reason: To correct typos, align the avian influenza language with proper OIE usage, correct e-mail addresses, and eliminate the \$500 fee for the compartment auditor training course.

Sponsor: Dr. Christina Lindsey, NPIP Compartmentalization Coordinator